Evaluation of $^{11}$C-BU99008, a Positron Emission Tomography ligand for the Imidazoline$_2$ binding sites in Rhesus brain

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† These two authors contributed equally to this work.

RUNNING TITLE: Evaluation of $^{11}$C-BU99008 in Rhesus

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Abstract
The development of a PET radioligand selective for I₂-imidazoline binding sites (I₂BS) would enable, for the first time, specific, measurable in vivo imaging of this target protein, along with assessment of alterations in expression patterns of this protein in disease pathophysiology.

Methods: BU99008 was identified as the most promising I₂BS radioligand candidate, and radiolabelled with carbon-11 via methylation. The in vivo binding properties of ¹¹C-BU99008 were assessed in rhesus monkeys in order to determine: brain penetration, brain distribution, binding specificity and selectivity (via the use of the unlabelled blockers), and the most appropriate kinetic model for analysing data generated with this PET radioligand.

Results: ¹¹C-BU99008 was demonstrated to readily enter to the brain resulting in a heterogeneous distribution (globus pallidus>cortical regions>cerebellum) consistent with the reported regional I₂BS densities as determined by human tissue section autoradiography and preclinical in vivo PET studies in the pig. In vivo competition studies revealed ¹¹C-BU99008 displayed reversible kinetics specific for the I₂BS. The MA1 multi-linear model was the most appropriate analysis method for this PET radioligand in this species. The selective, I₂BS blocker, BU224, was shown to cause a saturable, dose-dependent decrease in ¹¹C-BU99008 binding in all regions of the brain assessed, further demonstrating the heterogeneous distribution of I₂BS protein in the rhesus brain and binding specificity for this radioligand.

Conclusion: These data demonstrate ¹¹C-BU99008 represents a specific and selective PET radioligand for imaging and quantifying the I₂BS, in vivo, in rhesus monkey. Further work is underway to translate the use of ¹¹C-BU99008 to the clinic.

Keywords: Imidazoline₂ binding site, I₂BS, positron emission tomography, PET, BU99008
Introduction
The ability of the α₂-adrenoceptor agonist clonidine and the antagonist idazoxan, to label a sub-population of binding sites, not displaceable by the endogenous ligand noradrenaline, led to the discovery of the imidazoline binding sites some 20 years ago. These binding sites have subsequently been divided into three groups: the imidazoline₁ binding site that is preferentially labelled by ³H-clonidine, the imidazoline₂ binding site (I₂BS) that is preferentially labelled by ³H-idazoxan, and the imidazoline₃ binding site which is an atypical imidazoline site found on pancreatic β-cells (for review see (1)).

I₂BS are known to reside on the mitochondrial membranes of astrocytes (2). Changes in post-mortem binding density of the I₂BS has implicated them in a range of psychiatric conditions such as depression and addiction, along with neurodegenerative disorders such as Alzheimer’s disease and Huntington’s chorea (3). Functional interactions in preclinical models have also been shown in relation to the opioid system, where I₂BS ligands have been shown to affect tolerance to morphine (4) and alleviate some of the morphine withdrawal syndrome in rats (5). I₂BS ligands have also been shown acutely to affect feeding and appetite by an as yet undetermined mechanism (6). The location of I₂BS on glial cells and the possibility that they may in some way regulate glial fibrillary acidic protein (7) have led to increased interest into the role of I₂BS and I₂BS ligands in conditions characterised by marked gliosis. The density of I₂BS has been shown to increase in Alzheimer’s disease post mortem (3), and it has also been suggested that I₂BS may be a marker for human glioblastomas (8). Subsequent publications added weight to this argument showing that the density of I₂BS is increased in vivo with heat-induced gliosis (9). Additionally, Callado and coworkers have shown not only an increase in the I₂BS in human gliomas but this increase in binding sites was correlated with the severity and malignancy of the glioma (10).

Positron emission tomography (PET) is an in vivo imaging technique that uses radioligands as selective molecular probes to map the location and density of specific proteins. The development of a selective I₂BS PET radioligand would
allow for the characterisation of I$_2$BS \textit{in vivo} and their regulation in disease states. A number of ligands selective for I$_2$BS have been reported, but only two potential PET radioligands have been reported to date: the radiosynthesis of $^{11}$C-benzazoline, but its study \textit{in vivo} has not been reported \cite{11} and radiosynthesis and \textit{in vivo} imaging evaluation of $^{11}$C-FTIMD in non-human primate \cite{12} although the specific binding signal appears to be low for this radioligand.

We have recently reported the synthesis, \textit{in vitro} and \textit{in vivo} evaluation and radiosynthesis of a PET radioligand for the I$_2$BS, $^{11}$C-BU99008 \cite{13, 14}. In this manuscript, we report the preclinical \textit{in vivo} evaluation of $^{11}$C-BU99008, for imaging I$_2$BS, in rhesus monkey brain.
**Materials and Methods**

**Chemicals**

\(^3\)H-BU99008 (SA=1.04TBq/mmol) was custom synthesised by Sibtech. Challenge drugs moclobemide and lazabemide were obtained from commercial suppliers: Sigma-Aldrich Company Ltd. and Tocris Biosciences. Dr Husbands synthesized the BU224. All other chemicals and reagents were purchased from commercial suppliers and used without further purification.

**Animals**

All experiments in animals were carried out in accordance with the UK Animals (Scientific Procedures) Act 1986. PET imaging experiments in rhesus monkeys were conducted in accordance with a protocol approved by the Yale University Institutional Animal Care and Use Committee.

**In Vitro Competition Binding Studies**

Membrane preparation, competition binding studies and data analysis were conducted as previously described (13) with the following alterations. Rat (male, Wistar, 250-300g) and Cynomologus monkey brains were used and resulting membrane preparations stored at -80°C. The displacement binding studies for the \(^3\)H-BU99008 (1nM) were conducted at 37°C in assay buffer (50mM TRIS-HCl, 140mM NaCl, 1.5mM MgCl\(_2\), 5mM KCl, 1.5mM CaCl\(_2\), pH7.4), and radioactivity was determined using a Tricarb 2900 β-counter. Protein content was determined using a Pierce bicinchoninic acid kit.

**PET Imaging Studies in Rhesus Monkeys**

*Radiochemistry.* \(^11\)C-BU99008 was prepared by \(N\)-alkylation of the desmethyl precursor BU99007 with \(^11\)C-CH\(_3\)I in the AutoLoop synthesis module (Bioscan). A description of the synthetic methods can be found in supplementary information (Supplementary Figure 1 (supplemental materials are available online only at http://jnm.snmjournals.org)).
Study Design. Two Rhesus monkeys (Macaca mulatta; female, ~6kg, 7 years and female, ~7kg, 8 years) were used, with scanning days at least 14 days apart. Each animal had five scanning days. Each scanning day consisted of a baseline scan with $^{11}$C-BU99008 (120min); following this, animals received an i.v. injection of blocking drug over a 10min period, ~10min prior to initiation of a second scan with $^{11}$C-BU99008 (120min), to determine binding specificity and selectivity of the radioligand (Supplementary Table 1). Administration of the specific I$_2$BS ligand, BU224, was: monkey one, 0.01, 0.03 and 0.3mg/kg, and monkey two, 0.01, 0.03, 0.1mg/kg. To assess the selectivity of binding to I$_2$BS, both animals received an injection of the reversible MAO$_A$ inhibitor, moclobemide (1mg/kg) and the reversible MAO$_B$ inhibitor, lazabemide (0.5mg/kg). Data acquisition started simultaneously with ligand injection. Vital signs were monitored at least four times per hour, and more frequently following injection of tracer and blocking drugs.

MRI Imaging. Images were acquired for each monkey on a Siemens 3.0-T Trio scanner, using an extremity coil. $T_1$-weighted images were acquired in the coronal plane with a spin-echo sequence (TE=3.34, TR=2530, flip angle=7°, section thickness=0.50mm, field of view=140mm, image matrix=256×256×176 pixels, matrix size=0.547×0.547×0.500mm). Whole brain image was cropped to 176×176×176 pixels using MEDx software (Medical Numerics) before co-registration with PET image data.

PET Imaging Procedures. Animals were sedated with an intramuscular injection of ketamine hydrochloride (10±2mg/kg), ~2h prior to the start of scanning, transported to the PET facility, anaesthetized using isofluorane, intubated and maintained on oxygen and 1.5–2.5% isofluorane throughout the study. PET scans were performed on the FOCUS 220 PET scanner (Siemens Preclinical Solutions), with a reconstructed image resolution of ~1.5mm. Following a transmission scan, 170±14MBq (4.6±0.4mCi; mass dose=0.08±0.02µg/kg) of $^{11}$C-BU99008 was injected over 3min. List-mode data were acquired for 120min and binned into sinograms with the following frame timing: 6×30s; 3×1min; 2×2min; 22×5min.
Dynamic scan data were reconstructed with a filtered-back projection algorithm with corrections for attenuation, normalization, scatter, and randoms.

**Arterial Blood Sampling.** Arterial blood samples were collected for the determination of: whole blood and plasma input functions, and metabolite analysis and plasma free fraction ($f_p$) of $^{11}$C-BU99008. These procedures are described in detail in the supplementary information.

**Regional Time-Activity Curve Computation.** An existing region of interest (ROI) map defined on a template brain (a representative MR image of a rhesus brain) was used. The following, a priori defined, ROI were examined: cingulate, frontal, insula, and occipital cortex, brainstem, pons, cerebellum, caudate, putamen, globus pallidus and thalamus. A nonlinear transformation was estimated using the Bioimagesuite software (http://www.bioimagesuite.org/) to transfer the ROI template to the MR image of each animal used during this study. These regions were then transferred to the PET images based on a rigid transformation matrix (15), and used to generate time-radioactivity curves (TAC).

**Kinetic Modeling.** Regional TACs were analysed using one- and two-tissue compartment models (1TC and 2TC) and multi-linear analysis (MA1) (16) to calculate regional distribution volume ($V_T$). MA1 is a linear method related to Logan analysis, but with less noise-induced bias. Like Logan analysis, data are fitted starting at a specified time, $t^*$; here $t^*$=20min. The optimal model was based on quality of fit, and the uncertainty (standard error) of the $V_T$ parameter estimate. For blocking studies, since there was no suitable reference region, a graphical method was used to calculate the non-displaceable volume of distribution ($V_{ND}$) and global and regional receptor occupancy (17). Receptor occupancy data from different blocking drug doses was fitted to a one-site binding $E_{max}$ model where occupancy values ($Occ$) are plotted versus administered dose ($d$); where $Occ = d/(d + ED_{50})$, in order to derive the blocking dose needed to induce 50% global receptor occupancy ($ED_{50}$). Regional values were also derived from receptor occupancy calculated from the $V_T$ values at baseline and at various blocking doses.
using the equation above modified to add a parameter to permit maximum reduction in $V_T$ of $<100\%$, i.e., accounting for $V_{ND}$. 
RESULTS

**In vitro Competition Binding Studies**

\(^3\)H-BU99008 *in vitro* competition data demonstrated a two-site fit to the rodent brain with BU224 exhibiting an \(IC_{50(\text{high})}\) value of 50.5±12.9nM (Table 1), consistent with previous data (13). In contrast, competition of BU224 in cynomologus brain yielded a single site fit with an \(IC_{50}\) value of 130.2±33.9nM (Table 1). Competition of \(^3\)H-BU99008 from both rat and cynomologus brains by the MAO\(_B\) inhibitor, lazabemide, exhibited very poor inhibition of binding and the MAO\(_A\) inhibitor, moclobemide, showed no inhibition at the highest concentration used (Table 1).

**Radiochemistry**

Injection ready \(^{11}\)C-BU99008 was successfully synthesised with a chemical yield of 32±17% (decay-corrected), radiochemical purity of >99%, and a specific activity of 146±33MBq/nmol (3.95±0.90mCi/nmol, n=19) at the end of synthesis. The identity of the radiolabelled product was confirmed by co-injection with a sample of authentic BU99008, which, under the same elution conditions, showed an identical retention time.

**In vivo Blood Data**

Free fraction \((f_P)\) of \(^{11}\)C-BU99008 in the plasma was high, at 0.68±0.07 (n=19). The amount of total radioactivity measured in plasma was similar between baseline scans and following administration of either the MAO inhibitors or the \(I_2\)BS inhibitor (Figure 1A). In addition, radio-HPLC analysis revealed \(^{11}\)C-BU99008, under baseline conditions, to be rapidly metabolised in plasma, with the parent compound representing about 50% of the total radioactivity 20min following administration (Figure 1B). However, there was a small decrease in parent fraction of \(^{11}\)C-BU99008 following administration of all three inhibitors compared with data acquired under baseline conditions, where the parent compound represented about 50% of the total radioactivity at 15min following administration (Figure 1B).
In vivo PET Studies

Representative baseline PET images and corresponding TACs for $^{11}$C-BU99008 uptake into the rhesus brain are given in Figures 2B and 3A, respectively. $^{11}$C-BU99008 readily entered the brain with the highest uptake observed in the globus pallidus, caudate and thalamus, with moderate uptake in the cortical and putamen regions, and lowest uptake in the cerebellum and occipital cortex. Peak radioactivity concentrations were observed approximately 15–25 min after administration of $^{11}$C-BU99008, followed by a slow washout from all regions (Figure 3A).

The regional TACs were analysed by the reversible 1TC and 2TC models as well as by the MA1 model, (16). The 2TC model produced good fits to the data, but over 20% of fits to baseline data had unreliable $V_T$ estimates, i.e., standard errors (%SE) >20%. Poor reliability occurred most often in smaller, noisier regions and in baseline scans or scans with little effective blockade (see below). The 1TC model showed clear lack of fit in most cases. Also, the $V_T$ values from 1T underestimated those from 2TC by 10-40% (excluding 2TC values with high %SE), with the relationship of $V_T(1T) = 0.87 \times V_T(2T) - 4.2$, $r^2=0.92$. The MA1 method produced good fits and stable estimates for $V_T$ with small differences in $V_T$ values using different $t^*$ values from 20 and 40 min. In the cases where 2TC values had %SE <20%, the relationship between MA1 and 2TC values was $V_T(MA1) = 0.94 \times V_T(2T) - 1.3$, $r^2=0.97$.

Based on the bias from 1TC fits, and the numerous cases of high %SE from 2TC, MA1 with $t^*=20$ min, was chosen as the model of choice for derivation analysis.

Table 2 lists MA1-derived regional $V_T$ values from individual baseline scans. The baseline data acquired from the two rhesus monkeys were demonstrated to be reproducible and consistent throughout the course of the study (Figure 4). Using the MA1 model with $t^*=20$ min, baseline $V_T$ values were highest in the globus pallidus ($114.2 \pm 24.0$ mL/cm), caudate ($109.7 \pm 13.7$ mL/cm) and thalamus ($96.3 \pm 8.1$ mL/cm) and lowest in the cerebellum ($48.1 \pm 4.8$ mL/cm). This rank order of regional $V_T$ for $^{11}$C-BU99008 from the baseline data (globus pallidus>cortex>cerebellum, Table 2) is consistent with reported I$_2$BS densities and...
distribution determined by tissue-section autoradiography in humans (18) and in vivo pig PET (14). Furthermore, the mean rhesus V_T values were significantly correlated ($r^2=0.72$; P<0.05) with the mean V_T values from our previous in vivo porcine $^{11}$C-BU99008 PET data ((14) and unpublished data from the porcine study) (inset, Figure 4).

In vivo blocking studies utilising the MAO_A and MAO_B inhibitors, moclobemide and lazabemide, respectively, did not cause any significant change in binding signal of $^{11}$C-BU99008 to any regions studied (Figures 2C, 2E, and Supplementary Figure 2). In vivo competition using $^{11}$C-BU99008 plus increasing doses of the selective I2BS blocker, BU224, yielded a dose-dependent decrease in uptake of $^{11}$C-BU99008 in all regions studied. The highest dose administered (0.3mg/kg) yielded an apparent near-to-full blockade, suggesting high selectivity of this radioligand for the I2BS (Figures 2G, 3B, and 5, Table 3). The presence of a dose-dependent decrease in binding in the cerebellum suggests this is not a suitable reference region for analysis of $^{11}$C-BU99008.

A global receptor occupancy measure was calculated for the BU224 studies using the occupancy plot (17). Occupancy values ranged from 25-35% for the lowest dose of BU224 (0.01mg/kg) to 93% for the highest dose (0.3mg/kg). These occupancies were plotted vs. administered dose ($d$) in Figure 6 with a fit to the equation $Occ = d/(d + ED_{50})$, which yielded an $ED_{50}$ estimate of 0.022mg/kg for the whole brain (Table 3).

Regional $ED_{50}$ values for BU224 were also calculated for the globus pallidus, frontal cortex and cerebellum using the percentage reduction in $V_T$ instead of the estimated global occupancy value (Supplementary Figure 3A, B, C). Calculated regional $ED_{50}$ values were 0.017mg/kg for globus pallidus, 0.017mg/kg for frontal cortex, and 0.016mg/kg for the cerebellum (Supplementary Figure 3, Table 3).
DISCUSSION

This paper describes the radiosynthesis of $^{11}$C-BU99008 and its characterisation as a novel PET radioligand for the quantification of central I$_2$BS in vivo in rhesus monkeys.

BU99008 was selected as the most suitable compound for radiolabelling with a PET radioisotope for imaging the I$_2$BS, as reported previously by our group (13). In addition, BU99008 exhibited selectivity and nanomolar affinity for the I$_2$BS in the rodent and cynomologus brain (Table 1). Interestingly, a two-site model fit was preferred for the rodent and a one-site fit for the cynomologus brain tissue, suggesting that, in the rodent brain, BU99008 exhibits a degree of affinity for a second binding site which may be unrelated to the I$_2$BS, possibly reflecting a monoamine oxidase binding component (19). Importantly, due to the nature of the I$_2$BS being co-localised with monoamine oxidases (MAO) on the outer membrane mitochondrial enzymes (20), it was essential to investigate the relative affinities of selective MAO$_A$ and MAO$_B$ inhibitors for the I$_2$BS in order to proceed. *In vitro* studies using cynomologus brain demonstrated MAO inhibitors to possess a low affinity for the I$_2$BS in (Table 1). However in the rodent brain tissue, the MAO$_B$ inhibitor possessed a similar affinity for the low affinity binding site exhibited previously in the rodent brain by BU224 (Table 1). These data confirm the notion that the high affinity binding component, *in vitro*, in both the rat and cynomologus brain is expected to represent the I$_2$BS.

In view of these *in vitro* data, and combined with a successful radiolabelling feasibility assessment, we progressed the development of BU99008 as a PET ligand via evaluation in porcine brain, *in vivo* (14). In that study, $^{11}$C-BU99008 demonstrated reversible kinetics and a brain distribution consistent with the known binding site densities and localisation for the I$_2$BS protein, and a dose-dependent decrease in $V_T$ following administration of the selective I$_2$BS blocker, BU224. Further studies in pig showed a small, but relevant, binding component associated with MAO (manuscript in preparation). Therefore, given that relative expression levels of I$_2$BS and MAO differs from species to species, in order to progress
11C-BU99008 for use in human studies we decided to assess this PET ligand further, preclinically, in rhesus monkey, since we predict human brain uptake and in vivo binding characteristics of this particular PET ligand would be more similar to rhesus than porcine. BU99008 was successfully radiolabelled with carbon-11 with good reproducibility, radiochemical yields and high specific activities.

After injection into a rhesus monkey, the radioligand 11C-BU99008 metabolised fairly quickly, with only ~30% of parent compound remaining at 30min post injection. A slight acceleration in metabolism was also observed when blocking agents were given before 11C-BU99008 injection but this effect was modest and unlikely to affect the usability of this ligand.

In monkey brain 11C-BU99008 displayed differential regional uptake. This heterogeneous distribution of 11C-BU99008 exhibited the following rank order: globus pallidus and other basal ganglia regions>cortex>cerebellum, consistent with the known I2BS densities and results from human tissue-section autoradiography (18) and porcine PET imaging experiments (14). While the cerebellum showed the lowest brain uptake there was still a decrease in $V_T$ values after BU224 blockade (Figure 4), indicating cerebellum would be unsuitable as a reference region. This was consistent with the findings of our previous study in pig (14).

An assessment of the intra-subject variability was performed that yielded low to moderate variability in the $V_T$ values obtained for each ROI studied for each subject across multiple baseline scans (Figure 4). Furthermore, comparison of inter-species variability between the rhesus monkeys used in this study and pigs used previously by our group with this PET radioligand (14), demonstrated a significant correlation (inset, Figure 4), suggesting a degree of correspondence between these two species for the I2BS, albeit 11C-BU99008 appears to be affected by MAO inhibitors in the porcine which is a phenomenon not exhibited in the rhesus (manuscript in preparation).
The effects of the MAO\textsubscript{A} inhibitor, moclobemide, and the MAO\textsubscript{B} inhibitor, lazabemide, on \(^{11}\text{C}-\text{BU99008}\) binding were determined \textit{in vivo}, in rhesus monkey and found to cause no significant decrease in \(V_T\) in any of the ROIs assessed (\textit{Supplementary Figure 2}). This key finding suggests that in rhesus monkey brain any contribution of the \(^{11}\text{C}-\text{BU99008}\) signal due to binding to MAO is small or negligible and would not be expected to cause any significant interference with the assessment of \(I_2\text{BS}\) binding signal in this species.

Following administration of increasing blocking doses of BU224, a dose-dependent decrease in \(^{11}\text{C}-\text{BU99008}\) \(V_T\) was observed in all regions of the rhesus brain (Figure 5), confirming the absence of a reference region for this PET radioligand, and remaining consistent with the known distribution profile for \(I_2\text{BS}\) in the brain. The dose-dependent decrease in \(V_T\) observed for all ROI studied is not thought to represent a global change unrelated to the specific binding of \(^{11}\text{C}-\text{BU99008}\) and the blocking by BU224 given that: (a) a heterogeneous signal was observed across the ROI under baseline conditions, (b) differential levels of decrease in \(V_T\) values were observed for each region following increasing doses of BU224, and (c) a plateau at a \(V_T\) value of \(\sim 20\text{mL/cm}\) was achieved for all ROI following the highest dose of BU224 administered (0.3mg/kg; Figure 5). Additionally, the dose-dependent blockade by the selective \(I_2\text{BS}\) inhibitor in the rhesus brain confirmed the specificity of \(^{11}\text{C}-\text{BU99008}\) for the \(I_2\text{BS}\) and demonstrated a blockade of \(\sim 90\%\) across all ROI at the highest dose administered (0.3mg/kg). Interestingly, the \textit{in vivo} ED\textsubscript{50} of BU224 generated across all brain regions was consistent with the presence of one binding site and generated a value of 0.022mg/kg (Figure 6), which is consistent with the known \textit{in vitro} and \textit{ex vivo} properties of this compound (5).

Given these data, we predict that \(^{11}\text{C}-\text{BU99008}\) should demonstrate a similar binding distribution profile in the human brain to that observed from this study in rhesus monkey, where the rank order of regional brain uptake for this ligand would be expected to be globus pallidus>cortical regions>cerebellum. For modelling purposes, since the multi-linear analysis method, MA1, consistently produced good
fits to the data along with reliable and stable estimates of $V_T$ for all regions of the rhesus brain studied, the use of this particular model for analysis of PET data generated using $^{11}$C-BU99008 should be considered in future studies. Work is underway to assess the utility of $^{11}$C-BU99008 as a PET ligand for in vivo imaging and quantification of I$_2$BS in the human population, and if deemed useful, its applicability for determining alterations in I$_2$BS density and distribution in known disease states will be investigated.

**Conclusions**

This manuscript reports the radiolabelling and pharmacological investigation of $^{11}$C-BU99008 as a novel I$_2$BS PET radioligand in rhesus monkeys. *In vivo* distribution of $^{11}$C-BU99008 in the rhesus monkey brain demonstrated the following rank order of regional uptake: globus pallidus>cortex>cerebellum, consistent with the known distribution profile of the I$_2$BS. $^{11}$C-BU99008 displayed reversible kinetics and specificity for the I$_2$BS, with the MA1 model representing the most appropriate analysis method for the derivation of binding parameters for this PET radioligand. The data reported here provides evidence for $^{11}$C-BU99008 to represent a potentially useful PET imaging tool for probing the I$_2$BS. Work is underway in order to progress $^{11}$C-BU99008 for assessment of its clinical utility as a PET radioligand for I$_2$BS.
Acknowledgments

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Tables

Table 1. In vitro I$_2$BS binding affinities of the blocking drugs in rat and non-human primate brain using $^3$H-BU99008.

<table>
<thead>
<tr>
<th>Protein Target</th>
<th>Blocking drug</th>
<th>I/C$_{50}$±SD (nM)</th>
<th>Rat</th>
<th>Cynomologus</th>
</tr>
</thead>
<tbody>
<tr>
<td>I$_2$BS</td>
<td>BU224</td>
<td>50.5±12.9 (high)</td>
<td>130.2±33.9</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>9596±2494 (low)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>MAO$_A$</td>
<td>Moclobemide</td>
<td>&gt;100,000</td>
<td>&gt;100,000</td>
<td></td>
</tr>
<tr>
<td>MAO$_B$</td>
<td>Lazabemide</td>
<td>6445±1733</td>
<td>17370±2068</td>
<td></td>
</tr>
</tbody>
</table>

Displacement of $^3$H-BU99008 by BU224 was best fit to a 2-site model in rat only, for all other instances the data was best fit to a 1-site model; (n=4).
Table 2. Baseline regional $V_T$ values (mL/cm) for $^{11}$C-BU99008 using MA1. Values are computed with $t^* = 20\text{min}$ (n=8).

<table>
<thead>
<tr>
<th>Brain Region</th>
<th>MA1 $V_T$ (Mean±SD)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Globus Pallidus</td>
<td>114.2±24.0</td>
</tr>
<tr>
<td>Caudate</td>
<td>109.7±13.7</td>
</tr>
<tr>
<td>Thalamus</td>
<td>96.3±8.1</td>
</tr>
<tr>
<td>Putamen</td>
<td>75.7±7.8</td>
</tr>
<tr>
<td>Frontal Cortex</td>
<td>75.1±14.2</td>
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<tr>
<td>Cingulate Cortex</td>
<td>85.5±13.0</td>
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<tr>
<td>Occipital Cortex</td>
<td>50.5±7.3</td>
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<tr>
<td>Insula Cortex</td>
<td>76.3±9.2</td>
</tr>
<tr>
<td>Pons</td>
<td>70.1±6.9</td>
</tr>
<tr>
<td>Brainstem</td>
<td>75.2±8.5</td>
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<tr>
<td>Cerebellum</td>
<td>48.1±4.8</td>
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Table 3. Calculated fraction of I$_2$-BS occupied by BU224 using $^{11}$C-BU99008

<table>
<thead>
<tr>
<th>BU224 Dose (mg/kg)</th>
<th>Whole Brain</th>
<th>Globus Pallidus</th>
<th>Frontal Cortex</th>
<th>Cerebellum</th>
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<tr>
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<td>ED$_{50}$</td>
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<td>0.017</td>
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Number in parentheses indicates the animal used to derive the value: monkey 1, monkey 2. Whole brain occupancy was calculated using the occupancy plot (17) and is derived from the slope for all reported brain regions. Values calculated from the $V_T$ determined using the MA1 model. *Percentage occupancies generated from an averaged composite of baseline scans for monkey 1 due to failure to obtain a baseline scan on this study day.
Figure legends

Figure 1. Mean total plasma radioactivity (panel A) and metabolite data (panel B) for $^{11}$C-BU99008 scan. Symbols represent: baseline scan data •; blocking scan data using MAO inhibitors △; blocking scan data using BU224 ○. Each point represents the mean of all the scans in that group, vertical bars represent the standard deviation.

Figure 2. Representative coronal, transverse, and sagittal images of $^{11}$C-BU99008 uptake in rhesus brain. Images are summed from 30-45min following radioligand injection and are displayed as SUVs. A: structural MRI. B,C: baseline and moclobemide pre-block (1mg/kg). D,E: baseline and lazabemide pre-block (0.5mg/kg). F,G: baseline and BU224 pre-block (0.3mg/kg). Paired scans, (B,C), (D,E), and (F,G), were performed on the same day.

Figure 3. Representative time–activity curves for $^{11}$C-BU99008 in selected ROIs in rhesus brain. A: Baseline $^{11}$C-BU99008 scan. B: $^{11}$C-BU99008 scan after administration of BU224 (0.3mg/kg). The symbols represent: Caudate □; Cerebellum △; Frontal cortex ▲; Globus pallidus ▼; Occipital cortex ◆; Putamen ○; Thalamus +.

Figure 4. Bar chart showing the regional distribution volume ($V_T$) of $^{11}$C-BU99008 from two animals (monkey 2, open and monkey 1, filled bars). These represent the mean±SD from four separate baseline scans. The insert shows the significant correlation ($r^2=0.72; P<0.05$) between the mean $V_T$ from this study (both animals) and the pig (n=3; (14) and unpublished data from the pig study). Regional $V_T$ values were generated using the MA1 model ($t^*=20$min) for the rhesus data and the 1TC model for the porcine data.

Figure 5. Bar chart showing the regional distribution volume ($V_T$) of $^{11}$C-BU99008 and the effect of increasing doses of the I$_2$BS ligand BU224: 0.01mg/kg (striped);
0.03mg/kg (chequered); 0.1mg/kg (clear); 0.3mg/kg (grey) and baseline (black).

The bars represent the mean±SD; $V_T$ generated using the MA1 model.

Figure 6. Dose dependent occupancy by BU224 in whole brain, occupancy calculated using the occupancy plot (17) $ED_{50}=0.022$mg/kg. $V_T$ data generated using the MA1 model; monkey 1 • , monkey 2 ○ .
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$V_T$ data generated using the MA1 model; monkey 1 , monkey 2 .