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1 **Title:** Plasticity of fimbrial genotype and serotype within populations of *Bordetella pertussis*:
2 analysis by paired flow cytometry and genome sequencing

3

4 **Running Title:** *B. pertussis* fimbrial serotype plasticity

5

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7

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1 **Abstract**

2 The fimbriae of *Bordetella pertussis* are required for colonisation of the human respiratory tract.
3 Two serologically distinct fimbrial subunits, Fim2 and Fim3, having been considered important
4 vaccine components for many years, are included in the Sanofi Pasteur 5-component acellular
5 pertussis vaccine, and the World Health Organisation recommends the inclusion of strains
6 expressing both fimbrial serotypes in whole cell pertussis vaccines. Each of the fimbrial major
7 subunit genes, *fim2*, *fim3*, and also *fimX*, has a promoter poly(C) tract upstream of its -10 box. Such
8 monotonic DNA elements are susceptible to changes in length via slipped-strand mispairing *in vitro*
9 and *in vivo*, which can potentially cause on/off switching of genes at every cell division. Here we
10 describe intra-culture variability in poly(C) tract lengths and in the resulting fimbrial phenotypes in
11 22 recent United Kingdom *B. pertussis* isolates. Owing to the highly plastic nature of the fimbrial
12 promoters, we used the same cultures for both genome sequencing and flow cytometry. Individual
13 cultures of *B. pertussis* each contained multiple fimbrial serotypes and multiple different fimbrial
14 promoter poly(C) tract lengths, which supports earlier serological evidence that *B. pertussis*
15 expresses both serotypes during infection.

1 **Introduction**

2 *Bordetella pertussis* causes whooping cough and this continues to be a public health problem
3 despite high vaccine coverage. Rates of pertussis disease have increased in many countries in recent
4 years and this is thought to be due to a combination of strain evolution (Mooi *et al.*, 2013) and
5 faster waning immunity provided by acellular pertussis (aP) vaccines compared to whole cell
6 pertussis vaccines (wP) used previously (Tartof *et al.*, 2013). In addition, evidence from a baboon
7 model of pertussis indicates that acellular vaccines may be less effective than whole cell vaccines at
8 preventing acquisition and transmission of the organism (Warfel *et al.*, 2014). It is important for the
9 pathogenesis of pertussis disease that *B. pertussis* is able to adhere to the mucosal surface in the
10 upper respiratory tract via a number of adhesins including filamentous haemagglutinin, pertactin
11 and fimbriae (Mattoo & Cherry, 2005). *B. pertussis* expresses two serologically distinct fimbriae
12 which are composed of either Fim2 or Fim3 subunits (22.5 and 22.0 kDa respectively). Subtypes of
13 Fim2 (*fim2-1* and *fim2-2*) and Fim3 (*fim3-1*, *fim3-2* and *fim3-3*) have also been described (Packard
14 *et al.*, 2004; Tsang *et al.*, 2004). Both Fim2 and Fim3 subunits are assembled in a coil to produce
15 long filaments on the surface of the bacteria (Heck *et al.*, 1996). Fimbriae (co-purified Fim2/3) have
16 been shown to be protective antigens in mouse models of pertussis (Heininger *et al.*, 1998;
17 Robinson *et al.*, 1989a) and are components of an acellular vaccine which showed high efficacy in
18 phase III clinical trials (Gustafsson *et al.*, 1996; Olin, 1997). Household contact studies performed
19 during these trials showed a correlation between antibody responses to fimbriae and protection
20 against disease (Cherry *et al.*, 1998; Storsaeter *et al.*, 1998).

21 Expression of many *B. pertussis* virulence-related genes, including *fim2* and *fim3*, is regulated by
22 the BvgA/S two-component system. (Decker *et al.*, 2012). Transcription of *fim3* is proposed to
23 occur when a dimer of phosphorylated BvgA is bound to the *fim3* promoter's -35 region at the
24 same time as region 4 of the RNA polymerase σ^{80} subunit and the C-terminal domain of an RNA
25 polymerase α subunit (Decker *et al.*, 2011). Poly(C) tracts, and homopolymeric tracts generally, are

1 under-represented in the *B. pertussis* chromosome (Coenye & Vandamme, 2005), and the possibility
2 of rapid antigenic variation via slipped-strand mispairing at these sites has received limited
3 attention. However, each of the *fim* genes possesses a poly(C) tract between its -35 and -10
4 promoter regions. Transcription-permissive lengths for the fimbrial poly(C) tracts have been
5 determined (Willems *et al.*, 1990; Chen *et al.*, 2010), but the frequency of slipped-strand mispairing
6 within them has not been previously described. The fimbrial poly(C) tracts of *B. pertussis* have a
7 notable counterpart in the *porA* promoter of *Neisseria meningitidis*, where altered transcription
8 levels, caused by changes in the length of a poly(G) tract between the -35 and -10 regions, can
9 facilitate the evasion of bactericidal anti-PorA antibodies (Tauseef *et al.*, 2013).

10 In this study, we have analysed whole genome sequencing reads from a panel of 96 *B. pertussis*
11 strains isolated in the United Kingdom between 1920 and 2012, and shown that cultures of *B.*
12 *pertussis* contain multiple subpopulations with differing fimbrial poly(C) tract lengths. In addition,
13 we preserved aliquots from 22 of the cultures used for genome sequencing and have analysed Fim2
14 and Fim3 expression on individual bacteria using labelled anti-Fim2 and anti-Fim3 monoclonal
15 antibodies in two-colour flow cytometry. This demonstrated that bacteria expressing Fim2 and/or
16 Fim3 are present in cultures previously shown by standard slide agglutination methods to express
17 only one fimbrial serotype. Thus, these data show that expression of Fim2 and Fim3 by *B. pertussis*
18 is more flexible than generally acknowledged, and this flexibility may help to explain the rapid
19 shifts in predominant serotype observed in many countries.

20 **Materials and methods**

21 **Bacteria**

22 Ninety-six *B. pertussis* isolates UK001 to UK06, UK008 to UK011, UK014 to UK018, UK020 to
23 UK100 were obtained from the Respiratory and Vaccine Preventable Bacteria Reference Unit
24 (RVPBRU), Public Health England (PHE) Colindale, London. The majority of these isolates ($n =$

1 87) were submitted by hospital laboratories as part of the enhanced surveillance of pertussis (Van
2 Buynder *et al.*, 1999). Additional isolates were previously obtained from the Wellcome Bacterial
3 Collection, held by the National Collection of Type Cultures (NCTC) ($n = 4$); the NCTC ($n = 2$) and
4 the former Centre for Applied Microbiology and Research, now PHE Porton Down, Salisbury ($n =$
5 3) (Fry *et al.*, 2001).

6 Following culture for 48 to 72 h on blood charcoal agar (PHE Media Services, Colindale), *B.*
7 *pertussis* isolates were serotyped using a slide agglutination assay with rabbit polyclonal antisera
8 against antigens 1, 2, and 3 (89/596, 89/598, and 89/600, respectively; National Institute for
9 Biological Standards and Controls, Potters Bar, United Kingdom). Antigen 1 (agglutinogen 1) is
10 expressed by all isolates of *B. pertussis*, thus the notation Fim–, Fim2, Fim3 and Fim2,3 (Hallander
11 *et al.*, 2005) is used instead of the historical notation serotype 1, serotype 1,2 serotype 1,3 and
12 serotype 1,2,3 (Robinson *et al.*, 1989b) as previously described (Litt *et al.*, 2009). The terms Fim–
13 and Fim2–Fim3– are used interchangeably in the text. Cultures for this study were grown on
14 charcoal agar for 72h at 37°C. For flow cytometry analysis, bacteria were resuspended in PBS
15 containing 1% formaldehyde and the OD_{600 nm} adjusted to 2.0.

16 **Genome sequencing**

17 Genome sequencing of UK isolates UK001 to UK100 was performed at the Sanger Institute and
18 detailed analysis of these sequences will be the subject of a paper in preparation. Briefly, paired-end
19 libraries were prepared from genomic DNA, amplified with a KAPA HiFi library amplification kit
20 (Kapa Biosystems Inc, Wilmington MA), and sequenced as 150 bp paired-end reads using a MiSeq
21 desktop sequencer (Illumina Inc, San Diego, CA). The unassembled short reads are available in
22 fastq.gz format from the Sanger Institute's ftp site. Locations of the files are in Table S1
23 (supplementary materials). Additional genome sequence files (non-UK isolates) are listed in Table
24 S2 (supplementary materials).

1 **Extraction of poly(C)-tract–spanning reads from fastq.gz files**

2 Each fastq.gz file was read with *kseq_test*, a minimal example program for the `kseq.h` fastq parser
3 library (Li, 2011; Li, 2012). Output from *kseq_test* was piped into a script written in AWK (Aho *et*
4 *al.*, 1988), within which the sequence field of each Illumina read was put through two successive
5 filters.

6 The first filter compared each read to the set of all plausible fimbrial poly(C) tracts (6–20 base
7 pairs) flanked by a minimal *B. pertussis* Tohama I–derived context (Parkhill *et al.*, 2003). The
8 search patterns used in this first filter are shown as regular expressions in Table 1. The *fim2* regular
9 expressions contain a [CT] character list, corresponding to the deoxythymidine that punctuates the
10 poly(C) tract of *B. pertussis* Tohama. Three regular expressions are necessary to prevent *fim3* and
11 *fimX* sequences from being misidentified as each other. Reverse-complement patterns, not shown in
12 Table 1, were also used, giving a total of 308 patterns upon evaluation of the regular expressions.
13 Non-matching reads (including all reads ending within poly(C) tracts) were discarded, and exact
14 matches were tested in the second filter.

15 The second filter took the segments of each read outside the initial match, joined them and aligned
16 the resulting chimaera to the equivalent joined 30 bp segments from *B. pertussis* Tohama I. For this
17 alignment, EMBOSS *needle* (Rice *et al.*, 2000) was used via AWK’s `getline`. The alignment was
18 forced to be local with respect to query and global with respect to reference (by invoking *needle*
19 with an endweight, a zero end-open penalty and high gap open/extend penalties). Reads were given
20 a positive plausibility score if this second filter resulted in a plausible alignment (aligned join-sites,
21 $\geq 66.7\%$ identity across ≥ 10 bp where flanking sequence was available on both sides of the initial
22 match, $\geq 80\%$ identity across ≥ 20 bp where flanking sequence was available on only one side of
23 the initial match); otherwise reads were given a negative score. Each fastq record was reassembled

1 into a one-line tab-delimited format and augmented with gene name (*fim2*, *fim3* or *fimX*), isolate
2 name, fastq.gz file name and plausibility score.

3 The *kseq_test* preprocessing step and the *AWK/needle* double filter were combined as a single
4 Bourne shell script, and summary counts were extracted from the tab-delimited output using a
5 second shell script (supplementary materials: `MeasurePolyC.sh` and
6 `FastqplusToTable.sh`). The AWK script embedded in `MeasurePolyC.sh` induced a
7 memory leak in GNU *awk*, but not in *nawk* or *mawk*. Arithmetic means and standard deviations
8 (Table 3 and supplementary materials, `CoverageStatsUKstrains.awk`) were calculated
9 using the Welford–Knuth algorithm (Welford, 1962; Knuth, 1997, pp. 231–232).

10 **Detection of Fim2 and Fim3 by flow cytometry**

11 200 µl of each suspension ($OD_{600} = 0.1$) of plate-grown bacteria was added to a well of a 96-well
12 microtitre plate. The plate was centrifuged at $3060 \times g$ for 5 min. and supernatant was removed.
13 Fim2- and Fim3-specific monoclonal antibodies (06/124 and 06/128 respectively, obtained from the
14 National Institute of Biological Standards and Control, UK) were bound to AlexaFluor 488 and
15 Alexafluor 647, respectively, using a Zenon[®] Alexa Fluor[®] Mouse IgG₁ Labelling Kit (Life
16 Technologies, Paisley, UK). 100 µl of each antibody at a dilution of 1 in 5000 was added to each
17 test well and pellets were resuspended. The plate was incubated at 25 °C with shaking, for 30 min.
18 The plate was centrifuged as above and washed, and pellets were resuspended in 200 µl blocking
19 buffer (PBS, 1 % BSA). Samples were analysed using a Beckman Coulter CyanADP flow
20 cytometer, and Summit[®] software was used to calculate the median fluorescence of each
21 fluorophore for each sample. Unstained bacterial cells were used to verify the absence of
22 fluorophore-independent counts, which are minimised by the use of filter-sterilised buffers and
23 sheath fluid and by the collection of 10,000 events per sample.

1 A gate was drawn to include the bacteria plus conjugate control (no antibody), resulting in a median
2 fluorescence >20 being considered positive for Fim expression. To ascertain the percentage of
3 bacterial cells expressing Fim2 or Fim3, AlexaFluor 488 fluorescence was plotted against
4 AlexaFluor 647 fluorescence for each sample. Gates were drawn to enclose the negative control
5 sample and used to determine the demarcation between a positive and negative result. This gate was
6 then applied to all test samples, allowing the determination of the percentage of events, or
7 individual bacteria, which were not fluorescent, singly positive for AlexaFluor 488, singly positive
8 for AlexaFluor 647 or doubly positive for both Alexafluor 488 and Alexafluor 647, thereby
9 determining the percentage of individual bacteria expressing Fim2, Fim3 or both Fim2 and Fim3
10 within a population.

11 **Results**

12 **Multiple fimbrial promoter variants within cultures**

13 From the 96 unassembled UK *B. pertussis* genome sequences, 2648 reads were matched exactly to
14 one of the *fim2* regular expressions in Table 1 or to one of the reverse complements thereof. After a
15 second filter to eliminate false positives and uncertain matches, 2580 *fim2* poly(C) tract–spanning
16 reads were kept. The twice-filtered reads contained *fim2* poly(C) tracts ranging in length from 6 to
17 20 bp. Multiple *fim2* poly(C) tract lengths were found within samples (Table 2), with a mean intra–
18 isolate range of 4.2 bp (standard deviation = 2.3 bp). The deoxythymidine residue that punctuates
19 the *fim2* poly(C) tract of *B. pertussis* Tohama was also found in UK001, UK002 and UK004 (58%,
20 17% and 23% of reads respectively), and was counted as an internal deoxycytidine for the purpose
21 of poly(C) tract length enumeration. Our numbering convention thus differs from that of Chen *et al.*
22 (2010), who examined only T-punctuated *fim2* poly(C) tracts, and whose enumeration excluded
23 deocytidines on the 5' side of the punctuating deoxythymidine.

1 Using the equivalent double filter for *fim3*, 1914 reads were matched exactly by the first filter, and
2 1660 *fim3* poly(C) tract–spanning reads were kept after the second filter. The twice-filtered reads
3 contained *fim3* poly(C) tracts ranging in length from 6 to 20 bp. As with *fim2*, multiple poly(C) tract
4 lengths were found within samples (Table 2), with a mean intra–isolate range of 5.2 bp (standard
5 deviation = 3.0 bp).

6 For *fimX*, 4463 reads were matched by the first filter, and 4381 *fimX* poly(C) tract–spanning reads
7 were kept after the second filter. The *fimX* poly(C) tracts were less variable than those of the other
8 two fimbrial genes (mean intra–isolate range = 1.1 bp, standard deviation = 0.4). In 77 of the 96
9 genome-sequenced UK isolates, *fimX* poly(C) tracts did not exceed 7 bp in length. Isolate UK071
10 contained a *fimX* poly(C) tract of 9 bp, the longest among the UK isolates.

11 Read lengths for UK025 to UK100 were all 150 bp (average of 1.26 million reads per isolate,
12 standard deviation = 0.26 million), whereas read lengths for other available *B. pertussis* Illumina
13 sequences were 54 bp (average of 5.40 million reads per isolate for UK strains, standard deviation =
14 1.66 million). For *fim2* and *fimX*, poly(C) tract–spanning reads were more numerous in the genomes
15 sequenced at 54 bp per reads, whereas for *fim3*, poly(C) tract–spanning reads were more numerous
16 in the genomes sequenced at 150 bp per read (Table 3). Since a 54 bp read is less likely than a 150
17 bp read to span a given sequence, the higher tally of *fim2* and *fimX* poly(C) tracts in the 54 bp
18 genome sequences can be attributed to the higher coverage of those genomes (71.4-fold versus
19 46.1-fold for the 150 bp genome sequences). For *fim3*, the higher tally in the 150 bp genome
20 sequences can be attributed to the broader size range of poly(C) tracts ($\overline{range}_{fim3} = 5.2$ bp versus
21 $\overline{range}_{fim2} = 4.2$ bp), with the longer part of this range being more likely to be encompassed by
22 longer reads.

1 After normalisation to genome coverage, the yields of *fim2* and *fim3* poly(C) tract-spanning read
2 were higher from the 150 bp genomes than from the 54 bp genomes, but the yield for *fimX* stayed
3 higher for the 54 bp genomes (Table 3, column 8).

4 **Presence of multiple serotypes within cultures**

5 In this study an aliquot of culture prepared for genome sequencing from strains UK025 to UK046
6 was preserved by the addition of formaldehyde to 1 %. Individual bacteria in these preserved
7 suspensions were analysed for expression of Fim2 and Fim3 using anti-Fim2 and anti-Fim3
8 monoclonal antibodies and two-colour flow cytometry. The flow cytometry scatter plots obtained
9 and the poly(C) tract lengths determined for each isolate are shown in Figure 1. In each of 22
10 isolates examined by two-colour flow cytometry, *B. pertussis* cells of all four possible fimbrial
11 serotypes (Fim2⁻Fim3⁻, Fim2⁻Fim3⁺, Fim2⁺Fim3⁻ and Fim2⁺Fim3⁺) were present (Fig. 1,
12 scatterplots; Fig.2). One fimbrial serotype was dominant (>55 % of the population) in 21 out of the
13 22 isolates (Figure 2). In isolate UK043, which had a *bvg*⁻ colony morphology, Fim2⁻Fim3⁻ cells
14 were dominant (89% of the population). FimX surface expression was not investigated owing to the
15 lack of available antibodies.

16 Gates were set so that the bacteria plus conjugate control was 99% enclosed inside the Fim2⁻Fim3⁻
17 quadrant. Although this may misidentify some events as Fim2⁻Fim3⁻ (for instance the lower portion
18 of the Fim3⁺ cloud of UK042 in Figure 1), it does ensure that false positives are avoided. The
19 diagonal streaking that can be seen in the Fim2⁺Fim3⁺ quadrant of some scatterplots (Fig. 1) is
20 attributable to clumping of bacteria, with single bacteria towards the lower left and clumped
21 bacteria towards the upper right. This analysis used a gate that excluded most clumped bacteria, but
22 when a larger gate was used so as to include more clumped bacteria, the fraction of Fim2⁺ or Fim3⁺
23 bacteria remained very similar (data not plotted). Hence the results presented here are not dependent
24 on the chosen flow cytometry analysis gate.

1 In all 22 of these isolates, the *fim2* and *fim3* promoter poly(C) tracts were present in the Illumina
2 sequencing data as multiple variants. The broadest range for *fim2* occurred in UK086, with poly(C)
3 tract lengths ranging from 7 to 17. The broadest range for *fim3* occurred in UK057, with poly(C)
4 tract lengths ranging from 7 to 19.

5 **Relationship between median fimbrial poly(C) tract length and serotype**

6 Among the 21 *bvg*⁺ isolates studied by paired genome sequencing and two-colour flow cytometry,
7 Fim2 median fluorescence was high in 8 phenotypically *bvg*⁺ isolates that had median *fim2* poly(C)
8 tracts lengths of 12 to 16 (Table 2). In the other 13 *bvg*⁺ isolates tested in this paired analysis, Fim2
9 median fluorescence was at least 13-fold lower, and median poly(C) tract lengths were either 10 or
10 11. *fim2* median poly(C) tract lengths longer than 16 were not seen in the 22 isolates tested by flow
11 cytometry but were seen in four of the other 74 UK isolates.

12 Fim3 median fluorescence was high in 13 phenotypically *bvg*⁺ isolates that had median *fim3*
13 poly(C) tracts lengths of 13 or 14. In the other 8 *bvg*⁺ isolates tested, Fim3 median fluorescence was
14 at least 11-fold lower, and median *fim3* poly(C) tract lengths were in the range 10 to 13. *fim3*
15 median poly(C) tract lengths longer than 14 were not seen in the 22 isolates tested by flow
16 cytometry but were seen in four of the other 74 UK isolates.

17 In UK043, *fim2* and *fim3* median poly(C) tract lengths were 15 and 14 respectively, but median
18 fluorescence was low for both Fim2 and Fim3. Over 70% of cells were Fim2⁻Fim3⁻ (Fig 2). This
19 isolate had *bvg*⁻ colony morphology, and this decoupling of surface expression from poly(C) tract
20 length was therefore expected. It was, however, serotyped by agglutination as Fim3⁺.

21 Among the other 74 sequenced UK isolates (which were not analysed by flow cytometry) UK015
22 provided the sole example of a homogeneous *fim2* promoter (38 Illumina reads spanning the *fim2*
23 poly(C) tract, all with a poly(C) tract length of 9 nucleotides). The range of *fim2* poly(C) tract

1 lengths in UK010 was not determined since only one spanning read was available. There were no
2 unambiguous examples of homogenous *fim3* promoters (only one spanning read was available for
3 UK017).

4 For Fim2 and Fim3, the relationships between promoter poly(C) tract lengths and expression are
5 therefore as follows: a bvg⁺ population is Fim2⁺ if the median poly(C) tract length is > 11 (upper
6 bound not established, but at least 16), and a bvg⁺ population is Fim3⁺ if the median poly(C) tract
7 length is > 13 (upper bound not established, but at least 14).

8 Optimal poly(C) tract lengths for *B. pertussis* fimbrial promoter activity have been proposed from
9 observations made with luciferase reporter gene constructs (Chen *et al.*, 2010). In nineteen of the 21
10 bvg⁺ isolates described here, surface expression of Fim2 or Fim3 correlated with a majority of *fim2*
11 or *fim3* promoters having poly(C) tracts unambiguously in the proposed active range (Figure 1,
12 shaded in bar charts). . However, the correlation broke down for two isolates. In UK037, the
13 majority of *fim2* poly(C) tracts were outside the proposed optimal range, but the majority of cells in
14 this culture were Fim2⁺. Similarly, in UK044, the majority of *fim3* poly(C) tracts were outside the
15 proposed optimal range, while the majority of cells were Fim3⁺.

16 **Relationship between poly(C) tract length and serotype as determined by** 17 **agglutination**

18 Serotypes determined by agglutination were not uniformly in agreement with poly(C) tract length
19 distributions. Thirteen of the 96 genome-sequenced isolates (including UK043) had serotypes by
20 agglutination that conflicted with the rules proposed above connecting median poly(C) tract length
21 to expression (Table 4); and if upper bounds for “active” median poly(C) tract lengths could be
22 established, additional conflicting examples would probably be apparent. For 59 of the 96 isolates,
23 serotypes by agglutination were in agreement with median poly(C) tract lengths.

1 Serotypes by agglutination were in agreement with the flow cytometric measurements of Fim2 and
2 Fim3 for twenty of the 22 isolates that were analysed by flow cytometry (Table 2). The exceptions
3 were UK030 (Fim2⁻Fim3⁺ by agglutination, but 88% Fim2⁺Fim3⁻ by flow cytometry) and the *bvg*⁻
4 isolate UK043. Four of the 96 genome-sequenced UK isolates were serotyped as Fim2,3, and one
5 was serotyped as Fim⁻, but these five were not included in the flow cytometric analysis
6 (Supplementary Table S1: UK001, UK003, UK006, UK010 and UK068).

7 **Subpopulations with the potential to express FimX**

8 In 87 of the 96 UK isolates, all *fimX* poly(C) tracts were 8 bp or shorter, suggesting that these
9 strains are FimX⁻. The longest *fimX* poly(C) tract found in a UK isolate was 9 bp, in UK024, which
10 is shorter than the proposed optimum length for expression (Chen *et al.*, 2010).

11 Since no reagents are available to detect FimX protein, the possible existence of FimX⁺
12 subpopulations can be tested only by scanning genome sequences for *fimX* poly(C) tracts of
13 plausibly transcription-permissive lengths. A further 275 unassembled *B. pertussis* genome
14 sequences, all from non-United Kingdom isolates, were therefore filtered through the same script as
15 was used for the 96 UK isolates. In each of 195 non-UK isolates, the longest observed *fimX* poly(C)
16 tract length was 7 bp, and the longest *fimX* poly(C) tract lengths observed in the whole dataset were
17 10 bp (found in two isolates), which is shorter than the proposed optimum length for expression.

18 **Discussion**

19 Fimbriae are required by *B. pertussis* for colonisation of the human respiratory tract; and we have
20 shown in this study using two-colour flow cytometry, that individual cultures each contain cells of
21 more than one fimbrial serotype. In the same cultures, we have detected multiple variants of the
22 promoters for each of the serologically distinct fimbrial major subunits. This study therefore
23 provides cellular and genomic corroboration for the suggestion by Heikkinen *et al.* (2008), and

1 recently reported serological evidence, that *B. pertussis* expresses Fim2 and Fim3 during infection
2 irrespective of the predominant serotype of case isolates (Hallander *et al.*, 2013).

3 The intra-culture serotype heterogeneity described here reflects the intrinsic susceptibility of
4 repetitive DNA elements (in this case, promoter poly(C) tracts) to slipped-strand mispairing during
5 DNA replication. The steric feasibility of bulge defects in the base-pairing of nucleic acids was
6 demonstrated more than 50 years ago, and the formation of such defects by slipped-strand
7 mispairing was postulated immediately as a mechanism for point mutations in DNA regions of low
8 complexity (Fresco & Alberts, 1960). Insertions and deletions within runs of short (1 to 10 bp)
9 repeated elements have subsequently been observed in human genetic disorders and in antigenic
10 switching by pathogens (reviewed by van der Woude & Bäumlér, 2004). Sporadic Huntingdon's
11 disease arises from expansion of a run of sequential CAG glutamine codons in the *HTT* gene
12 (Myers *et al.*, 1993), and similar CAG expansions are seen in several other neurodegenerative
13 disorders. Analogous in-frame trinucleotide repeats (AGT or TCT) occur in the MgPa adhesin
14 operon of *Mycoplasma genitalium*, and variations in the number of repeat units have been observed
15 within and between isolates (Ma *et al.*, 2012). The opacity genes of *Neisseria gonorrhoeae* are
16 subject to reversible frameshifts, at a rate of 10^{-4} – 10^{-3} /cell/generation, resulting from changes in the
17 number of copies of a CTCTT repeat in the signal peptide-encoding sequence (Murphy *et al.*,
18 1989). Variation in the number of copies of a TAAA repeat in the *nadA* promoter of *Neisseria*
19 *meningitidis* MC58 leads to colonies with differential NadA expression appearing on the same
20 culture plate (Martin *et al.*, 2003).

21 The simplest possible repeat elements, homopolymeric tracts, account for many reported instances
22 of slipped-strand mispairing in bacteria. There are 5684 homopolymeric tracts longer than 4 bp in
23 the *B. pertussis* Tohama I genome sequence (Parkhill *et al.*, 2003), of which 78% are 5 bp in length.
24 The 6 bp homopolymeric tracts account for 15% of the total, and frequency decreases with

1 increasing length. In a survey of expected versus observed homopolymeric tract frequencies in 139
2 sequenced bacterial genomes, *B. pertussis* was ranked twelfth lowest with an expected/observed
3 ratio of 0.23 (Coenye & Vandamme, 2005), and most reports of homopolymeric tract-associated
4 antigenic variation are from bacteria with higher expected/observed ratios (for instance, 2.1 in
5 *Neisseria meningitidis* and 3.5 in *Helicobacter pylori*). It is therefore striking that the earliest report
6 of *B. pertussis* slipped-strand mispairing was in the *bvgS* gene, where the impact is magnified by
7 pleiotropy. Stibitz *et al.* (1989) found that bvg^- strains arose by frameshifting expansion of a C₆
8 tract at *bvgS* codons 1103–4, and that the frequency of the $bvg^{+/-}$ switch varied between two strains.

9 Transcription from the fimbrial promoters requires that their poly(C) tracts be longer (Chen *et al.*,
10 2010) than the footprints of some DNA polymerases (Beese *et al.*, 1993; Eom *et al.*, 1996), so the
11 heterogeneity of fimbrial promoters reported here is perhaps unsurprising. Fazekas *et al.* (2010)
12 were able to improve the fidelity of PCR amplification across homopolymeric tracts by using PCR
13 enzymes with extended DNA-binding footprints. Compared to a 68°C PCR extension, DNA
14 replication in *B. pertussis* should be less vulnerable to slipped-strand mispairing because the 3' end
15 of the nascent strand is less likely to dissociate at physiological temperatures. Nevertheless, the
16 imperfect fidelity of C₆ replication in *bvgS*, along with the apparent selection against
17 homopolymeric tracts throughout the genome, suggests that *B. pertussis* is unable to replicate
18 homopolymeric tracts with high fidelity if they are longer than 5 bp. The fimbrial poly(C) tracts of
19 *B. pertussis*, being several nucleotides longer, are directly analogous to the poly(C) tract of *opc*
20 (Sarkari *et al.*, 1994) and the poly(G) tract of *porA* (van der Ende *et al.*, 1995; Tauseef *et al.*, 2013),
21 both in *N. meningitidis*. In each of these examples, expression of a surface antigen is modulated by
22 frequent and apparently stochastic changes in the spacing between the –35 and –10 promoter
23 regions.

1 In an organism that uses slipped-strand mispairing to effect antigenic variation, cell division must,
2 at least some of the time, result in daughter cells inheriting surface antigens that they are unable to
3 express anew. The correlation between fimbrial poly(C) tract length distribution and surface
4 expression is therefore likely to be imperfect, especially if antigenic variation is rapid. This may be
5 exemplified in the present study by the two isolates with poly(C) tracts lengths predominantly
6 outside the transcription-permissive range estimated by Chen *et al.* (2010); though an alternative
7 explanation might be that the estimates of transcription-permissive lengths were compromised by
8 slipped-strand mispairing in the reporter gene constructs used. In this study, we obtained statistical
9 estimates of transcription-permissive fimbrial poly(C) tract lengths, as distinct from the previous
10 reporter gene-based estimates, but isolates with conflicting serotype (by agglutination assay) were
11 still apparent (Table 4). However, the objective of the present study was not to determine precise
12 limits for promoter activity, and the range of poly(C) tract lengths in individual cultures was broad
13 enough to encompass the multiple serotypes seen in the same cultures (Figure 1).

14 The major impediment to estimating *in vivo* poly(C) tract diversity lies in the potential for
15 laboratory DNA polymerases to add to the diversity *in vitro*. In our hands, *B. pertussis* fimbrial
16 poly(C) tracts can be sequenced successfully using dideoxy chemistry if they are amplified in
17 *Escherichia coli* as plasmid inserts, implying correct amplification by the *E. coli* DNA polymerase.
18 However, when PCR-amplified fimbrial promoters are sequenced without cloning, multiple
19 superimposed traces appear on the downstream side of the poly(C) tract, representing multiple
20 poly(C) tract lengths of which some might represent slipped-stranded mispairing in the PCR tube
21 rather than in the organism. For the present study, we sought to avoid PCR-generated diversity by
22 using high coverage sequence data. The 10-cycle amplification in the Kapa HiFi library
23 amplification protocol amounts to 10000-fold less amplification that takes place in a 30-cycle PCR
24 reaction, so the number of *in vitro*-generated poly(C) tract lengths is likely to be minimal. In

1 addition, only a few of the poly(C) tract length distributions in Figure 1 resemble the bell-shaped
2 distribution that would be expected for purely PCR-generated diversity.

3 The distribution of *B. pertussis* fimbrial serotypes, as determined by agglutination, has varied
4 between countries and over time, and may have been influenced by differing vaccination policies
5 (reviewed by Poolman & Hallander, 2007). Our data indicate that the result returned by an
6 agglutination assay is the serotype of the largest subpopulation in the sample being assayed; so the
7 fact that temporal and geographical trends can be discerned at all suggests that *B. pertussis*
8 populations are stably diverse, with selective pressure being required to displace a dominant
9 serotype. Detection limits for current serotyping assays are not known, but could be determined
10 with blended samples following separate flow cytometric analyses. The proportion of United
11 Kingdom isolates with serotype Fim2⁺ (by agglutination) dropped sharply from 47 % (*n* = 110) in
12 1988 to 2001, to 3 % (*n* = 184) in 2002 to 2004, and 2 % (*n* = 166) in 2005 to 2008, after the
13 introduction of aP vaccines (a preschool booster in 2001 and a replacement for the previous wP
14 given at 2, 3 and 4 months in 2004 (Litt *et al.*, 2009), but increased in 2011–2012 (Alexander *et al.*,
15 2012) to levels comparable with the pre-vaccine and whole-cell vaccine eras. It is possible that
16 population immunity to the predominant serotype builds up until a tipping point is reached and the
17 expression of the alternative serotype is favoured. However, there is no clear pattern in the median
18 poly(C) tract lengths over these periods, partly because of a sampling bias in favour of 2012 isolates
19 (Figure S1 supplementary materials).

20 FimX is 73% similar to Fim2 and 70% similar to Fim3, but has been detected only by mass
21 spectrometry and only in one *B. pertussis* isolate (Tefon *et al.*, 2011). No laboratory antibodies are
22 available for more rapid detection, and the capacity of FimX to substitute for Fim2 and Fim3 is
23 unknown. Most of the *fimX* poly(C) tracts counted in the present study were 6 to 8 bp long, which is
24 probably too short to allow transcriptional initiation. The emergence of a FimX⁺ strain from one

1 with exclusively short *fimX* poly(C) tracts might conceivably start with a low-probability DNA
2 replication error followed by increasingly probable recurrences as the poly(C) tract grew further
3 beyond the size of the DNA polymerase footprint in successive generations. The analogous
4 phenomenon in human CAG expansion disorders, where successive generations are at increased
5 risk as their CAG repeats expand, is termed genetic anticipation (Walker, 2007). The 10 bp *fimX*
6 poly(C) tracts found in two strains are comparable in length to some of the *fim2* and *fim3* poly(C)
7 tracts (Figure 1); but a robust investigation of FimX genetic anticipation requires more data than
8 were available in the genome sequences (Table 3 column 4), and would be better accomplished by
9 generating specific antibodies and using them in flow cytometry.

10 Thus, in this study we have characterised the distribution of poly(C) tract lengths in the promoters
11 of *fim2* and *fim3* and shown that cultures contain multiple sub-populations with differing poly(C)
12 lengths. For a subset of strains, we determined expression of Fim2 and Fim3 on each bacterium
13 using two-colour flow cytometry, and showed that whilst there is a predominant serotype in each
14 culture, multiple combinations of Fim2 and Fim3 expression occur in a single culture. The optimal
15 poly(C) tract lengths for expression of Fim2 and Fim3 were greater than 11 bp or 13 bp respectively
16 for most strains, but for 13 out of 96 strains the observed serotype conflicted with that predicted by
17 median poly(C) tract length (Table 4). This demonstrates that expression of Fim2 and Fim3 is more
18 fluid than previously thought and confirms the serological evidence that *B. pertussis* expresses both
19 Fim2 and Fim3 *in vivo*. It is therefore important for *B. pertussis* vaccines that rely on protection
20 mediated by fimbriae to elicit antibodies against both fimbrial antigens.

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1 Tables

2 Table 1

Gene		Regular expression
<i>fim2</i>	(1)	<code>gccc[ct]c\{2,17\}taagacc</code>
	(2)	<code>gaatcagccc[ct]c\{2,17\}t</code>
<i>fim3</i>	(1)	<code>gc\{6,20\}ggacctg</code>
	(2)	<code>ccatcagc\{6,20\}gg</code>
	(3)	<code>accatcagc\{6,20\}g</code>
<i>fimX</i>	(1)	<code>gc\{6,20\}gaggcgt</code>
	(2)	<code>ccatcagc\{6,20\}ga</code>
	(3)	<code>tccatcagc\{6,20\}g</code>

3 Table 1. Patterns used in the first of two sequential text filters to identify genome sequence reads
4 spanning the promoter poly(C) tracts of *fim2*, *fim3* or *fimX*. The patterns are printed in ED regular
5 expression syntax (Thompson & Ritchie, 1971). The reverse-complement of each pattern was also
6 used.

1 **Table 2**

Isolate	Year of isolation	Serotype by agglutination	ENA identifier	Fim2				Fim3			
				No. of reads	Median poly(C) tract length	Range of poly(C) tract lengths	Median fluorescence	No. of reads	Median poly(C) tract length	Range of poly(C) tract lengths	Median fluorescence
UK025	2008	Fim3	ERS176852	33	10	3 (9 to 11)	4.6	8	14	6 (13 to 18)	320.2
UK026	2008	Fim3	ERS176853	28	10	2 (10 to 11)	4.3	12	14	3 (13 to 15)	251.4
UK027	2008	Fim3	ERS176854	24	10	3 (9 to 11)	4.6	15	14	4 (12 to 15)	335.3
UK028	2009	Fim3	ERS176855	27	10	3 (9 to 11)	4.6	10	14	4 (12 to 15)	360.7
UK029	2009	Fim2	ERS176856	6	16	4 (14 to 17)	708.8	20	13	5 (11 to 15)	12.8
UK030	2011	Fim3	ERS176857	4	16	3 (15 to 17)	703.3	26	10	3 (9 to 11)	8.1
UK031	2011	Fim2	ERS176858	9	15	4 (13 to 16)	666.5	24	10	3 (9 to 11)	8.1
UK032	2011	Fim3	ERS176859	39	11	3 (10 to 12)	4.6	22	14	8 (12 to 19)	209.8
UK033	2011	Fim3	ERS176860	30	10	2 (9 to 10)	4.6	5	14	2 (14 to 15)	246.7
UK034	2011	Fim2	ERS176861	6	15	4 (13 to 16)	608.7	20	11	3 (10 to 12)	9.3
UK035	2012	Fim2	ERS176862	11	15	7 (11 to 17)	361	21	12	3 (11 to 13)	12
UK036	2012	Fim3	ERS176863	31	10	3 (9 to 11)	4.1	6	14	2 (13 to 14)	666.5
UK037	2012	Fim2	ERS176864	12	14	6 (11 to 16)	255.8	38	10	3 (9 to 11)	9.9
UK038	2012	Fim3	ERS176865	30	10	3 (9 to 11)	4.9	17	14	8 (12 to 19)	394.6
UK039	2012	Fim3	ERS176866	28	10	2 (9 to 10)	4.6	11	14	9 (12 to 20)	341.5
UK040	2012	Fim2	ERS176867	11	15	7 (13 to 19)	270	37	12	8 (11 to 18)	9.9
UK041	2012	Fim2	ERS176868	7	15	5 (13 to 17)	270.2	27	12	5 (11 to 15)	9.6
UK042	2012	Fim3	ERS176869	28	10	2 (10 to 11)	4.7	6	14	3 (12 to 14)	178.3
UK043	2012	Fim3	ERS176870	11	15	8 (9 to 16)	4.6	11	14	3 (13 to 15)	15.3
UK044	2012	Fim3	ERS176871	36	10	3 (9 to 11)	4.5	6	13.5	8 (11 to 18)	291
UK045	2012	Fim3	ERS176872	35	10	3 (10 to 12)	4.2	5	14	3 (13 to 15)	312.8
UK046	2012	Fim3	ERS176873	43	10	4 (9 to 12)	4	9	14	6 (12 to 17)	191.9

2 Table 2. Promoter poly(C) tract lengths and surface expression of Fim2 and Fim3 in *B. pertussis* isolates UK025 to UK046. ENA identifiers are
3 European Nucleotide Archive sample names. Serotype by agglutination uses the nomenclature of Hallander *et al.* (2005). Each isolate was cultured
4 twice: once for serotyping by agglutination and once for the other two analyses, with cultures being split between flow cytometry and whole genome
5 sequencing. Values (apart from fluorescence) for the other 74 UK isolates are in Table S3 (supplementary materials).

6

1 **Table 3**

Isolates	Read length	No. of reads	Genome coverage	No. of reads spanning poly(C) tracts		Yield of reads spanning poly(C) tract		Gene
				1 st filter	2 nd filter	1 st filter	2 nd filter	
UK001 to UK024 (20 isolates)	54	5401318 (<i>1704184</i>)	71.4-fold (<i>22.5-fold</i>)	33.95 (<i>33.30</i>)	32.70 (<i>32.56</i>)	45.91% (<i>40.75%</i>)	44.22% (<i>39.73%</i>)	<i>fim2</i>
				18.50 (<i>15.40</i>)	15.10 (<i>14.14</i>)	24.93% (<i>18.72%</i>)	15.10 (<i>14.14</i>)	<i>fim3</i>
				80.15 (<i>33.26</i>)	79.40 (<i>32.98</i>)	109.70% (<i>21.94%</i>)	108.66% (<i>22.04%</i>)	<i>fimX</i>
UK025 to UK100 (76 isolates)	150	1255255 (<i>259298</i>)	46.1-fold (<i>9.5-fold</i>)	25.91 (<i>16.18</i>)	25.34 (<i>16.12</i>)	56.71% (<i>33.85%</i>)	55.45% (<i>33.80%</i>)	<i>fim2</i>
				20.32 (<i>14.25</i>)	17.87 (<i>13.83</i>)	43.56% (<i>26.83%</i>)	38.25% (<i>25.89%</i>)	<i>fim3</i>
				37.63 (<i>11.54</i>)	36.75 (<i>11.26</i>)	81.30% (<i>16.52%</i>)	79.46% (<i>16.47%</i>)	<i>fimX</i>

- 2 Table 3. Availability of Illumina reads spanning the fimbrial poly(C) tracts in *B. pertussis* isolates UK001 to UK024 and UK025 to UK100. All values
3 other than read length are arithmetic means per isolate (standard deviations *italicised*). Yields are expressed as percentages of genome coverage.
4 Genome sequences were not available for UK007, UK012, UK013 and UK019.

1 **Table 4**

Isolate	Serotype		Median poly(C) tract length	
	Observed	Predicted	<i>fim2</i>	<i>fim3</i>
UK001	Fim–	Fim2	14	9
UK003	Fim2,3	Fim2	16	6
UK005	Fim3	Fim2	17	13
UK010	Fim2,3	Fim2	17	10.5
UK011	Fim3	Fim2	14	13
UK015	Fim3	Fim–	9	10.5
UK030	Fim3	Fim2	16	10
UK043	Fim3	Fim2,3	15	14
UK051	Fim3	Fim2	12	13
UK061	Fim3	Fim–	11	8
UK064	Fim3	Fim2,3	14	14
UK072	Fim3	Fim2	12	13
UK081	Fim3	Fim2	16	9

2 Table 4. *B. pertussis* isolates with serotype by agglutination conflicting with serotype
3 predicted from *fim2* and *fim3* median poly(C) tract length. Observed, by agglutination;
4 Predicted, from median poly(C) tract lengths. Serotype nomenclature is as described by
5 Hallander *et al.* (2005). Discrepant fimbrial serotypes between those observed and predicted
6 are emboldened.

1 **Figure legends**

2 Figure 1. Bar charts: distribution of *fim2* and *fim3* promoter poly(C) tract lengths in
3 unassembled genome sequencing reads for 22 United Kingdom isolates of *B. pertussis*.
4 Scatterplots: surface expression of Fim2 and Fim3 in aliquots of the same cultures as were
5 used for the genome sequencing, determined by two-colour flow cytometry. The dominant
6 serotype by agglutination in each sample is stated in parentheses in the bar chart titles. Shaded
7 bars indicate poly(C) tract lengths that were previously reported to be transcription-permissive
8 (Chen *et al.*, 2010). *fim2* poly(C) tract lengths include the C residues that precede the T
9 residue that interrupts sequences analysed by Chen *et al.* The particles on the outside bounds
10 of the scatterplots are those with a very high fluorescence that exceeds the dynamic range of
11 the flow cytometer: they are more likely to be clumps of AlexaFluor-conjugated antibody than
12 to be bacteria.

13 Figure 2. Relative abundances of four fimbrial serotype permutations in individual cultures of
14 22 UK isolates of *B. pertussis* analysed by two-colour flow cytometry.

Figure 1 (part 1 of 5)

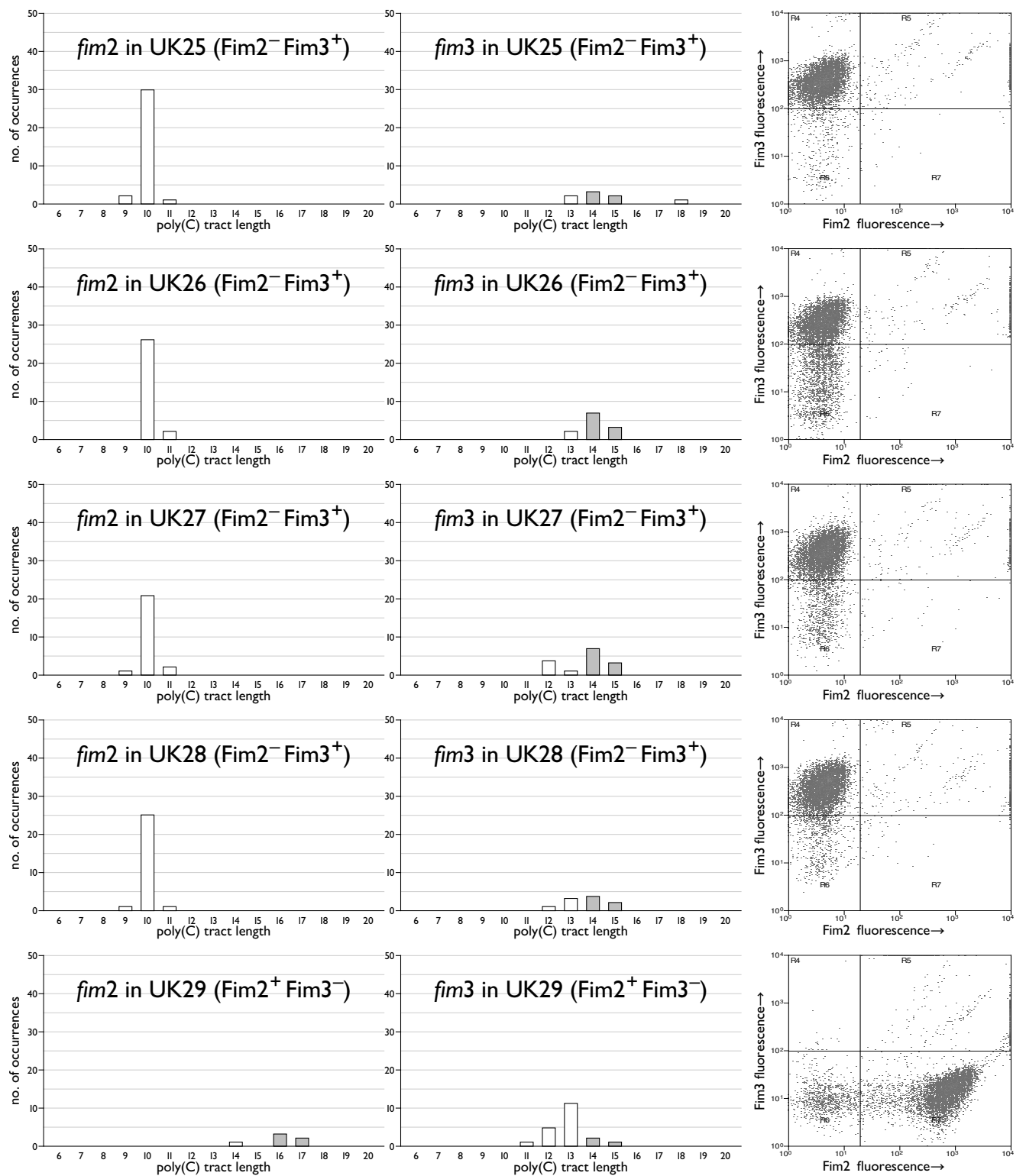


Figure 1 (part 2 of 5)

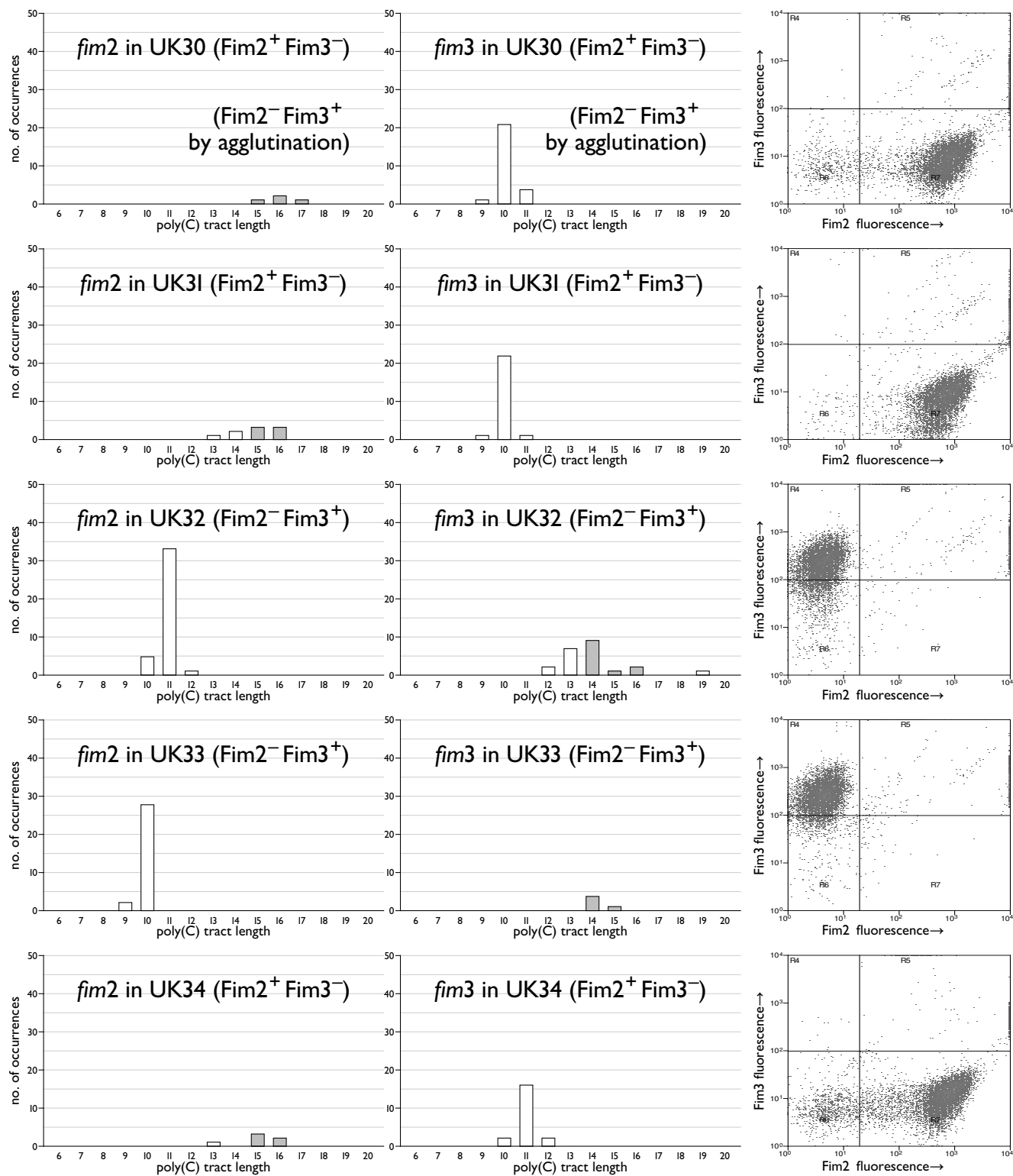


Figure 1 (part 3 of 5)

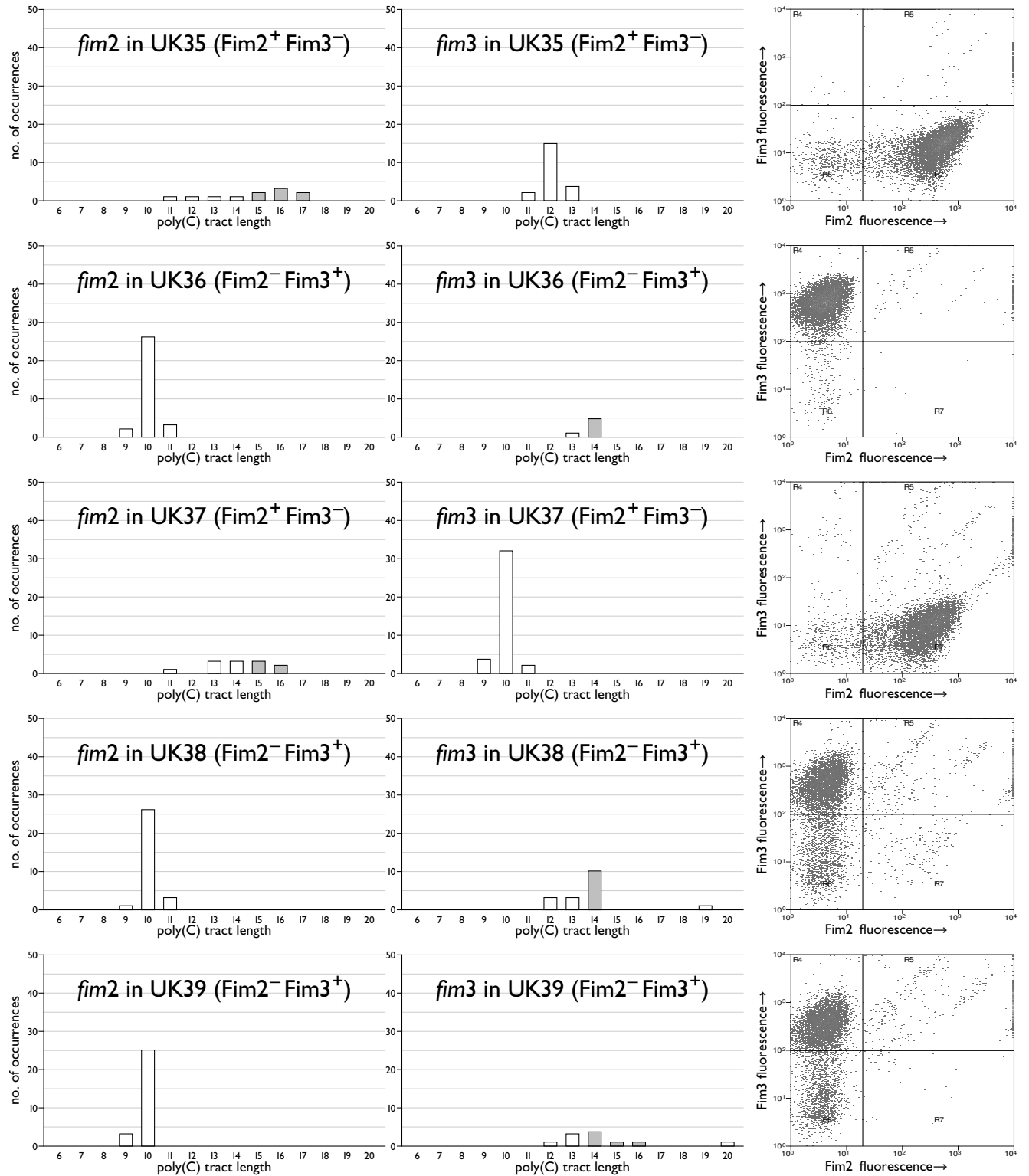


Figure 1 (part 4 of 5)

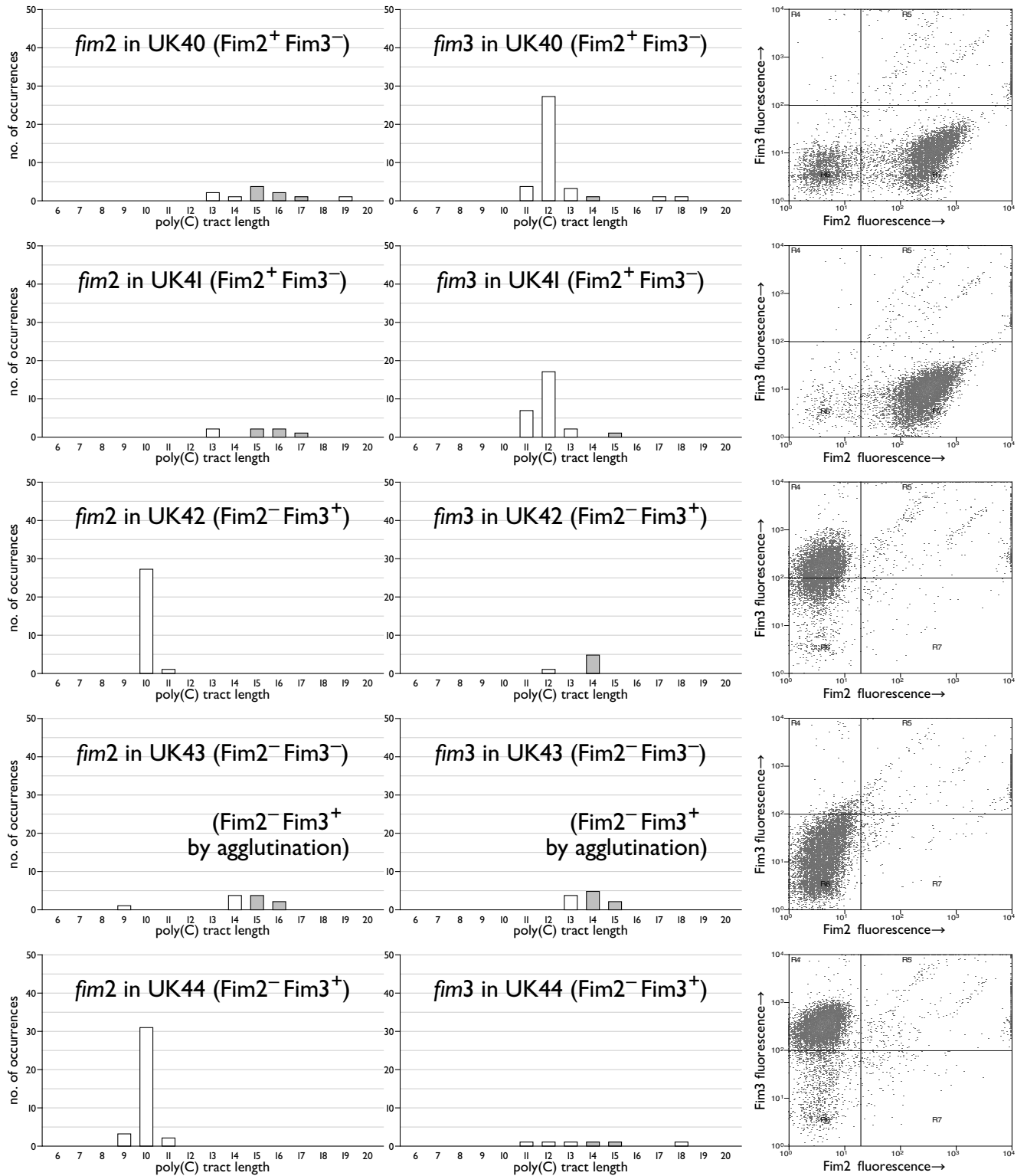


Figure 1 (part 5 of 5)

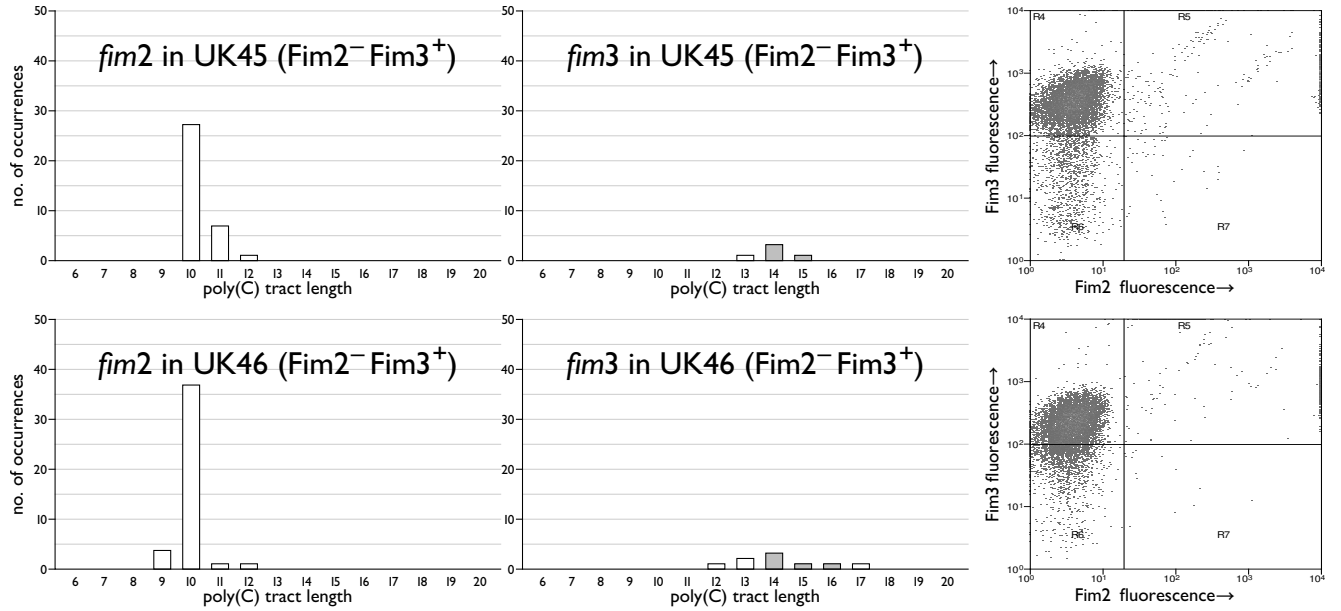
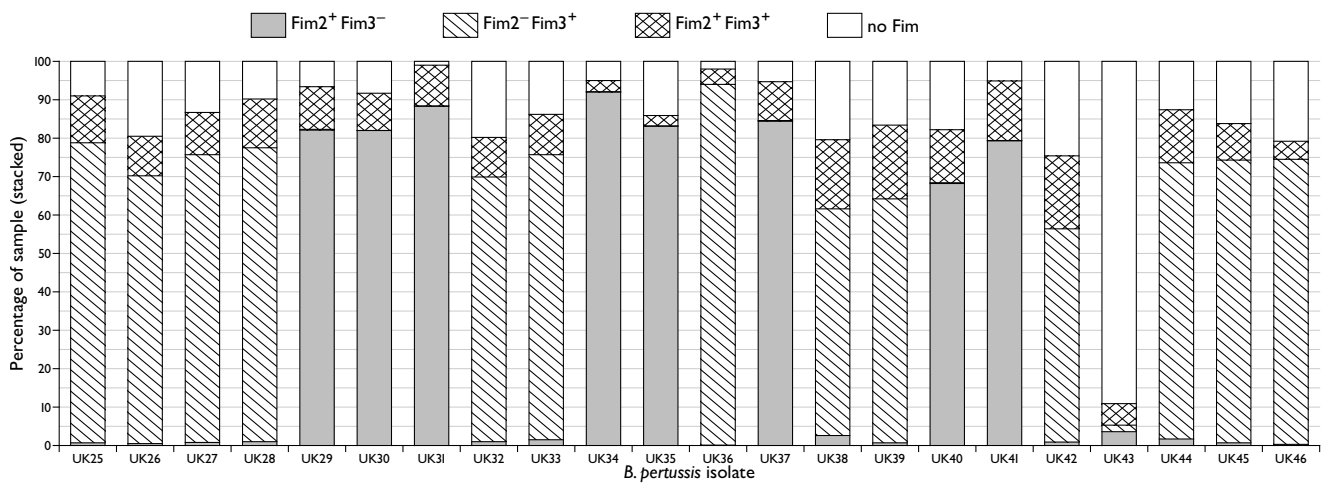


Figure 2



Supplementary tables and figures for
Plasticity of fimbrial genotype and serotype within populations of
Bordetella pertussis: analysis by paired flow cytometry and
genome sequencing

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Page 26	Table S3 (Supplementary to Table 2 in main text)

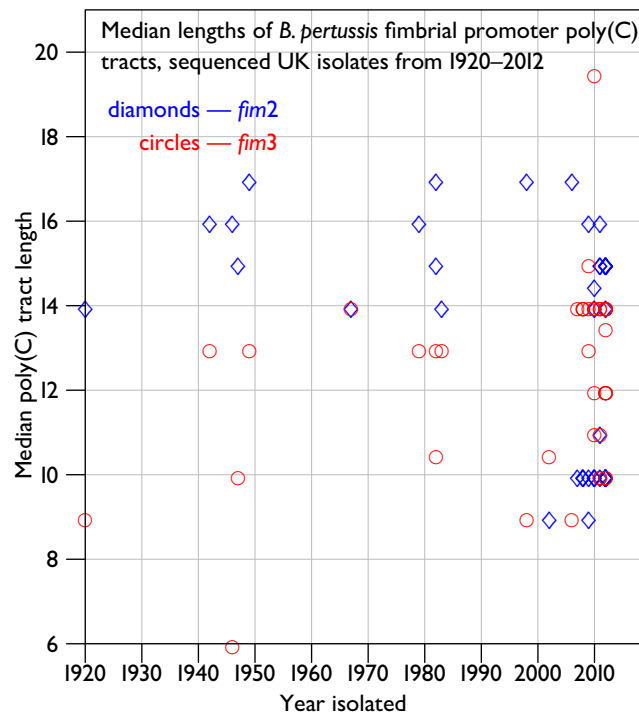


Figure S1 Median poly(C) tract lengths of *B. pertussis* isolates UK001 to UK048, isolated from 1920 to 2012.

Isolate	ENA identifier	File URL
UK001	ERS018345	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037420/ERR037420_1.fastq.gz
UK001	ERS018345	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037420/ERR037420_2.fastq.gz
UK002	ERS018346	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037421/ERR037421_1.fastq.gz
UK002	ERS018346	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037421/ERR037421_2.fastq.gz
UK003	ERS018347	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037422/ERR037422_1.fastq.gz
UK003	ERS018347	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037422/ERR037422_2.fastq.gz
UK004	ERS018348	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037396/ERR037396_1.fastq.gz
UK004	ERS018348	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037396/ERR037396_2.fastq.gz
UK005	ERS018349	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037400/ERR037400_1.fastq.gz
UK005	ERS018349	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037400/ERR037400_2.fastq.gz
UK006	ERS018350	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037401/ERR037401_1.fastq.gz
UK006	ERS018350	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037401/ERR037401_2.fastq.gz
UK008	ERS018351	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037402/ERR037402_1.fastq.gz
UK008	ERS018351	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037402/ERR037402_2.fastq.gz
UK009	ERS018352	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037403/ERR037403_1.fastq.gz
UK009	ERS018352	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037403/ERR037403_2.fastq.gz
UK010	ERS018353	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037404/ERR037404_1.fastq.gz
UK010	ERS018353	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037404/ERR037404_2.fastq.gz
UK011	ERS018354	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037405/ERR037405_1.fastq.gz
UK011	ERS018354	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037405/ERR037405_2.fastq.gz
UK014	ERS018355	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037406/ERR037406_1.fastq.gz
UK014	ERS018355	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037406/ERR037406_2.fastq.gz
UK015	ERS018356	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037407/ERR037407_1.fastq.gz
UK015	ERS018356	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037407/ERR037407_2.fastq.gz
UK016	ERS018357	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037397/ERR037397_1.fastq.gz
UK016	ERS018357	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037397/ERR037397_2.fastq.gz
UK017	ERS018358	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037398/ERR037398_1.fastq.gz
UK017	ERS018358	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037398/ERR037398_2.fastq.gz
UK018	ERS018359	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037399/ERR037399_1.fastq.gz
UK018	ERS018359	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037399/ERR037399_2.fastq.gz

Table S1 Unassembled genome sequence files for UK isolates used in this study (*continues on next page*).

Isolate	ENA identifier	File URL
UK020	ERS018360	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037408/ERR037408_1.fastq.gz
UK020	ERS018360	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037408/ERR037408_2.fastq.gz
UK021	ERS018361	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037411/ERR037411_1.fastq.gz
UK021	ERS018361	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037411/ERR037411_2.fastq.gz
UK022	ERS018362	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037412/ERR037412_1.fastq.gz
UK022	ERS018362	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037412/ERR037412_2.fastq.gz
UK023	ERS018363	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037413/ERR037413_1.fastq.gz
UK023	ERS018363	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037413/ERR037413_2.fastq.gz
UK024	ERS018364	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037414/ERR037414_1.fastq.gz
UK024	ERS018364	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037414/ERR037414_2.fastq.gz
UK049	ERS227750	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR304/ERR304770/ERR304770_1.fastq.gz
UK049	ERS227750	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR304/ERR304770/ERR304770_2.fastq.gz
UK050	ERS227751	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR304/ERR304771/ERR304771_1.fastq.gz
UK050	ERS227751	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR304/ERR304771/ERR304771_2.fastq.gz
UK051	ERS227752	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR304/ERR304772/ERR304772_1.fastq.gz
UK051	ERS227752	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR304/ERR304772/ERR304772_2.fastq.gz
UK052	ERS227753	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR304/ERR304773/ERR304773_1.fastq.gz
UK052	ERS227753	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR304/ERR304773/ERR304773_2.fastq.gz
UK053	ERS227754	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR304/ERR304774/ERR304774_1.fastq.gz
UK053	ERS227754	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR304/ERR304774/ERR304774_2.fastq.gz
UK054	ERS227755	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR304/ERR304775/ERR304775_1.fastq.gz
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UK055	ERS227756	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR304/ERR304776/ERR304776_1.fastq.gz
UK055	ERS227756	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR304/ERR304776/ERR304776_2.fastq.gz
UK056	ERS227757	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR304/ERR304777/ERR304777_1.fastq.gz
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UK057	ERS227758	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR304/ERR304778/ERR304778_1.fastq.gz
UK057	ERS227758	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR304/ERR304778/ERR304778_2.fastq.gz
UK058	ERS227759	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR304/ERR304779/ERR304779_1.fastq.gz
UK058	ERS227759	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR304/ERR304779/ERR304779_2.fastq.gz

Table SI continued Unassembled genome sequence files for UK isolates used in this study (*continues on next page*).

Isolate	ENA identifier	File URL
UK059	ERS227760	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR304/ERR304780/ERR304780_1.fastq.gz
UK059	ERS227760	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR304/ERR304780/ERR304780_2.fastq.gz
UK060	ERS227761	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR304/ERR304781/ERR304781_1.fastq.gz
UK060	ERS227761	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR304/ERR304781/ERR304781_2.fastq.gz
UK061	ERS227762	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR304/ERR304782/ERR304782_1.fastq.gz
UK061	ERS227762	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR304/ERR304782/ERR304782_2.fastq.gz
UK062	ERS227763	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR304/ERR304783/ERR304783_1.fastq.gz
UK062	ERS227763	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR304/ERR304783/ERR304783_2.fastq.gz
UK063	ERS227764	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR304/ERR304784/ERR304784_1.fastq.gz
UK063	ERS227764	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR304/ERR304784/ERR304784_2.fastq.gz
UK064	ERS227765	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR304/ERR304785/ERR304785_1.fastq.gz
UK064	ERS227765	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR304/ERR304785/ERR304785_2.fastq.gz
UK065	ERS227766	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR304/ERR304786/ERR304786_1.fastq.gz
UK065	ERS227766	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR304/ERR304786/ERR304786_2.fastq.gz
UK066	ERS227767	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR304/ERR304787/ERR304787_1.fastq.gz
UK066	ERS227767	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR304/ERR304787/ERR304787_2.fastq.gz
UK084	ERS227785	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR305/ERR305950/ERR305950_1.fastq.gz
UK084	ERS227785	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR305/ERR305950/ERR305950_2.fastq.gz
UK085	ERS227786	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR305/ERR305951/ERR305951_1.fastq.gz
UK085	ERS227786	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR305/ERR305951/ERR305951_2.fastq.gz
UK086	ERS227787	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR305/ERR305952/ERR305952_1.fastq.gz
UK086	ERS227787	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR305/ERR305952/ERR305952_2.fastq.gz
UK087	ERS227788	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR305/ERR305953/ERR305953_1.fastq.gz
UK087	ERS227788	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR305/ERR305953/ERR305953_2.fastq.gz
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Table SI continued Unassembled genome sequence files for UK isolates used in this study (*continues on next page*).

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UK091	ERS227792	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR305/ERR305957/ERR305957_1.fastq.gz
UK091	ERS227792	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR305/ERR305957/ERR305957_2.fastq.gz
UK092	ERS227793	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR305/ERR305958/ERR305958_1.fastq.gz
UK092	ERS227793	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR305/ERR305958/ERR305958_2.fastq.gz
UK093	ERS227794	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR305/ERR305959/ERR305959_1.fastq.gz
UK093	ERS227794	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR305/ERR305959/ERR305959_2.fastq.gz
UK094	ERS227795	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR305/ERR305960/ERR305960_1.fastq.gz
UK094	ERS227795	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR305/ERR305960/ERR305960_2.fastq.gz
UK095	ERS227796	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR305/ERR305961/ERR305961_1.fastq.gz
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UK098	ERS227799	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR305/ERR305964/ERR305964_1.fastq.gz
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UK099	ERS227800	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR305/ERR305965/ERR305965_1.fastq.gz
UK099	ERS227800	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR305/ERR305965/ERR305965_2.fastq.gz
UKI00	ERS227801	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR305/ERR305966/ERR305966_1.fastq.gz
UKI00	ERS227801	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR305/ERR305966/ERR305966_2.fastq.gz

Table S1 continued Unassembled genome sequence files for UK isolates used in this study.

Isolate	ENA identifier	File URL
(n/a)	ERS001360	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR007/ERR007653/ERR007653_1.fastq.gz
(n/a)	ERS001360	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR007/ERR007653/ERR007653_2.fastq.gz
Bp02-46	ERS018336	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037419/ERR037419_1.fastq.gz
Bp02-46	ERS018336	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037419/ERR037419_2.fastq.gz
Bp03-030	ERS018337	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037423/ERR037423_1.fastq.gz
Bp03-030	ERS018337	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037423/ERR037423_2.fastq.gz
Bp03-040	ERS018338	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037424/ERR037424_1.fastq.gz
Bp03-040	ERS018338	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037424/ERR037424_2.fastq.gz
Bp06-089	ERS018339	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037425/ERR037425_1.fastq.gz
Bp06-089	ERS018339	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037425/ERR037425_2.fastq.gz
Bp06-090	ERS018340	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037426/ERR037426_1.fastq.gz
Bp06-090	ERS018340	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037426/ERR037426_2.fastq.gz
Bp06-246	ERS018341	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037427/ERR037427_1.fastq.gz
Bp06-246	ERS018341	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037427/ERR037427_2.fastq.gz
Bp06-274	ERS018342	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037428/ERR037428_1.fastq.gz
Bp06-274	ERS018342	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037428/ERR037428_2.fastq.gz
Bp06-371	ERS018343	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037429/ERR037429_1.fastq.gz
Bp06-371	ERS018343	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037429/ERR037429_2.fastq.gz
Bp06-401	ERS018344	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037430/ERR037430_1.fastq.gz
Bp06-401	ERS018344	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037430/ERR037430_2.fastq.gz
bp4376	ERS018365	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037415/ERR037415_1.fastq.gz
bp4376	ERS018365	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037415/ERR037415_2.fastq.gz
bp4389	ERS018366	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037416/ERR037416_1.fastq.gz
bp4389	ERS018366	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037416/ERR037416_2.fastq.gz
bp4397	ERS018367	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037417/ERR037417_1.fastq.gz
bp4397	ERS018367	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037417/ERR037417_2.fastq.gz
bp4396	ERS018368	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037418/ERR037418_1.fastq.gz
bp4396	ERS018368	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037418/ERR037418_2.fastq.gz
bp4398	ERS018369	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037409/ERR037409_1.fastq.gz
bp4398	ERS018369	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037409/ERR037409_2.fastq.gz

Table S2 Unassembled genome sequence files for non-United Kingdom isolates referred to in this study (continues on next page).

Isolate	ENA identifier	File URL
bp4307	ERS018370	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037410/ERR037410_1.fastq.gz
bp4307	ERS018370	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR037/ERR037410/ERR037410_2.fastq.gz
(n/a)	ERS045992	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044160/ERR044160_1.fastq.gz
(n/a)	ERS045992	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044160/ERR044160_2.fastq.gz
(n/a)	ERS045996	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044164/ERR044164_1.fastq.gz
(n/a)	ERS045996	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044164/ERR044164_2.fastq.gz
(n/a)	ERS046000	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044168/ERR044168_1.fastq.gz
(n/a)	ERS046000	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044168/ERR044168_2.fastq.gz
(n/a)	ERS046004	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044172/ERR044172_1.fastq.gz
(n/a)	ERS046004	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044172/ERR044172_2.fastq.gz
(n/a)	ERS046008	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044176/ERR044176_1.fastq.gz
(n/a)	ERS046008	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044176/ERR044176_2.fastq.gz
(n/a)	ERS046012	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044180/ERR044180_1.fastq.gz
(n/a)	ERS046012	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044180/ERR044180_2.fastq.gz
(n/a)	ERS046016	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044184/ERR044184_1.fastq.gz
(n/a)	ERS046016	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044184/ERR044184_2.fastq.gz
(n/a)	ERS046020	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044188/ERR044188_1.fastq.gz
(n/a)	ERS046020	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044188/ERR044188_2.fastq.gz
(n/a)	ERS046024	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044192/ERR044192_1.fastq.gz
(n/a)	ERS046024	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044192/ERR044192_2.fastq.gz
(n/a)	ERS046028	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044196/ERR044196_1.fastq.gz
(n/a)	ERS046028	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044196/ERR044196_2.fastq.gz
(n/a)	ERS046032	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044200/ERR044200_1.fastq.gz
(n/a)	ERS046032	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044200/ERR044200_2.fastq.gz
(n/a)	ERS046036	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044204/ERR044204_1.fastq.gz
(n/a)	ERS046036	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044204/ERR044204_2.fastq.gz
(n/a)	ERS046040	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044208/ERR044208_1.fastq.gz
(n/a)	ERS046040	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044208/ERR044208_2.fastq.gz
(n/a)	ERS046044	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044212/ERR044212_1.fastq.gz
(n/a)	ERS046044	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044212/ERR044212_2.fastq.gz

Table S2 (continued) Unassembled genome sequence files for non-United Kingdom isolates referred to in this study (*continues on next page*).

Isolate	ENA identifier	File URL
(n/a)	ERS046048	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044216/ERR044216_1.fastq.gz
(n/a)	ERS046048	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044216/ERR044216_2.fastq.gz
(n/a)	ERS046052	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044220/ERR044220_1.fastq.gz
(n/a)	ERS046052	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044220/ERR044220_2.fastq.gz
(n/a)	ERS046056	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044224/ERR044224_1.fastq.gz
(n/a)	ERS046056	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044224/ERR044224_2.fastq.gz
(n/a)	ERS046060	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044228/ERR044228_1.fastq.gz
(n/a)	ERS046060	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044228/ERR044228_2.fastq.gz
(n/a)	ERS046064	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044232/ERR044232_1.fastq.gz
(n/a)	ERS046064	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044232/ERR044232_2.fastq.gz
(n/a)	ERS046068	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044236/ERR044236_1.fastq.gz
(n/a)	ERS046068	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044236/ERR044236_2.fastq.gz
(n/a)	ERS046072	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044240/ERR044240_1.fastq.gz
(n/a)	ERS046072	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044240/ERR044240_2.fastq.gz
(n/a)	ERS046076	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044244/ERR044244_1.fastq.gz
(n/a)	ERS046076	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044244/ERR044244_2.fastq.gz
(n/a)	ERS046080	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044248/ERR044248_1.fastq.gz
(n/a)	ERS046080	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044248/ERR044248_2.fastq.gz
(n/a)	ERS046084	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044252/ERR044252_2.fastq.gz
(n/a)	ERS046086	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044254/ERR044254_1.fastq.gz
(n/a)	ERS046086	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044254/ERR044254_2.fastq.gz
(n/a)	ERS046090	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044258/ERR044258_1.fastq.gz
(n/a)	ERS046090	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR044/ERR044258/ERR044258_2.fastq.gz
(n/a)	ERS046110	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR043/ERR043365/ERR043365_1.fastq.gz
(n/a)	ERS046110	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR043/ERR043365/ERR043365_2.fastq.gz
(n/a)	ERS046114	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR043/ERR043369/ERR043369_1.fastq.gz
(n/a)	ERS046114	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR043/ERR043369/ERR043369_2.fastq.gz
(n/a)	ERS046118	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR043/ERR043373/ERR043373_1.fastq.gz
(n/a)	ERS046118	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR043/ERR043373/ERR043373_2.fastq.gz
(n/a)	ERS046122	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR043/ERR043377/ERR043377_1.fastq.gz

Table S2 (continued) Unassembled genome sequence files for non-United Kingdom isolates referred to in this study (*continues on next page*).

Isolate	ENA identifier	File URL
(n/a)	ERS046122	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR043/ERR043377/ERR043377_2.fastq.gz
(n/a)	ERS053508	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR045/ERR045441/ERR045441_1.fastq.gz
(n/a)	ERS053508	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR045/ERR045441/ERR045441_2.fastq.gz
(n/a)	ERS053515	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR045/ERR045475/ERR045475_1.fastq.gz
(n/a)	ERS053515	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR045/ERR045475/ERR045475_2.fastq.gz
(n/a)	ERS053517	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR045/ERR045438/ERR045438_1.fastq.gz
(n/a)	ERS053517	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR045/ERR045438/ERR045438_2.fastq.gz
(n/a)	ERS053523	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR045/ERR045449/ERR045449_1.fastq.gz
(n/a)	ERS053523	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR045/ERR045449/ERR045449_2.fastq.gz
(n/a)	ERS053525	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR045/ERR045453/ERR045453_1.fastq.gz
(n/a)	ERS053525	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR045/ERR045453/ERR045453_2.fastq.gz
(n/a)	ERS053530	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR045/ERR045461/ERR045461_1.fastq.gz
(n/a)	ERS053530	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR045/ERR045461/ERR045461_2.fastq.gz
(n/a)	ERS053535	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR045/ERR045471/ERR045471_1.fastq.gz
(n/a)	ERS053535	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR045/ERR045471/ERR045471_2.fastq.gz
(n/a)	ERS053548	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR045/ERR045445/ERR045445_1.fastq.gz
(n/a)	ERS053548	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR045/ERR045445/ERR045445_2.fastq.gz
(n/a)	ERS053552	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR045/ERR045457/ERR045457_1.fastq.gz
(n/a)	ERS053552	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR045/ERR045457/ERR045457_2.fastq.gz
(n/a)	ERS053554	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR045/ERR045468/ERR045468_1.fastq.gz
(n/a)	ERS053554	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR045/ERR045468/ERR045468_2.fastq.gz
(n/a)	ERS054025	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046401/ERR046401_1.fastq.gz
(n/a)	ERS054025	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046401/ERR046401_2.fastq.gz
(n/a)	ERS054026	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046406/ERR046406_1.fastq.gz
(n/a)	ERS054026	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046406/ERR046406_2.fastq.gz
(n/a)	ERS054027	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046410/ERR046410_1.fastq.gz
(n/a)	ERS054027	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046410/ERR046410_2.fastq.gz
(n/a)	ERS054028	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046414/ERR046414_1.fastq.gz
(n/a)	ERS054028	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046414/ERR046414_2.fastq.gz
(n/a)	ERS054029	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046418/ERR046418_1.fastq.gz

Table S2 (continued) Unassembled genome sequence files for non-United Kingdom isolates referred to in this study (*continues on next page*).

Isolate	ENA identifier	File URL
(n/a)	ERS054029	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046418/ERR046418_2.fastq.gz
(n/a)	ERS054030	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046423/ERR046423_1.fastq.gz
(n/a)	ERS054030	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046423/ERR046423_2.fastq.gz
(n/a)	ERS054031	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046426/ERR046426_1.fastq.gz
(n/a)	ERS054031	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046426/ERR046426_2.fastq.gz
(n/a)	ERS054032	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046430/ERR046430_1.fastq.gz
(n/a)	ERS054032	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046430/ERR046430_2.fastq.gz
(n/a)	ERS054033	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046434/ERR046434_1.fastq.gz
(n/a)	ERS054033	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046434/ERR046434_2.fastq.gz
(n/a)	ERS054034	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046439/ERR046439_1.fastq.gz
(n/a)	ERS054034	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046439/ERR046439_2.fastq.gz
(n/a)	ERS054035	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046442/ERR046442_1.fastq.gz
(n/a)	ERS054035	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046442/ERR046442_2.fastq.gz
(n/a)	ERS054036	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046446/ERR046446_1.fastq.gz
(n/a)	ERS054036	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046446/ERR046446_2.fastq.gz
(n/a)	ERS054037	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046451/ERR046451_1.fastq.gz
(n/a)	ERS054037	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046451/ERR046451_2.fastq.gz
(n/a)	ERS054038	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046455/ERR046455_1.fastq.gz
(n/a)	ERS054038	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046455/ERR046455_2.fastq.gz
(n/a)	ERS054039	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046459/ERR046459_1.fastq.gz
(n/a)	ERS054039	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046459/ERR046459_2.fastq.gz
(n/a)	ERS054040	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046463/ERR046463_1.fastq.gz
(n/a)	ERS054040	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046463/ERR046463_2.fastq.gz
(n/a)	ERS054031	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046516/ERR046516_1.fastq.gz
(n/a)	ERS054031	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046516/ERR046516_2.fastq.gz
(n/a)	ERS005390	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR019/ERR019606/ERR019606_2.fastq.gz
(n/a)	ERS001339	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR007/ERR007510/ERR007510_1.fastq.gz
(n/a)	ERS001339	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR007/ERR007509/ERR007509_1.fastq.gz
(n/a)	ERS001339	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR007/ERR007510/ERR007510_2.fastq.gz
(n/a)	ERS001339	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR007/ERR007509/ERR007509_2.fastq.gz

Table S2 (continued) Unassembled genome sequence files for non-United Kingdom isolates referred to in this study (*continues on next page*).

Isolate	ENA identifier	File URL
(n/a)	ERS054027	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046473/ERR046473_1.fastq.gz
(n/a)	ERS054027	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046473/ERR046473_2.fastq.gz
(n/a)	ERS001338	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR007/ERR007506/ERR007506_2.fastq.gz
(n/a)	ERS005388	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR019/ERR019603/ERR019603_2.fastq.gz
(n/a)	ERS054026	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046469/ERR046469_1.fastq.gz
(n/a)	ERS001338	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR007/ERR007506/ERR007506_1.fastq.gz
(n/a)	ERS054028	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046477/ERR046477_1.fastq.gz
(n/a)	ERS054029	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046481/ERR046481_1.fastq.gz
(n/a)	ERS054026	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046469/ERR046469_2.fastq.gz
(n/a)	ERS054029	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046481/ERR046481_2.fastq.gz
(n/a)	ERS054028	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046477/ERR046477_2.fastq.gz
(n/a)	ERS054033	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046524/ERR046524_1.fastq.gz
(n/a)	ERS001361	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR007/ERR007648/ERR007648_1.fastq.gz
(n/a)	ERS054033	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046524/ERR046524_2.fastq.gz
(n/a)	ERS001361	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR007/ERR007648/ERR007648_2.fastq.gz
(n/a)	ERS054034	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046490/ERR046490_1.fastq.gz
(n/a)	ERS054034	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046490/ERR046490_2.fastq.gz
(n/a)	ERS054039	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046510/ERR046510_1.fastq.gz
(n/a)	ERS054036	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046497/ERR046497_1.fastq.gz
(n/a)	ERS054035	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046493/ERR046493_1.fastq.gz
(n/a)	ERS054039	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046510/ERR046510_2.fastq.gz
(n/a)	ERS054036	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046497/ERR046497_2.fastq.gz
(n/a)	ERS054035	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046493/ERR046493_2.fastq.gz
(n/a)	ERS054030	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046486/ERR046486_1.fastq.gz
(n/a)	ERS054040	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046514/ERR046514_1.fastq.gz
(n/a)	ERS054030	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046486/ERR046486_2.fastq.gz
(n/a)	ERS054032	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046520/ERR046520_1.fastq.gz
(n/a)	ERS054038	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046506/ERR046506_1.fastq.gz
(n/a)	ERS054040	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046514/ERR046514_2.fastq.gz
(n/a)	ERS054032	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046520/ERR046520_2.fastq.gz

Table S2 (continued) Unassembled genome sequence files for non-United Kingdom isolates referred to in this study (*continues on next page*).

Isolate	ENA identifier	File URL
(n/a)	ERS054038	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046506/ERR046506_2.fastq.gz
(n/a)	ERS054025	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046464/ERR046464_1.fastq.gz
(n/a)	ERS054025	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046464/ERR046464_2.fastq.gz
(n/a)	ERS054037	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046502/ERR046502_1.fastq.gz
(n/a)	ERS054037	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR046/ERR046502/ERR046502_2.fastq.gz
939980_2	ERS005382	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR019/ERR019594/ERR019594_1.fastq.gz
939980_2	ERS005382	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR019/ERR019594/ERR019594_2.fastq.gz
Bp02-027	ERS014547	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029992/ERR029992_1.fastq.gz
Bp02-027	ERS014547	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029992/ERR029992_2.fastq.gz
Bp02-045	ERS014550	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029984/ERR029984_1.fastq.gz
Bp02-045	ERS014550	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029984/ERR029984_2.fastq.gz
DKI2	ERS014537	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029331/ERR029331_1.fastq.gz
DKI2	ERS014537	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029331/ERR029331_2.fastq.gz
I326620_2	ERS005378	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR019/ERR019597/ERR019597_1.fastq.gz
I326620_2	ERS005378	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR019/ERR019597/ERR019597_2.fastq.gz
BP92001	ERS014612	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028888/ERR028888_1.fastq.gz
BP92001	ERS014612	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028888/ERR028888_2.fastq.gz
BP94025	ERS014615	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028891/ERR028891_1.fastq.gz
BP94025	ERS014615	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028891/ERR028891_2.fastq.gz
BII78	ERS014208	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029300/ERR029300_1.fastq.gz
BII78	ERS014208	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029300/ERR029300_2.fastq.gz
BPI82	ERS014509	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029313/ERR029313_1.fastq.gz
BPI82	ERS014509	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029313/ERR029313_2.fastq.gz
PT3783	ERS014611	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028885/ERR028885_1.fastq.gz
PT3783	ERS014611	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028885/ERR028885_2.fastq.gz
BP94035	ERS014616	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028892/ERR028892_1.fastq.gz
BP94035	ERS014616	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028892/ERR028892_2.fastq.gz
BPI28	ERS014511	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029315/ERR029315_1.fastq.gz
BPI28	ERS014511	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029315/ERR029315_2.fastq.gz
B0907	ERS014207	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029299/ERR029299_1.fastq.gz

Table S2 (continued) Unassembled genome sequence files for non-United Kingdom isolates referred to in this study (*continues on next page*).

Isolate	ENA identifier	File URL
B0907	ERS014207	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029299/ERR029299_2.fastq.gz
L599	ERS014594	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028869/ERR028869_1.fastq.gz
L599	ERS014594	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028869/ERR028869_2.fastq.gz
BII4I	ERS014210	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029302/ERR029302_1.fastq.gz
BII4I	ERS014210	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029302/ERR029302_2.fastq.gz
(n/a)	ERS005376	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR019/ERR019595/ERR019595_1.fastq.gz
(n/a)	ERS005376	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR019/ERR019595/ERR019595_2.fastq.gz
BP70	ERS014507	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029311/ERR029311_1.fastq.gz
BP70	ERS014507	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029311/ERR029311_2.fastq.gz
BP56	ERS014514	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029307/ERR029307_1.fastq.gz
BP56	ERS014514	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029307/ERR029307_2.fastq.gz
B0919	ERS014213	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029294/ERR029294_1.fastq.gz
B0919	ERS014213	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029294/ERR029294_2.fastq.gz
BII73	ERS014205	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029297/ERR029297_1.fastq.gz
BII73	ERS014205	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029297/ERR029297_2.fastq.gz
PRCB640	ERS014531	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029336/ERR029336_1.fastq.gz
PRCB640	ERS014531	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029336/ERR029336_2.fastq.gz
DKI3	ERS014538	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029332/ERR029332_1.fastq.gz
DKI3	ERS014538	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029332/ERR029332_2.fastq.gz
BP31	ERS014513	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029306/ERR029306_1.fastq.gz
BP31	ERS014513	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029306/ERR029306_2.fastq.gz
L570	ERS014586	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028849/ERR028849_1.fastq.gz
L570	ERS014586	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028849/ERR028849_2.fastq.gz
B0930	ERS014209	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029301/ERR029301_1.fastq.gz
B0930	ERS014209	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029301/ERR029301_2.fastq.gz
PRCB52I	ERS014529	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029334/ERR029334_1.fastq.gz
PRCB52I	ERS014529	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029334/ERR029334_2.fastq.gz
BII34	ERS014211	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029292/ERR029292_1.fastq.gz
BII34	ERS014211	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029292/ERR029292_2.fastq.gz
DK7	ERS014533	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029338/ERR029338_1.fastq.gz

Table S2 (continued) Unassembled genome sequence files for non-United Kingdom isolates referred to in this study (*continues on next page*).

Isolate	ENA identifier	File URL
DK7	ERS014533	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029338/ERR029338_2.fastq.gz
BP230	ERS014518	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029322/ERR029322_1.fastq.gz
BP230	ERS014518	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029322/ERR029322_2.fastq.gz
IS5840_2	ERS005370	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR019/ERR019587/ERR019587_1.fastq.gz
IS5840_2	ERS005370	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR019/ERR019587/ERR019587_2.fastq.gz
2035970_2	ERS005374	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR019/ERR019588/ERR019588_1.fastq.gz
2035970_2	ERS005374	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR019/ERR019588/ERR019588_2.fastq.gz
B0917	ERS014504	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029308/ERR029308_1.fastq.gz
B0917	ERS014504	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029308/ERR029308_2.fastq.gz
BP279	ERS014519	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029323/ERR029323_1.fastq.gz
BP279	ERS014519	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029323/ERR029323_2.fastq.gz
bp5776	ERS014560	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028822/ERR028822_1.fastq.gz
bp5776	ERS014560	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028822/ERR028822_2.fastq.gz
B0881	ERS018621	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031100/ERR031100_1.fastq.gz
B0881	ERS018621	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031100/ERR031100_2.fastq.gz
DK11	ERS014536	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029330/ERR029330_1.fastq.gz
DK11	ERS014536	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029330/ERR029330_2.fastq.gz
BPI20	ERS014510	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029314/ERR029314_1.fastq.gz
BPI20	ERS014510	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029314/ERR029314_2.fastq.gz
L590	ERS014592	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028867/ERR028867_1.fastq.gz
L590	ERS014592	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028867/ERR028867_2.fastq.gz
L582	ERS014590	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028865/ERR028865_1.fastq.gz
L582	ERS014590	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028865/ERR028865_2.fastq.gz
BPI01	ERS014526	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029319/ERR029319_1.fastq.gz
BPI01	ERS014526	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029319/ERR029319_2.fastq.gz
bp5766	ERS014557	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028830/ERR028830_1.fastq.gz
bp5766	ERS014557	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028830/ERR028830_2.fastq.gz
L695	ERS014607	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028883/ERR028883_1.fastq.gz
L695	ERS014607	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028883/ERR028883_2.fastq.gz
BP95003	ERS014617	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028893/ERR028893_1.fastq.gz

Table S2 (continued) Unassembled genome sequence files for non-United Kingdom isolates referred to in this study (*continues on next page*).

Isolate	ENA identifier	File URL
BP95003	ERS014617	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028893/ERR028893_2.fastq.gz
B0949	ERS018626	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031094/ERR031094_1.fastq.gz
B0949	ERS018626	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031094/ERR031094_2.fastq.gz
BPII9	ERS014515	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029317/ERR029317_1.fastq.gz
BPII9	ERS014515	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029317/ERR029317_2.fastq.gz
C900	ERS018609	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030013/ERR030013_1.fastq.gz
C900	ERS018609	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030013/ERR030013_2.fastq.gz
D492	ERS018612	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030005/ERR030005_1.fastq.gz
D492	ERS018612	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030005/ERR030005_2.fastq.gz
PRCB649	ERS014528	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029333/ERR029333_1.fastq.gz
PRCB649	ERS014528	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029333/ERR029333_2.fastq.gz
BI865	ERS018623	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031102/ERR031102_1.fastq.gz
BI865	ERS018623	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031102/ERR031102_2.fastq.gz
L709	ERS014609	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028874/ERR028874_1.fastq.gz
L709	ERS014609	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028874/ERR028874_2.fastq.gz
B0441	ERS018616	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031095/ERR031095_1.fastq.gz
B0441	ERS018616	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031095/ERR031095_2.fastq.gz
B0583	ERS018619	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031098/ERR031098_1.fastq.gz
B0583	ERS018619	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031098/ERR031098_2.fastq.gz
L468	ERS014579	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028853/ERR028853_1.fastq.gz
L468	ERS014579	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028853/ERR028853_2.fastq.gz
B0496	ERS018618	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031097/ERR031097_1.fastq.gz
B0496	ERS018618	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031097/ERR031097_2.fastq.gz
B0585	ERS018620	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031099/ERR031099_1.fastq.gz
B0585	ERS018620	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031099/ERR031099_2.fastq.gz
E495	ERS018625	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031093/ERR031093_1.fastq.gz
E495	ERS018625	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031093/ERR031093_2.fastq.gz
D898	ERS018613	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030006/ERR030006_1.fastq.gz
D898	ERS018613	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030006/ERR030006_2.fastq.gz
L580	ERS014588	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028863/ERR028863_1.fastq.gz

Table S2 (continued) Unassembled genome sequence files for non-United Kingdom isolates referred to in this study (*continues on next page*).

Isolate	ENA identifier	File URL
L580	ERS014588	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028863/ERR028863_2.fastq.gz
B385	ERS018599	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030028/ERR030028_1.fastq.gz
B385	ERS018599	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030028/ERR030028_2.fastq.gz
FR3207	ERS014572	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028844/ERR028844_1.fastq.gz
FR3207	ERS014572	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028844/ERR028844_2.fastq.gz
L453	ERS014576	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028850/ERR028850_1.fastq.gz
L453	ERS014576	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028850/ERR028850_2.fastq.gz
FR1139	ERS014570	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028842/ERR028842_1.fastq.gz
FR1139	ERS014570	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028842/ERR028842_2.fastq.gz
B1153	ERS014212	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029293/ERR029293_1.fastq.gz
B1153	ERS014212	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029293/ERR029293_2.fastq.gz
L452	ERS014575	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028846/ERR028846_1.fastq.gz
L452	ERS014575	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028846/ERR028846_2.fastq.gz
FR145	ERS014566	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028838/ERR028838_1.fastq.gz
FR145	ERS014566	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028838/ERR028838_2.fastq.gz
L595	ERS014593	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028868/ERR028868_1.fastq.gz
L595	ERS014593	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028868/ERR028868_2.fastq.gz
B1161	ERS014503	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029304/ERR029304_1.fastq.gz
B1161	ERS014503	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029304/ERR029304_2.fastq.gz
L573	ERS014587	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028859/ERR028859_1.fastq.gz
L573	ERS014587	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028859/ERR028859_2.fastq.gz
D300	ERS018611	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030015/ERR030015_1.fastq.gz
D300	ERS018611	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030015/ERR030015_2.fastq.gz
Bp02-044	ERS014549	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029983/ERR029983_1.fastq.gz
Bp02-044	ERS014549	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029983/ERR029983_2.fastq.gz
bp5822	ERS014552	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028825/ERR028825_1.fastq.gz
bp5822	ERS014552	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028825/ERR028825_2.fastq.gz
B0935	ERS014202	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029291/ERR029291_1.fastq.gz
B0935	ERS014202	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029291/ERR029291_2.fastq.gz
BP93001	ERS014613	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028889/ERR028889_1.fastq.gz

Table S2 (continued) Unassembled genome sequence files for non-United Kingdom isolates referred to in this study (*continues on next page*).

Isolate	ENA identifier	File URL
BP93001	ERS014613	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028889/ERR028889_2.fastq.gz
B0908	ERS014203	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029295/ERR029295_1.fastq.gz
B0908	ERS014203	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029295/ERR029295_2.fastq.gz
A214	ERS018624	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031092/ERR031092_1.fastq.gz
A214	ERS018624	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031092/ERR031092_2.fastq.gz
bp3092	ERS014553	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028826/ERR028826_1.fastq.gz
bp3092	ERS014553	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028826/ERR028826_2.fastq.gz
bp1773	ERS014556	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028829/ERR028829_1.fastq.gz
bp1773	ERS014556	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028829/ERR028829_2.fastq.gz
D909	ERS018614	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030007/ERR030007_1.fastq.gz
D909	ERS018614	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030007/ERR030007_2.fastq.gz
D075	ERS018610	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030014/ERR030014_1.fastq.gz
D075	ERS018610	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030014/ERR030014_2.fastq.gz
C670	ERS018607	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030011/ERR030011_1.fastq.gz
C670	ERS018607	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030011/ERR030011_2.fastq.gz
PRCB637	ERS014527	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029329/ERR029329_1.fastq.gz
PRCB637	ERS014527	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029329/ERR029329_2.fastq.gz
(n/a)	ERS005377	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR019/ERR019596/ERR019596_1.fastq.gz
(n/a)	ERS005377	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR019/ERR019596/ERR019596_2.fastq.gz
Bp02-035	ERS014548	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029982/ERR029982_1.fastq.gz
Bp02-035	ERS014548	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029982/ERR029982_2.fastq.gz
L481	ERS014580	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028854/ERR028854_1.fastq.gz
L481	ERS014580	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028854/ERR028854_2.fastq.gz
BP6901	ERS018579	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031087/ERR031087_1.fastq.gz
BP6901	ERS018579	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031087/ERR031087_2.fastq.gz
(n/a)	ERS005380	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR019/ERR019591/ERR019591_1.fastq.gz
(n/a)	ERS005380	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR019/ERR019591/ERR019591_2.fastq.gz
BP94016	ERS014614	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028890/ERR028890_1.fastq.gz
BP94016	ERS014614	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028890/ERR028890_2.fastq.gz
BP6	ERS014523	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029326/ERR029326_1.fastq.gz

Table S2 (continued) Unassembled genome sequence files for non-United Kingdom isolates referred to in this study (*continues on next page*).

Isolate	ENA identifier	File URL
BP6	ERS014523	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029326/ERR029326_2.fastq.gz
B0442	ERS018617	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031096/ERR031096_1.fastq.gz
B0442	ERS018617	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031096/ERR031096_2.fastq.gz
FRI003	ERS014569	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028841/ERR028841_1.fastq.gz
FRI003	ERS014569	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028841/ERR028841_2.fastq.gz
2146970_2	ERS005373	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR019/ERR019598/ERR019598_1.fastq.gz
2146970_2	ERS005373	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR019/ERR019598/ERR019598_2.fastq.gz
606770_2	ERS005386	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR019/ERR019592/ERR019592_1.fastq.gz
606770_2	ERS005386	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR019/ERR019592/ERR019592_2.fastq.gz
Bp02-016	ERS014545	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029990/ERR029990_1.fastq.gz
Bp02-016	ERS014545	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029990/ERR029990_2.fastq.gz
L619	ERS014600	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028876/ERR028876_1.fastq.gz
L619	ERS014600	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028876/ERR028876_2.fastq.gz
Bp02-019	ERS014546	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029991/ERR029991_1.fastq.gz
Bp02-019	ERS014546	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029991/ERR029991_2.fastq.gz
L617	ERS014599	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028872/ERR028872_1.fastq.gz
L617	ERS014599	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028872/ERR028872_2.fastq.gz
L581	ERS014589	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028864/ERR028864_1.fastq.gz
L581	ERS014589	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028864/ERR028864_2.fastq.gz
bp5783	ERS014559	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028832/ERR028832_1.fastq.gz
bp5783	ERS014559	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028832/ERR028832_2.fastq.gz
DK15	ERS014540	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029985/ERR029985_1.fastq.gz
DK15	ERS014540	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029985/ERR029985_2.fastq.gz
C455	ERS018604	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030008/ERR030008_1.fastq.gz
C455	ERS018604	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030008/ERR030008_2.fastq.gz
B0922	ERS014206	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029298/ERR029298_1.fastq.gz
B0922	ERS014206	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029298/ERR029298_2.fastq.gz
FR743	ERS014568	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028840/ERR028840_1.fastq.gz
FR743	ERS014568	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028840/ERR028840_2.fastq.gz
C477	ERS018606	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030010/ERR030010_1.fastq.gz

Table S2 (continued) Unassembled genome sequence files for non-United Kingdom isolates referred to in this study (*continues on next page*).

Isolate	ENA identifier	File URL
C477	ERS018606	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030010/ERR030010_2.fastq.gz
FR3713	ERS014574	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028836/ERR028836_1.fastq.gz
FR3713	ERS014574	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028836/ERR028836_2.fastq.gz
BPI05	ERS018573	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031081/ERR031081_1.fastq.gz
BPI05	ERS018573	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031081/ERR031081_2.fastq.gz
B490	ERS018601	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030019/ERR030019_1.fastq.gz
B490	ERS018601	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030019/ERR030019_2.fastq.gz
B316	ERS018598	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030027/ERR030027_1.fastq.gz
B316	ERS018598	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030027/ERR030027_2.fastq.gz
I758970_2	ERS005381	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR019/ERR019590/ERR019590_1.fastq.gz
I758970_2	ERS005381	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR019/ERR019590/ERR019590_2.fastq.gz
FR3407	ERS014573	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028835/ERR028835_1.fastq.gz
FR3407	ERS014573	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028835/ERR028835_2.fastq.gz
bp5640	ERS014551	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028821/ERR028821_1.fastq.gz
bp5640	ERS014551	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028821/ERR028821_2.fastq.gz
L711	ERS014610	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028875/ERR028875_1.fastq.gz
L711	ERS014610	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028875/ERR028875_2.fastq.gz
L655	ERS014605	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028881/ERR028881_1.fastq.gz
L655	ERS014605	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028881/ERR028881_2.fastq.gz
C467	ERS018605	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030009/ERR030009_1.fastq.gz
C467	ERS018605	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030009/ERR030009_2.fastq.gz
BI59	ERS017001	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030021/ERR030021_1.fastq.gz
BI59	ERS017001	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030021/ERR030021_2.fastq.gz
bp6632	ERS014562	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028824/ERR028824_1.fastq.gz
bp6632	ERS014562	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028824/ERR028824_2.fastq.gz
DK14	ERS014539	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029981/ERR029981_1.fastq.gz
DK14	ERS014539	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029981/ERR029981_2.fastq.gz
C855	ERS018608	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030012/ERR030012_1.fastq.gz
C855	ERS018608	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030012/ERR030012_2.fastq.gz
L518	ERS014584	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028847/ERR028847_1.fastq.gz

Table S2 (continued) Unassembled genome sequence files for non-United Kingdom isolates referred to in this study (*continues on next page*).

Isolate	ENA identifier	File URL
L518	ERS014584	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028847/ERR028847_2.fastq.gz
Bp02-012	ERS014544	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029989/ERR029989_1.fastq.gz
Bp02-012	ERS014544	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029989/ERR029989_2.fastq.gz
BP601	ERS018576	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031084/ERR031084_1.fastq.gz
BP601	ERS018576	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031084/ERR031084_2.fastq.gz
bp890	ERS014554	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028827/ERR028827_1.fastq.gz
bp890	ERS014554	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028827/ERR028827_2.fastq.gz
BP96004	ERS014619	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028895/ERR028895_1.fastq.gz
BP96004	ERS014619	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028895/ERR028895_2.fastq.gz
B0403	ERS018615	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031091/ERR031091_1.fastq.gz
B0403	ERS018615	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031091/ERR031091_2.fastq.gz
B0887	ERS018622	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031101/ERR031101_1.fastq.gz
B0887	ERS018622	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031101/ERR031101_2.fastq.gz
BP95004	ERS014618	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028894/ERR028894_1.fastq.gz
BP95004	ERS014618	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028894/ERR028894_2.fastq.gz
BPI06	ERS018574	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031082/ERR031082_1.fastq.gz
BPI06	ERS018574	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031082/ERR031082_2.fastq.gz
BPI21	ERS014512	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029305/ERR029305_1.fastq.gz
BPI21	ERS014512	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029305/ERR029305_2.fastq.gz
FR3080	ERS014571	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028843/ERR028843_1.fastq.gz
FR3080	ERS014571	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028843/ERR028843_2.fastq.gz
PRCB450	ERS014530	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029335/ERR029335_1.fastq.gz
PRCB450	ERS014530	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029335/ERR029335_2.fastq.gz
BP317	ERS014521	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029325/ERR029325_1.fastq.gz
BP317	ERS014521	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029325/ERR029325_2.fastq.gz
BPI43	ERS014516	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029320/ERR029320_1.fastq.gz
BPI43	ERS014516	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029320/ERR029320_2.fastq.gz
bp4834	ERS014555	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028828/ERR028828_1.fastq.gz
bp4834	ERS014555	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028828/ERR028828_2.fastq.gz
B0913	ERS014506	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029310/ERR029310_1.fastq.gz

Table S2 (continued) Unassembled genome sequence files for non-United Kingdom isolates referred to in this study (*continues on next page*).

Isolate	ENA identifier	File URL
B0913	ERSO14506	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029310/ERR029310_2.fastq.gz
Bp02-001	ERSO14541	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029986/ERR029986_1.fastq.gz
Bp02-001	ERSO14541	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029986/ERR029986_2.fastq.gz
B225	ERSO18597	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030026/ERR030026_1.fastq.gz
B225	ERSO18597	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030026/ERR030026_2.fastq.gz
BP824	ERSO18578	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031086/ERR031086_1.fastq.gz
BP824	ERSO18578	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031086/ERR031086_2.fastq.gz
L508	ERSO14583	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028857/ERR028857_1.fastq.gz
L508	ERSO14583	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028857/ERR028857_2.fastq.gz
B215	ERSO18596	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030025/ERR030025_1.fastq.gz
B215	ERSO18596	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030025/ERR030025_2.fastq.gz
Bp02-005	ERSO14543	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029988/ERR029988_1.fastq.gz
Bp02-005	ERSO14543	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029988/ERR029988_2.fastq.gz
B1137	ERSO14505	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029309/ERR029309_1.fastq.gz
B1137	ERSO14505	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029309/ERR029309_2.fastq.gz
DK8	ERSO14534	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029339/ERR029339_1.fastq.gz
DK8	ERSO14534	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029339/ERR029339_2.fastq.gz
CI39	ERSO18603	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030004/ERR030004_1.fastq.gz
CI39	ERSO18603	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030004/ERR030004_2.fastq.gz
B200	ERSO18593	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030022/ERR030022_1.fastq.gz
B200	ERSO18593	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030022/ERR030022_2.fastq.gz
L491	ERSO14581	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028855/ERR028855_1.fastq.gz
L491	ERSO14581	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028855/ERR028855_2.fastq.gz
L584	ERSO14591	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028866/ERR028866_1.fastq.gz
L584	ERSO14591	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028866/ERR028866_2.fastq.gz
Bp02-002	ERSO14542	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029987/ERR029987_1.fastq.gz
Bp02-002	ERSO14542	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029987/ERR029987_2.fastq.gz
L535	ERSO14585	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028848/ERR028848_1.fastq.gz
L535	ERSO14585	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028848/ERR028848_2.fastq.gz
DK6	ERSO14532	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029337/ERR029337_1.fastq.gz

Table S2 (continued) Unassembled genome sequence files for non-United Kingdom isolates referred to in this study (*continues on next page*).

Isolate	ENA identifier	File URL
DK6	ERS014532	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029337/ERR029337_2.fastq.gz
I26690_2	ERS005379	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR019/ERR019593/ERR019593_1.fastq.gz
I26690_2	ERS005379	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR019/ERR019593/ERR019593_2.fastq.gz
BP706	ERS018577	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031085/ERR031085_1.fastq.gz
BP706	ERS018577	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031085/ERR031085_2.fastq.gz
bpl535	ERS014561	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028823/ERR028823_1.fastq.gz
bpl535	ERS014561	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028823/ERR028823_2.fastq.gz
L708	ERS014608	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028873/ERR028873_1.fastq.gz
L708	ERS014608	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028873/ERR028873_2.fastq.gz
BP7	ERS014525	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029318/ERR029318_1.fastq.gz
BP7	ERS014525	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029318/ERR029318_2.fastq.gz
KKKI33	ERS018591	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030002/ERR030002_1.fastq.gz
KKKI33	ERS018591	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030002/ERR030002_2.fastq.gz
B453	ERS018600	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030018/ERR030018_1.fastq.gz
B453	ERS018600	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030018/ERR030018_2.fastq.gz
IS8205_2	ERS005372	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR019/ERR019589/ERR019589_1.fastq.gz
IS8205_2	ERS005372	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR019/ERR019589/ERR019589_2.fastq.gz
B733	ERS018602	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030020/ERR030020_1.fastq.gz
B733	ERS018602	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030020/ERR030020_2.fastq.gz
Cz	ERS014564	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028834/ERR028834_1.fastq.gz
Cz	ERS014564	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028834/ERR028834_2.fastq.gz
BP22	ERS014508	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029312/ERR029312_1.fastq.gz
BP22	ERS014508	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029312/ERR029312_2.fastq.gz
BP312	ERS018575	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031083/ERR031083_1.fastq.gz
BP312	ERS018575	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031083/ERR031083_2.fastq.gz
L467	ERS014578	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028852/ERR028852_1.fastq.gz
L467	ERS014578	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028852/ERR028852_2.fastq.gz
PRCB406	ERS018590	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030001/ERR030001_1.fastq.gz
PRCB406	ERS018590	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030001/ERR030001_2.fastq.gz
A818	ERS018592	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030017/ERR030017_1.fastq.gz

Table S2 (continued) Unassembled genome sequence files for non-United Kingdom isolates referred to in this study (continues on next page).

Isolate	ENA identifier	File URL
A818	ERS018592	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030017/ERR030017_2.fastq.gz
B203	ERS018595	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030024/ERR030024_1.fastq.gz
B203	ERS018595	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030024/ERR030024_2.fastq.gz
BP2	ERS014524	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029327/ERR029327_1.fastq.gz
BP2	ERS014524	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029327/ERR029327_2.fastq.gz
Hav	ERS014565	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028837/ERR028837_1.fastq.gz
Hav	ERS014565	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028837/ERR028837_2.fastq.gz
bp6334	ERS014558	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028831/ERR028831_1.fastq.gz
bp6334	ERS014558	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028831/ERR028831_2.fastq.gz
B202	ERS018594	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030023/ERR030023_1.fastq.gz
B202	ERS018594	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030023/ERR030023_2.fastq.gz
DK10	ERS014535	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029340/ERR029340_1.fastq.gz
DK10	ERS014535	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029340/ERR029340_2.fastq.gz
BP310	ERS014520	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029324/ERR029324_1.fastq.gz
BP310	ERS014520	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029324/ERR029324_2.fastq.gz
L646	ERS014603	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028879/ERR028879_1.fastq.gz
L646	ERS014603	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028879/ERR028879_2.fastq.gz
B0934	ERS014204	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029296/ERR029296_1.fastq.gz
B0934	ERS014204	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029296/ERR029296_2.fastq.gz
PRCB352	ERS018589	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030000/ERR030000_1.fastq.gz
PRCB352	ERS018589	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR030/ERR030000/ERR030000_2.fastq.gz
L649	ERS014604	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028880/ERR028880_1.fastq.gz
L649	ERS014604	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028880/ERR028880_2.fastq.gz
L687	ERS014606	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028882/ERR028882_1.fastq.gz
L687	ERS014606	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028882/ERR028882_2.fastq.gz
PRCBI79	ERS018587	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029998/ERR029998_1.fastq.gz
PRCBI79	ERS018587	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029998/ERR029998_2.fastq.gz
KKKI277	ERS018581	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031089/ERR031089_1.fastq.gz
KKKI277	ERS018581	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031089/ERR031089_2.fastq.gz
L454	ERS014577	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028851/ERR028851_1.fastq.gz

Table S2 (continued) Unassembled genome sequence files for non-United Kingdom isolates referred to in this study (*continues on next page*).

Isolate	ENA identifier	File URL
L454	ERS014577	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028851/ERR028851_2.fastq.gz
BP227	ERS014517	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029321/ERR029321_1.fastq.gz
BP227	ERS014517	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029321/ERR029321_2.fastq.gz
PRCBII8	ERS018586	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029997/ERR029997_1.fastq.gz
PRCBII8	ERS018586	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029997/ERR029997_2.fastq.gz
L610	ERS014597	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028861/ERR028861_1.fastq.gz
L610	ERS014597	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028861/ERR028861_2.fastq.gz
L504	ERS014582	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028856/ERR028856_1.fastq.gz
L504	ERS014582	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028856/ERR028856_2.fastq.gz
PRCB272	ERS018588	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029999/ERR029999_1.fastq.gz
PRCB272	ERS018588	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029999/ERR029999_2.fastq.gz
FR287	ERS014567	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028839/ERR028839_1.fastq.gz
FR287	ERS014567	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028839/ERR028839_2.fastq.gz
L604	ERS014595	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028870/ERR028870_1.fastq.gz
L604	ERS014595	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028870/ERR028870_2.fastq.gz
PRCB2	ERS018585	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029996/ERR029996_1.fastq.gz
PRCB2	ERS018585	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029996/ERR029996_2.fastq.gz
I9773	ERS018583	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029994/ERR029994_1.fastq.gz
I9773	ERS018583	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029994/ERR029994_2.fastq.gz
PRCB7H	ERS018584	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029995/ERR029995_1.fastq.gz
PRCB7H	ERS018584	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR029/ERR029995/ERR029995_2.fastq.gz
L638	ERS014601	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028877/ERR028877_1.fastq.gz
L638	ERS014601	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028877/ERR028877_2.fastq.gz
KKK22	ERS018580	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031088/ERR031088_1.fastq.gz
KKK22	ERS018580	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR031/ERR031088/ERR031088_2.fastq.gz
L644	ERS014602	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028878/ERR028878_1.fastq.gz
L644	ERS014602	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028878/ERR028878_2.fastq.gz
L609	ERS014596	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028860/ERR028860_1.fastq.gz
L609	ERS014596	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028860/ERR028860_2.fastq.gz
(n/a)	ERS005384	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR019/ERR019600/ERR019600_1.fastq.gz

Table S2 (continued) Unassembled genome sequence files for non-United Kingdom isolates referred to in this study (*continues on next page*).

Isolate	ENA identifier	File URL
(n/a)	ERS005384	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR019/ERR019600/ERR019600_2.fastq.gz
L614	ERS014598	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028862/ERR028862_1.fastq.gz
L614	ERS014598	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028862/ERR028862_2.fastq.gz
3690970_2	ERS005383	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR019/ERR019601/ERR019601_1.fastq.gz
3690970_2	ERS005383	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR019/ERR019601/ERR019601_2.fastq.gz
(n/a)	ERS005387	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR019/ERR019604/ERR019604_1.fastq.gz
(n/a)	ERS005387	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR019/ERR019604/ERR019604_2.fastq.gz
(n/a)	ERS005389	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR019/ERR019605/ERR019605_1.fastq.gz
(n/a)	ERS005389	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR019/ERR019605/ERR019605_2.fastq.gz
273980_2	ERS005385	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR019/ERR019602/ERR019602_1.fastq.gz
273980_2	ERS005385	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR019/ERR019602/ERR019602_2.fastq.gz
BP96006	ERS014620	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028886/ERR028886_1.fastq.gz
BP96006	ERS014620	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028886/ERR028886_2.fastq.gz
BP96013	ERS014621	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028887/ERR028887_1.fastq.gz
BP96013	ERS014621	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR028/ERR028887/ERR028887_2.fastq.gz
PT2019_2	ERS005405	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR019/ERR019585/ERR019585_1.fastq.gz
PT2019_2	ERS005405	ftp://ftp.sra.ebi.ac.uk/voll/fastq/ERR019/ERR019585/ERR019585_2.fastq.gz

Table S2 (continued) Unassembled genome sequence files for non–United Kingdom isolates referred to in this study.

Isolate	Year of isolation	Serotype by agglutination	ENA identifier	Fim2				Fim3			
				No. of reads	Median poly(C) tract length	Range of poly(C) tract lengths	Median fluorescence	No. of reads	Median poly(C) tract length	Range of poly(C) tract lengths	Median fluorescence
UK001	1920	I	ERS018345	80	14	4 (12 to 15)	nd	46	9	4 (7 to 10)	nd
UK002	1942	I,2	ERS018346	30	16	10 (8 to 17)	nd	22	13	5 (12 to 16)	nd
UK003	1946	I,2,3	ERS018347	18	16	3 (15 to 17)	nd	0	nd	nd	nd
UK004	1947	I,2	ERS018348	22	15.5	2 (15 to 16)	nd	35	10	3 (9 to 11)	nd
UK005	1949	I,3	ERS018349	11	17	7 (14 to 20)	nd	13	13	9 (11 to 19)	nd
UK006	1967	I,2,3	ERS018350	5	14	2 (14 to 15)	nd	5	14	7 (13 to 19)	nd
UK008	1979	I,2	ERS018349	6	16	4 (15 to 18)	nd	19	13	9 (11 to 19)	nd
UK009	1982	I,2	ERS018350	6	15	5 (12 to 16)	nd	4	12.5	5 (9 to 13)	nd
UK010	1982	I,2,3	ERS018353	1	17	1 (17 to 17)	nd	1	15	1 (15 to 15)	nd
UK011	1983	I,3	ERS018354	12	14	8 (11 to 18)	nd	2	16.5	8 (13 to 20)	nd
UK014	1998	I,2	ERS018355	16	17	10 (8 to 17)	nd	23	9	1 (9 to 9)	nd
UK015	2002	I,3	ERS018356	38	9	1 (9 to 9)	nd	1	14	1 (14 to 14)	nd
UK016	2006	I,2	ERS018357	20	17	3 (15 to 17)	nd	27	9	4 (7 to 10)	nd
UK017	2007	I,3	ERS018358	66	10	6 (6 to 11)	nd	1	14	1 (14 to 14)	nd
UK018	2010	I,3	ERS018359	37	10	2 (9 to 10)	nd	7	19	7 (14 to 20)	nd
UK020	2009	I,3	ERS018360	83	9	2 (9 to 10)	nd	5	15	4 (13 to 16)	nd
UK021	2010	I,2	ERS018361	5	14	4 (13 to 16)	nd	34	12	3 (11 to 13)	nd
UK022	2010	I,2	ERS018362	7	15	5 (12 to 16)	nd	35	11	3 (10 to 12)	nd
UK023	2010	I,3	ERS018363	92	10	3 (9 to 11)	nd	11	15	9 (12 to 20)	nd
UK024	2010	I,3	ERS018364	99	10	3 (9 to 11)	nd	11	15	8 (13 to 20)	nd
UK049	2012	I,3	ERS227750	51	10	3 (9 to 11)	nd	2	15	3 (14 to 16)	nd
UK050	2012	I,2	ERS227751	15	16	6 (13 to 18)	nd	21	10	3 (9 to 11)	nd
UK051	2011	I,3	ERS227752	48	12	3 (11 to 13)	nd	6	13	3 (12 to 14)	nd
UK052	2011	I,2	ERS227753	6	13.5	6 (10 to 15)	nd	22	11	3 (10 to 12)	nd
UK053	2011	I,3	ERS227754	37	10	3 (9 to 11)	nd	5	14	3 (13 to 15)	nd
UK054	2011	I,3	ERS227755	18	10	2 (10 to 11)	nd	3	14	2 (13 to 14)	nd
UK055	2012	I,2	ERS227756	8	16.5	7 (12 to 18)	nd	31	10	3 (9 to 11)	nd
UK056	2012	I,2	ERS227757	15	14	4 (12 to 15)	nd	15	10	2 (9 to 10)	nd
UK057	2012	I,3	ERS227758	48	9	2 (8 to 9)	nd	5	15	13 (7 to 19)	nd
UK058	2012	I,2	ERS227759	12	15	7 (12 to 18)	nd	14	12	3 (11 to 13)	nd
UK059	2012	I,3	ERS227760	63	10	3 (9 to 11)	nd	12	14.5	8 (12 to 19)	nd
UK060	2012	I,2	ERS227761	9	16	4 (13 to 16)	nd	18	12	4 (10 to 13)	nd
UK061	2012	I,3	ERS227762	36	11	3 (10 to 12)	nd	2	11	9 (7 to 15)	nd
UK062	2012	I,2	ERS227763	9	15	5 (13 to 17)	nd	19	11	4 (10 to 13)	nd
UK063	2012	I,3	ERS227764	45	10	2 (9 to 10)	nd	5	13	5 (11 to 15)	nd
UK064	2012	I,3	ERS227765	27	14	8 (9 to 16)	nd	9	15	7 (13 to 19)	nd
UK065	2012	I,3	ERS227766	62	10	4 (8 to 11)	nd	12	14	8 (13 to 20)	nd
UK066	2012	I,3	ERS227767	50	9	2 (9 to 10)	nd	14	13	6 (11 to 16)	nd
UK067	2012	I,2	ERS227768	9	15	3 (14 to 16)	nd	33	12	4 (10 to 13)	nd
UK068	2012	I,2,3	ERS227769	13	15	8 (9 to 16)	nd	11	14	7 (14 to 20)	nd
UK069	2012	I,3	ERS227770	41	11	4 (9 to 12)	nd	11	13	2 (13 to 14)	nd
UK070	2012	I,3	ERS227771	42	11	4 (10 to 13)	nd	16	13	7 (9 to 15)	nd
UK071	2012	I,2	ERS227772	7	14	8 (9 to 16)	nd	12	13	3 (12 to 14)	nd
UK072	2012	I,3	ERS227773	39	12	4 (10 to 13)	nd	14	13.5	3 (13 to 15)	nd
UK073	2012	I,2	ERS227774	14	15	7 (12 to 18)	nd	19	12	3 (11 to 13)	nd
UK074	2012	I,3	ERS227775	45	9	3 (8 to 10)	nd	11	14	9 (11 to 19)	nd
UK075	2012	I,3	ERS227776	44	10	2 (9 to 10)	nd	11	14	7 (11 to 17)	nd
UK076	2012	I,2	ERS227777	10	15	6 (12 to 17)	nd	53	12	12 (7 to 18)	nd
UK077	2012	I,3	ERS227778	53	10	3 (9 to 11)	nd	15	14	3 (13 to 15)	nd
UK078	2012	I,2	ERS227779	12	15.5	5 (12 to 16)	nd	38	12	11 (10 to 20)	nd
UK079	2012	I,2	ERS227780	14	14.5	5 (12 to 16)	nd	29	12	4 (11 to 14)	nd
UK080	2012	I,3	ERS227781	33	9	1 (9 to 9)	nd	8	14.5	10 (6 to 15)	nd
UK081	2012	I,3	ERS227782	13	16	5 (13 to 17)	nd	81	9	3 (8 to 10)	nd
UK082	2012	I,3	ERS227783	41	10	3 (9 to 11)	nd	11	14	10 (6 to 15)	nd
UK083	2012	I,2	ERS227784	9	16	3 (15 to 17)	nd	51	11	4 (9 to 12)	nd
UK084	2012	I,2	ERS227785	7	15	4 (13 to 16)	nd	34	10	3 (9 to 11)	nd
UK085	2012	I,3	ERS227786	44	10	5 (7 to 11)	nd	9	14	8 (13 to 20)	nd
UK086	2012	I,2	ERS227787	9	16	11 (7 to 17)	nd	26	10	3 (9 to 11)	nd
UK087	2012	I,3	ERS227788	37	9	3 (8 to 10)	nd	12	14	4 (12 to 15)	nd
UK088	2012	I,3	ERS227789	29	9	3 (8 to 10)	nd	5	14	3 (13 to 15)	nd
UK089	2012	I,3	ERS227790	35	10	4 (9 to 12)	nd	16	14	4 (12 to 15)	nd
UK090	2012	I,3	ERS227791	27	10	3 (9 to 11)	nd	12	14	5 (13 to 17)	nd
UK091	2012	I,3	ERS227792	29	11	3 (10 to 12)	nd	5	15	4 (13 to 16)	nd
UK092	2012	I,2	ERS227793	11	15	11 (6 to 16)	nd	41	11	4 (10 to 13)	nd
UK093	2012	I,3	ERS227794	33	10	3 (9 to 11)	nd	10	15	10 (7 to 16)	nd
UK094	2012	I,2	ERS227795	4	16	3 (14 to 16)	nd	47	10	3 (9 to 11)	nd
UK095	2012	I,3	ERS227796	47	10	3 (9 to 11)	nd	16	14	13 (6 to 18)	nd
UK096	2012	I,2	ERS227797	4	7.5	3 (6 to 8)	nd	0	nd	nd	nd
UK097	2012	I,3	ERS227798	41	10	3 (9 to 11)	nd	21	14	7 (10 to 16)	nd
UK098	2012	I,3	ERS227799	24	10	3 (9 to 11)	nd	14	14	13 (6 to 18)	nd
UK099	2012	I,2	ERS227800	9	14	11 (6 to 16)	nd	27	11	5 (9 to 13)	nd
UK100	2012	I,3	ERS227801	38	10	2 (10 to 11)	nd	10	14	4 (12 to 15)	nd

Table S3 Promoter poly(C) tract lengths of *fim2* & *fim3*, and serotypes by agglutination, of *B. pertussis* isolates UK001 to UK024 and UK049 to UK100. ENA identifiers are European Nucleotide Archive sample names. Serotype by agglutination uses the nomenclature of Robinson *et al.* (1989b). Surface fluorescence was not determined (nd). Values (including fluorescence) for the other 22 UK isolates are in Table 2.