“Integrated” and “insulated” boronate-based fluorescent probes for the detection of hydrogen peroxide
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Integrated and insulated boronate-based fluorescent probes have been evaluated for the detection of hydrogen peroxide in the presence of saccharides.

Hydrogen peroxide (H$_2$O$_2$) is the simplest peroxide (a compound with an oxygen-oxygen single bond) and it is a strong oxidizing agent, which has been widely employed as a bleach and as a cleaning reagent to reduce BOD$^1$ and COD$^2$ from industrial wastewater. Hydrogen peroxide, one of reactive oxygen species (ROS), plays an important role as a signalling molecule in the regulation of a variety of biological processes, such as immune response, cell signalling,3 Alzheimer’s diseases3 and cancer.5 The importance of H$_2$O$_2$ has led to researchers seeking effective and applicable approaches for its detection.

Among the powerful tools available for H$_2$O$_2$ detection are synthetic fluorescent probes. Hydrogen peroxide reacts with arylboronic acids under mild alkaline conditions to generate phenols (Scheme 1).6 Over the last decade, several groups introduced different fluorescent probes for the detection of hydrogen peroxide by utilising this property. Chang and co-workers have developed a series of boronate-based derivatives for the fluorescent detection of H$_2$O$_2$ in living systems,7–13 and Tomapatanaghet al. used the H$_2$O$_2$ generated from glucose by the action of glucose oxidase to convert boronic acids to phenols in their glucose sensing system.14

We have a long-standing interest in boronic acids for saccharide binding, operating a PET mechanism.22

Bearing this in mind, we decided to investigate how the reaction of H$_2$O$_2$ with two classes of boronic acid (integrated and insulated) fluorescent probes changes in the presence of saccharides. This is conceptually similar to the inspirational work by Jiang into the effect of saccharides on Suzuki homocoupling reactions.23 Understanding the effect of saccharides on this reaction and fluorescence may be important in developing advanced intracellular probes to accurately map H$_2$O$_2$ concentrations in living systems. Probes 1 and 2 (Scheme 2) and their saccharide (α-fructose) was chosen as a model saccharide since it has a high binding constant) complexes were evaluated as fluorescent indicators for hydrogen peroxide both in neutral and alkaline aqueous solutions. In the corresponding experiments, fluorescence titrations with H$_2$O$_2$ were carried out at 25 °C in pH 7.20 buffer (52.1% methanol in water with KCl, 10 mM; KH$_2$PO$_4$, 2.752 mM; Na$_2$HPO$_4$, 2.757 mM and the pH was adjusted to 7.20 using HCl) and pH 9.70 buffer (10.0% methanol in water with Na$_2$CO$_3$, 50 mM; NaHCO$_3$, 50 mM). The saccharide-boronic acid complexes were formed by mixing free boronic acid (10 µM) with α-fructose (100 mM) in situ.25,26

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† Joseph Priestley, the discoverer of oxygen, was born in 1733. This paper is dedicated to the 280th anniversary of his birth.
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The binding α-fructose to 2-naphthylboronic acid (1), in the absence of H₂O₂, causes a decrease and an increase in fluorescence intensity at pH 7.20 (Fig. S1, ESI‡) and pH 9.70 (Fig. S2, ESI‡) respectively. When H₂O₂ is present, 2-naphthylboronic acid is oxidatively converted to 2-naphthol (Scheme 1), and the fluorescence is reduced at 340 nm and red-shifted to 410 nm from pH 7.2 to 9.7, when excited at 290 nm (Fig. S4, ESI‡).

Fig. 1 summarises the reaction scheme and the fluorescence intensity changes $F\text{T}/F\text{0}$ (in the presence of H₂O₂)/$F\text{T}/F\text{0}$ (in the absence of H₂O₂) for fluorescent probe 1 and the 1-α-fructose complex in pH 7.20 and 9.70 buffer. It is known that on saccharide binding and formation of a cyclic boronate ester, the pKₐ of the boronic acid is enhanced, or in other words the ‘ester’ is more acidic than the ‘acid’. Therefore, binding with α-fructose increases the electrophilicity of boron in probe 1, making it more easily oxidised by H₂O₂. This observation was borne out by our experiments. As shown in Fig. 1(b), the fluorescence decrease over the observed time (55 min) of probe 1 (10 μM) was ca. 0.85 ($F\text{T}/F\text{0}$) while there was a larger change for the 1-α-fructose complex, ca. 0.80 ($F\text{T}/F\text{0}$) upon exposure to H₂O₂ (0.10 mM) over the same period in neutral buffer solution. Under alkaline conditions, the 1-α-fructose boronate ester complex reacted even faster with H₂O₂ (0.10 mM) ($F\text{T}/F\text{0} = ca. 0.30$) than the unbound boronic acid 1 ($F\text{T}/F\text{0} = ca. 0.60$). The responses of 1 and the 1-α-fructose complex towards H₂O₂ became more rapid at high pH since not only is the binding between boronic acid and saccharide enhanced, but also H₂O₂ is also more reactive at higher pH.

Fluorescence was confirmed to be due to the generation of 2-naphthol by comparison with spectroscopic properties of an authentic sample. i.e. The emission maxima for 2-naphtol (10 μM, pKₐ = 9.51) was found at 340 nm in pH 7.20 PBS buffer while the maxima shifted to 410 nm in pH 9.70 alkaline buffer (Fig. S4, ESI‡). These results demonstrate the proton-induced fluorescence switching of 2-naphthol at pH 7.20 and the electron donation of the negative oxygen to the fluorophore causing the red-shift at pH 9.70.

In the case of N-methyl-α-(aminomethyl)phenylboronic acid (2), a significant ‘off–on’ signal response has been seen on binding with α-fructose (Fig. S5, ESI‡) due to a PET mechanism. However, when the arylboronic acid moiety of probe 2 was transformed into a phenol upon adding H₂O₂, the fluorescence was further reduced due to the stronger PET from the amine in the boron free system (Fig. S5–S7, ESI‡).

The reaction scheme and fluorescence intensity changes for fluorescent probe 2 and the 2-α-fructose complex in pH 7.20 and 9.70 buffer are given in Fig. 2. From Fig. 2(b), it can be seen that the fluorescence intensity ratios with time ($F\text{T}/F\text{0}$) of 2-α-fructose complex slowly change to ca. 0.72, while for the saccharide free system a bigger change $F\text{T}/F\text{0}$ = ca. 0.55 on addition of 0.05 mM H₂O₂ over three hours in pH 7.20 PBS buffer (probe 2, 10 μM). The difference between 2 and the 2-α-fructose complex towards H₂O₂ became much larger in pH 9.70 buffer solution with $F\text{T}/F\text{0}$ = ca. 0.05 within 20 min upon addition of only 0.02 mM H₂O₂ (Fig. S8, ESI‡). More importantly, the signal to noise ratio output of probe 2 was enhanced through binding with saccharide allowing for colorimetric detection (bright blue to colourless) in the reaction with H₂O₂ (Fig. 3, Fig. S9, ESI‡).

The fluorescence of 2 is turned on by saccharide binding, since boronic ester formation causes an enhanced interaction between the neighbouring amine and the boron atom (mediated via an inserted solvent molecule). This N–B interaction hinders the reaction between boron and H₂O₂ in the presence of saccharides resulting in a slower decrease in fluorescence (cf. saccharide free system).

In conclusion, α-fructose effects the reaction of integrated (compound 1) and insulated (compound 2) boronic acid-based fluorescent probes with hydrogen peroxide in opposite directions. Integrated boronic acid fluorophores (compound 1) display enhanced reactivity with H₂O₂ in the presence of α-fructose. While the PET fluorophore systems (compound 2) display reduced reactivity with H₂O₂ in the presence of α-fructose. The insulated PET systems are particularly interesting, (compound 2) because, in the presence of α-fructose the initial fluorescence intensity is much higher and produces a blue visible fluorescence, which implies that they could be used as temporal fluorescent probes to map both intracellular H₂O₂ and saccharide concentrations. The insulated systems also produce...
Therefore, we believe that insulated (PET) systems are the fluorescent probes of choice for use as imaging agents and sensors for H$_2$O$_2$.

We are currently exploring the development of longer wavelength insulated systems for biologically relevant imaging applications.

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Notes and references

1. Biochemical oxygen demand (BOD) is the amount of dissolved oxygen needed by aerobic biological organisms in a body of water to break down organic material present in a given water sample at certain temperature over a specific time period.

2. In environmental chemistry, the chemical oxygen demand (COD) test is commonly used to indirectly measure the amount of organic compounds in water.


29. Given that the reactions take time to reach completion we confirmed that H$_2$O$_2$ is stable in pH 9.70 buffer at 25 °C for two hours. [Fig. S10, ESI].
