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1 **Fundamental differences in physiology of *Bordetella pertussis* dependent on the two-component**  
2 **system Bvg, revealed by gene essentiality studies.**

3 Conditional gene essentiality in *B. pertussis*.

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16

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18

19 Repositories: Sequence data files are available from the European Nucleotide Archive  
20 (<https://www.ebi.ac.uk/ena>), with accession numbers ERS2490348 (Bvg+ phase data) and  
21 ERS2490349 (Bvg- phase data).

22

23 **Abstract.**

24 The identification of genes essential for a bacterium's growth reveals much about their basic  
25 physiology under different conditions. *Bordetella pertussis*, the causative agent of whooping cough,  
26 adopts both virulent and avirulent states through the activity of the two-component system, Bvg.  
27 The genes essential for *B. pertussis* growth in vitro were defined using transposon sequencing, for  
28 different Bvg-determined growth states. In addition, comparison of the insertion indices of each  
29 gene between Bvg phases identified those genes whose mutation exerted a significantly different  
30 fitness cost between phases. As expected, many of the genes identified as essential for growth in  
31 other bacteria were also essential for *B. pertussis*. However, the essentiality of some genes was  
32 dependent on Bvg. In particular, a number of key cell wall biosynthesis genes, including the entire  
33 *mre/mrd* locus, were essential for growth of the avirulent (Bvg minus) phase but not the virulent  
34 (Bvg plus) phase. In addition, cell wall biosynthesis was identified as a fundamental process that  
35 when disrupted produced greater fitness costs for the Bvg minus phase compared to the Bvg plus  
36 phase. Bvg minus phase growth was more susceptible than Bvg plus phase growth to the cell wall  
37 disrupting antibiotic ampicillin demonstrating the increased susceptibility of the Bvg minus phase to  
38 disruption of cell wall synthesis. This Bvg-dependent conditional essentiality was not due to Bvg-  
39 regulation of expression of cell wall biosynthesis genes; suggesting that this fundamental process  
40 differs between the Bvg phases in *B. pertussis* and is more susceptible to disruption in the Bvg minus  
41 phase. The ability of a bacterium to modify its cell wall synthesis is important when considering the  
42 action of antibiotics, particularly if developing novel drugs targeting cell wall synthesis.

43 **Impact Statement.**

44 This study identifies differences in gene essentiality for *B. pertussis*, dependent on the activity of the  
45 key Bvg regulatory system. The role of Bvg in regulating the pathogenicity of *B. pertussis* is well-

46 established. Here we demonstrate a role in fundamental physiology and surprising differences in cell  
47 wall synthesis between Bvg phases. The use of functional genomics to identify the genetic bases for  
48 key traits has greatly enhanced our understanding of genotype-phenotype links. Here, we  
49 demonstrate that the role of key genes can be conditional, in this case dependent on the activity of a  
50 two-component regulatory system. We identify cell wall biogenesis as a process that appears to  
51 differ in *B. pertussis* between Bvg states. This suggests novel links between fundamental *B. pertussis*  
52 cell growth processes and its pathogenicity and virulence. Our understanding of the role of bacterial  
53 physiology, including metabolism in bacterial infection has expanded significantly in recent years.  
54 Elucidating the role in *B. pertussis* of Bvg in regulating cell wall biogenesis will deepen this  
55 understanding.

56

#### 57 **Data Summary.**

58 Sequence data files are available from the European Nucleotide Archive  
59 (<https://www.ebi.ac.uk/ena>), with accession numbers ERS2490348 (Bvg+ phase data) and  
60 ERS2490349 (Bvg- phase data).

61

#### 62 **Introduction**

63 **Essential genes.** Allelic replacement mutagenesis is a cornerstone in the study of bacterial  
64 pathogenicity. A defined mutation is constructed to produce a strain that is identical to wild type  
65 (WT) except for the loss of the targeted gene. Comparison between the phenotypes of WT and  
66 mutant reveals differences in behaviour that are ascribed to the loss of the targeted gene from the  
67 mutant. In this way, the genetic bases of numerous traits have been established and targeted  
68 mutagenesis used to perturb traits to understand them. However, genes that are essential for the  
69 viability of a bacterium cannot be studied in this way because, by definition, the bacteria cannot

70 withstand a knock out mutation in these genes and remain viable. Often it is not easy to establish  
71 that a gene is essential, as the failure to obtain viable mutants is a negative screen and its not  
72 obvious if this failure is because the gene is essential or because the many steps required to achieve  
73 the necessary homologous recombination have failed. There is great interest in defining the  
74 essential genes of bacteria as part of understanding gene function and genetic and pathway  
75 networks (1). This identifies the core processes required by the bacterium for cell growth,  
76 maintenance and division, and can reveal important information about the basic physiology and  
77 anatomy of the cell, such as particular metabolic pathways used. Furthermore, identification of  
78 processes not absolutely required by the bacterium for viability can be just as informative. Currently,  
79 there is great interest in identifying genes essential for viability of bacterial pathogens, as these  
80 represent potential targets for the design of novel antimicrobials to combat the relentless rise of  
81 antimicrobial resistance (2).

82 **TraDIS/TnSeq.** Post-genomic techniques have proved powerful, high-throughput approaches to  
83 dissecting the genetic bases for numerous different traits. In particular, Transposon Directed  
84 Insertion Sequencing (TraDIS), or TnSeq, is well suited to identifying the essential genes for bacterial  
85 viability for whatever conditions the screen has been performed under (3, 4). A large scale, high  
86 density, library of transposon mutants is generated. These mutants are pooled and genomic DNA  
87 extracted. Utilising next generation sequencing, the boundaries between the transposon insertions  
88 and neighbouring DNA are sequenced and mapped to the corresponding genome sequence. This  
89 identifies precisely the insertion sites for all of the different transposon mutants recovered. If the  
90 density of insertions is sufficiently high, then there is high confidence that any genes for which no  
91 insertions are observed are those genes that cannot be mutated by insertion to produce a viable  
92 mutant, i.e. are the essential genes.

93 ***B. pertussis.*** *B. pertussis* is the primary causative agent of whooping cough, or pertussis. A well-  
94 documented resurgence of pertussis has been observed in many countries in recent years (5). This

95 has been attributed to several factors, notably a switch from whole cell to acellular pertussis  
96 vaccines in many developed countries during the early 2000's (5). These vaccines appear to protect  
97 infants from serious disease but induce immunity that wanes more quickly than that induced by  
98 whole cell vaccines, and does not prevent colonisation to the same degree as whole cell vaccines.  
99 Combined, it is thought there is increased circulation/transmission of *B. pertussis* in countries using  
100 acellular vaccines. In addition, there are signatures of vaccine-induced selection among *B. pertussis*  
101 and it is feared this selection is favouring the emergence of lineages that are less well controlled by  
102 vaccination (6, 7).

103 **Bvg.** The *Bordetella* contain a two component regulator, Bvg, comprised of an inner membrane  
104 histidine kinase, the sensor, BvgS; and the response regulator, BvgA (8). Two component systems  
105 sense stimuli, often from the bacterium's environment, and respond to it by regulating changes in  
106 gene expression patterns through the activity of the response regulator, that are usually  
107 transcriptional regulators. Bvg directs *B. pertussis* through a spectrum of different phenotypes. The  
108 phenotypes at the extremes of this spectrum are the best characterised. In the absence of stimulus,  
109 the levels of phosphorylated BvgA, and thus its transcriptional activating activity, are low. Many of  
110 the genes implicated in *B. pertussis* infection, including toxins, adhesins and immune evasion factors  
111 are expressed at very low levels (if at all) whereas other genes are expressed at high levels. This state  
112 is referred to as the Bvg minus (Bvg-) phase. On sensing stimulus, activated BvgS phosphorylates  
113 BvgA which triggers its transcriptional activating activity. Through binding to the promoters of target  
114 genes, BvgA enhances the transcription of many virulence factors, and also represses a number of  
115 the genes that were highly expressed in the Bvg- phase. This virulence-competent state is referred to  
116 as the Bvg plus (Bvg+) phase. Bvg transitions *B. pertussis* through a spectrum of states, dependent on  
117 the magnitude and duration of the stimulus sensed by BvgS. Initial studies using microarrays  
118 identified 291 *B. pertussis* genes differentially regulated by Bvg (9). RNAseq analysis has identified  
119 around 550 genes whose expression is as changed at least 2 fold between Bvg+ and Bvg- phase  
120 conditions (10), comprising 14% of the *B. pertussis* Tohama I gene repertoire. Thus, Bvg regulates the

121 expression of a major portion of *B. pertussis* genes. In addition to genes involved in infection, Bvg  
122 appears to regulate many other processes, including metabolism. For example, the Bvg- phase is  
123 characterized by a faster growth rate than the Bvg+ phase (11). Thus, Bvg is regarded as a master  
124 regulator of *B. pertussis*, directing it between very different phenotypes. The stimuli sensed by Bvg  
125 are unclear, but in vitro the system responds to temperature in that *B. pertussis* growing at 37°C are  
126 in the Bvg+ phase whereas they transition to the Bvg- phase at temperatures below around 27°C.  
127 Also, the Bvg- phase is induced by growth at 50mM MgSO<sub>4</sub>, even at 37°C.

128 Previously, the *B. pertussis* genes essential for growth in vitro, and those conditionally essential in  
129 vivo in a mouse model of infection were defined (12). Here we define the genes essential for growth  
130 of *B. pertussis* in vitro under both Bvg+ and Bvg- phase conditions. As expected, many of the core  
131 processes involved in bacterial growth and replication were identified as essential. However, a  
132 surprising number of genes were identified as conditionally essential, dependent on Bvg phase.  
133 Furthermore, our use of two different conditions enabled the identification of genes which although  
134 not essential, their mutation decreases the fitness of *B. pertussis* in a Bvg-dependent manner. Our  
135 data identified cell wall biogenesis as a process whose disruption is less well tolerated by Bvg- phase  
136 *B. pertussis* compared to Bvg+ phase. The ability of Bvg+ *B. pertussis* to grow despite mutation of a  
137 range of key cell wall biosynthesis genes suggests that this fundamental process differs significantly  
138 between Bvg phases.

## 139 **Methods**

140 **Bacteria and plasmids.** *B. pertussis* BP536 is a streptomycin-resistant clone of Tohama I. BP536 was  
141 grown on charcoal agar at 37°C for 3 days. Bvg- phase conditions were achieved by plating on  
142 charcoal agar containing 50mM MgSO<sub>4</sub> or 16mM nicotinic acid. *E. coli* ST18 was used a donor strain  
143 for conjugations (13). It was grown in Luria Bertani (LB) broth with shaking or on LB agar, at 37°C.  
144 ST18 is auxotrophic for aminolevulinic acid (ALA) and thus media were supplemented with 50mM  
145 ALA. *E. coli* 5 alpha (NEB, Hitchin, U.K.) were used for routine cloning and plasmid maintenance.

146 Selection of strains carrying plasmid pBam1 (14) or ep1 was achieved by supplementing media with  
147 50µg ml<sup>-1</sup> kanamycin. pBBR1MCS (15) was used as a shuttle vector to carry a plasmid borne copy of  
148 *mreB* in *B. pertussis*.

149 **DNA manipulations.** Routine DNA manipulations were performed using reagents from NEB or  
150 Qiagen (Manchester, U.K.) according to the manufacturer's instructions.

151 **Construction of ep1.** Plasmid pBAM1 (14) was modified by the addition of additional *PmeI*  
152 restriction endonuclease recognition sites adjacent to the mosaic ends that are recognised by the  
153 transposase TnpA, to produce plasmid ep1. This enables cleavage of plasmid derived DNA to prevent  
154 sequencing from plasmid that might be purified from conjugation plates. *PmeI* cuts only once in the  
155 *B. pertussis* BP536 genome (within BP0823).

156 **Construction of transposon mutant libraries.** *B. pertussis* BP536 were conjugated with *E. coli* ST18  
157 ep1, as described previously (16). Conjugants were selected by plating onto charcoal plates  
158 supplemented with 50µg ml<sup>-1</sup> kanamycin and incubating at 37°C for three days. For collection of high  
159 density transposon mutant libraries, nine independent conjugations were performed, and each  
160 plated onto multiple 140mm diameter agar plates. Colonies were recovered into PBS and processed  
161 immediately for genomic DNA extraction using the Gentra Pure Yeast/Bact kit (Qiagen, Manchester,  
162 U.K.). For this, portions of each resuspension were mixed in proportion to the number of colonies  
163 obtained from the conjugation to give two separate pools of genomic DNA for Bvg+ phase and Bvg-  
164 phase conjugants. The purified DNA was digested with *PmeI* to prevent sequencing from ep1  
165 plasmid DNA that was harvested from dead donor *E. coli* present on the conjugation plates.

166 **DNA sequencing of transposon mutant libraries.** 2µg of genomic DNA was fragmented by Covaris to  
167 an average size of ~300 bp. Following end-repair and 'A' tailing a modified Illumina adapter,  
168 synthesized and annealed by IDT using oligonucleotides SplA5\_top and SplA5\_bottom (Table 3) was  
169 ligated to the fragments for 40 mins at 20C. Ligated fragments were cleaned using Ampure XP beads



170 (Beckman Coulter, High Wycombe, U.K.) with a beads to sample ratio of 0.8 to 1. PCR enrichment of  
171 fragments containing transposon was performed using primers homologous to each end of the  
172 transposon (EP1 5'PCR or EP1 3'PCR) in conjunction with an adapter-specific primer containing an  
173 index tag (SplAP5.x). PCR products were cleaned using Ampure XP beads as before, quantified by  
174 qPCR and then pooled. PhiX library was added to a level of 6%. The pooled libraries were sequenced  
175 on a HiSeq2500 using a specially modified recipe to overcome difficulties generated by the  
176 monotemplate transposon sequence. Briefly, a transposon-specific sequencing primer (EP1 5'seq or  
177 EP1 3'seq) anneals to the transposon 10 bases away from the junction between transposon and *B.*  
178 *pertussis* chromosome. Sequencing takes place with no imaging for the first 10 cycles followed by  
179 imaging for the next 50 cycles. This gives a 50bp genomic DNA read. The template is denatured and  
180 the same sequencing primer is re-annealed and 10 cycles of sequencing takes place to give a 10bp  
181 transposon read. To reduce background plasmid DNA libraries were digested with *PmeI* at the post-  
182 ligation stage for the 3' transposon ends and a blocking oligo (added during PCR enrichment) for the  
183 5' transposon ends.

Primer	Sequence (5' to 3')
SplA5_top	G*AGATCGGTCTCGGCATTCTGCTGAACCGCTCTCCGATC*T
SplA5_bottom	/5Phos/G*ATCGGAAGAGCGGTTTCAGCAGGTTTTTTTTTTTCAAAAAA*A
EP1 5'PCR	AATGATACGGCGACCACCGAGATCTACACTTATTGTTTCATGATGATATATTTTTATCTTGT GC
EP1 3'PCR	AATGATACGGCGACCACCGAGATCTACACGCAGGTCGACTCTAGAGGATCCCC
EP1 5'seq	TAACATCAGAGATTTTGAGACACAAGACGTCAGATGTGTA
EP1 3'seq	GCGGCCTAGGCGGCCTTAATTAAGATGTGTA
5'plasmid blocking oligo	CATCAGATTCTGGAAAACGGGAAAGGTTCCGTTTCAGGACGCTACTTGTGTAGTTTAAACC AGCTGG**
SplAP5.1	C*AAGCAGAAGACGGCATAACGATGAGATCGGTCTCGGCATTC*C
SplAP5.2	C*AAGCAGAAGACGGCATAACATCGGAGATCGGTCTCGGCATTC*C
SplAP5.3	C*AAGCAGAAGACGGCATAACGATATGCCTAAGAGATCGGTCTCGGCATTC*C
SplAP5.4	C*AAGCAGAAGACGGCATAACGATAGTGGTCAGAGATCGGTCTCGGCATTC*C
SplAP5.5	C*AAGCAGAAGACGGCATAACGATACCACTGTGAGATCGGTCTCGGCATTC*C
SplAP5.6	C*AAGCAGAAGACGGCATAACGATACATTGGCGAGATCGGTCTCGGCATTC*C
SplAP5.7	C*AAGCAGAAGACGGCATAACGATCAGATCTGGAGATCGGTCTCGGCATTC*C
SplAP5.8	C*AAGCAGAAGACGGCATAACGATCATCAAGTGAAGATCGGTCTCGGCATTC*C
SplAP5.9	C*AAGCAGAAGACGGCATAACGATCGCTGATCGAGATCGGTCTCGGCATTC*C
SplAP5.10	C*AAGCAGAAGACGGCATAACGATACAAGCTAGAGATCGGTCTCGGCATTC*C
SplAP5.11	C*AAGCAGAAGACGGCATAACGATCTGTAGCCGAGATCGGTCTCGGCATTC*C
SplAP5.12	C*AAGCAGAAGACGGCATAACGATAGTACAAGGAGATCGGTCTCGGCATTC*C
mreBLF	AAAAGGTCTCTCGAGATTCGGGGGCGTTGG

mreBLR	AAAAGGTCTCCATGTGCATGGGAGCTCAGCTAGATTC
mreBRF	AAAAGGTCTCAGGTCTGAGCCTGTCTCGCG
mreBRR	AAAAGGTCTCGAACTGGGCGGCTCGTACAGC
KanF	AAAAAAGGTCTCCACATGACGTCTTGTGTCTCAAATCTC
KanR	AAAAAAGGTCTCAGACCTTAGAAAAATTCATCCAGCATC

184 Table 3. Primers used in this study.

185 \*: phosphorothioate group modification. \*\* dideoxy base.

186 **Analysis of DNA sequence.** Sequencing data was analysed as described (3) to process reads, map  
187 them to the Tohama I reference genome sequence, define the number of unique insertions, the  
188 insertion index for each gene, predict the essentiality of each gene, and to compare mutant  
189 frequencies between the Bvg+ and Bvg- phase conditions. Briefly, an insertion index for each gene  
190 was calculated by dividing the number of unique insertion sites within the gene by gene length (to  
191 normalise for gene length). The distribution of insertion indices was bimodal with a narrow peak  
192 corresponding to essential genes without insertions and a broader distribution for genes able to  
193 tolerate insertions. Fitting gamma distributions to these two modes enabled calculation of Log2  
194 likelihood ratios between the two modes. An LLR of < -2 was used to assign a gene as essential  
195 (indicating it was four times more likely to be essential than non-essential), while an LLR of > 2 was  
196 used to assign as non-essential. Genes falling between these values were classed as ambiguous in  
197 terms of their essentiality. Identification of genes for which there were significantly different  
198 numbers of read between Bvg phases was conducted as described (3).

199 **Availability of Data and Materials.** Sequence data files are available from the European Nucleotide  
200 Archive (<https://www.ebi.ac.uk/ena>), with accession numbers ERS2490348 (Bvg+ phase data) and  
201 ERS2490349 (Bvg- phase data).

202 **Antibiotic susceptibility testing.** The MICs of Bvg- and Bvg+ phase *B. pertussis* for antibiotics were

203 determined using E-test<sup>R</sup> strips (BioMérieux, Basingstoke, U.K.). BP536 was grown on charcoal agar  
204 and resuspended in PBS to an OD<sub>600</sub>= 0.2. 100µls of this suspension was plated onto charcoal agar or  
205 charcoal agar supplemented with 50mM MgSO<sub>4</sub>. Once the plates were dry, an E-test strip was  
206 placed centrally and the plates were incubated at 37°C for 72 hours. MICs were read as described:  
207 [http://www.biomerieux.co.uk/sites/subsidiary\\_uk/files/etest-reading-guide-aerobic-bacteria.pdf](http://www.biomerieux.co.uk/sites/subsidiary_uk/files/etest-reading-guide-aerobic-bacteria.pdf).

208 The average and standard error of MICs for triplicate samples were calculated and significance  
209 determined using a T-test.

210 **Construction of *mreB* mutant.** Allelic exchange mutagenesis was used to delete the *mreB* gene and  
211 replace it with a kanamycin resistance cassette. Regions flanking *mreB*, and a kanamycin resistance  
212 gene cassette were amplified by PCR (Q5 high-fidelity polymerase, NEB) using the primers indicated  
213 in Table 1. They were cloned using Golden Gate cloning into plasmid pCR8 (Invitrogen,  
214 Loughborough, U.K.) modified for Golden Gate cloning. This cloned region was subcloned into the  
215 allelic exchange vector pSS4940 using Gateway technology (Invitrogen). The resulting plasmid was  
216 transformed into the conjugation donor *E. coli* strain ST18, and introduced into *B. pertussis* by  
217 conjugation as described previously (16) Conjugants were selected on charcoal agar supplemented  
218 with 50mM MgSO<sub>4</sub> and 30µM gentamycin at 37°C for 3-4 days. Individual clones were passaged on  
219 this media prior to passaging on charcoal agar supplemented with 50µg/ml kanamycin at 37°C for 3-  
220 4 days to select for allelic exchange mutants. Individual clones were analysed using PCR to confirm  
221 the replacement of the *mreB* gene by the kanamycin resistance cassette.

222 **Viable counts of *mreB* mutant.** WT and *B. pertussis mreB* were grown on charcoal agar. Bacteria  
223 were suspended in PBS and serially diluted. 100µls of dilutions were plated onto charcoal agar and  
224 charcoal agar supplemented with 50mM MgSO<sub>4</sub> or 16mM nicotinic acid, incubated at 37°C for 4 days  
225 and the resulting colonies were imaged.

226

227 **Results.**

228 To create the library of transposon mutants, nine independent conjugations were performed in  
229 which the transient expression of Tn5 transposase led to mTn insertions in to the *B. pertussis*  
230 chromosome. Plasmid ep1 contains a modified mTn5 containing the Tn5 direct repeats flanking a  
231 kanamycin resistance cassette. It carries the Tn5 transposase gene but outside of the repeats on  
232 which it acts. Plasmid ep1 cannot replicate in *B. pertussis* but on delivery into these bacteria by  
233 conjugation, the transient expression of the transposase enables transposition of the kanamycin  
234 cassette onto the *B. pertussis* chromosome but subsequent loss of the plasmid. Conjugants from  
235 each conjugation were selected on agar under both Bvg+ and Bvg- phase conditions. Approximately  
236 490 000 transposon insertion mutants were recovered for each Bvg phase condition. Bvg+ phase and  
237 Bvg- phase mutants were recovered separately for each conjugation. Single pools of Bvg+ and Bvg-  
238 phase mutants were created by combining colonies obtained from each conjugation in proportion to  
239 the number of colonies recovered in each, and genomic DNA was extracted for each pool and  
240 subjected to sequencing. For the Bvg+ and Bvg- mutant libraries, 300 581 and 316 281 unique  
241 insertion sites were identified, corresponding to an average of an insertion every 13.6 and 12.9 bp  
242 respectively across the 4,086,189 bp *B. pertussis* genome, i.e. very high-density transposon libraries  
243 had been recovered.

244 Using the approach described previously (3, 17), genes essential for growth under Bvg+ and Bvg-  
245 phase conditions were calculated. Due to the uncertainty regarding the ability of bacteria to tolerate  
246 insertions in the termini of essential genes, here insertions in the terminal 10% of each gene were  
247 discounted for the purpose of essential gene identification. The number of unique insertion sites  
248 within each gene was normalised by gene length to give an insertion index for each gene. Plotting  
249 the frequency distribution of insertion indices produces a bimodal distribution representing essential  
250 and non-essential genes. This was used to calculate a likelihood ratio of whether each gene was  
251 more likely to be in the essential gene peak of the distribution and a cut off value was calculated to  
252 define whether a gene was considered essential or non-essential. The gamma-fits of these  
253 distributions for insertion indices for the Bvg+ and Bvg- phase mutants are shown in Figure 1.

254 Three hundred and ninety six genes were identified as essential, in that they were required for  
 255 growth of both Bvg+ and Bvg- phase *B. pertussis* under the conditions tested here, Table S1. A  
 256 further 26 genes were ambiguous for essentiality for each of the Bvg+ and Bvg- phases, Table S2, in  
 257 that their insertion indices fell between the two peaks in the distribution of frequencies.

258 The broad functional categories of these genes are listed in Table 1. Three broad functional classes  
 259 are richly represented among these genes, comprising 203 of the 396 essential genes:  
 260 translation/ribosome structure, cell wall and cofactor biosynthesis. These processes are also richly  
 261 represented in the essential gene lists of many other bacteria, highlighting the importance of these  
 262 core processes for cellular viability and growth, and the large number of genes involved in these  
 263 processes. Also, as expected, key metabolic genes were identified as essential. For example, *B.*  
 264 *pertussis* does not possess a functional glycolytic pathway and does not metabolise sugars as a  
 265 carbon source (18, 19). It is assumed that gluconeogenesis is used to synthesise the sugar  
 266 components of *B. pertussis* via the metabolism of glutamate, the main carbon source in the media  
 267 used here (20). In agreement with this, every gene encoding a gluconeogenesis enzyme is essential.

268 Table 1. Functional categories of essential and fitness-affected genes.

	Unconditional	Bvg+ only	Bvg- only
Amino acid biosynthesis	19	0 (1)	9 (4)
Capsule biosynthesis	0	0 (7)	0 (0)
Cell division	12	1 (3)	2 (3)
Cell wall	54	4 (2)	19 (17)
Chaperone	4	0 (0)	0 (0)
Chemotaxis	0	0 (0)	1 (0)
Cofactor biosynthesis	52	1 (1)	4 (1)
DNA repair	4	3 (0)	2 (1)
DNA replication	16	0 (2)	3 (2)

Electron transport	6	0 (0)	0 (3)
Energy metabolism	23	0 (0)	1(0)
General metabolism	1	1 (4)	0 (1)
Glycolysis/gluconeogenesis	10	0 (0)	0 (0)
Homeostasis	14	0 (4)	5 (2)
Lipid metabolism	14	0 (1)	1 (2)
Mobile Genetic Element	0	0 (1)	0 (0)
Nucleotide metabolism	7	0 (0)	0 (0)
Oligosaccharide biosynthesis	0	0 (3)	0 (0)
Pentose phosphate pathway	3	0 (0)	0 (0)
Peptidase	1	0 (0)	0 (0)
Phospholipid metabolism	6	0 (0)	0 (0)
Protein secretion	10	0 (0)	1 (1)
Purine metabolism	6	0 (0)	0 (0)
Pyrimidine metabolism	3	0 (0)	0 (1)
Response regulation	2	0 (1)	1 (1)
RNA metabolism	1	0 (0)	0 (0)
Small molecule transport	2	0 (4)	3 (4)
Stress response	3	0 (2)	0 (1)
TCA cycle	8	0 (0)	6 (0)
Transcription	10	2 (2)	2 (2)
Translation/ribosome structure	98	2 (1)	12 (1)
Unknown	7	5 (11)	7 (4)
Total	396	19 (50)	79 (51)

269 **Table 1.** The number of genes within different functional categories identified as essential under  
270 both Bvg+ and Bvg- phase conditions (unconditional), or in only the Bvg+ or Bvg- phase. The number  
271 of genes within each functional category that although not essential produce a statistically different  
272 fitness cost between phases is in parentheses.

273 There were just 7 genes identified as being essential for which involvement in a particular cellular  
274 process could not be predicted. These represent potentially novel core processes in *B. pertussis* or  
275 novel functions in previously characterised core processes. Homologues of all of the predicted  
276 proteins encoded by these genes are present in other bacteria. Of these 7 predicted proteins, only 1  
277 contains strong homology to conserved domains as revealed by BlastP homology searches. On the  
278 basis of these results, BP2815 is a putative glycosyl hydrolase. However the substrate, and thus  
279 cellular function, of BP2815 is unknown.

280 Of particular interest was the identification of genes that were essential in only either the Bvg+ or  
281 Bvg- phase, i.e. conditionally essential genes. Nineteen genes were identified as being essential in  
282 the Bvg+ phase but not Bvg- phase, Table S1. Six of these genes were, however, described in the  
283 ambiguous category in the Bvg- phase meaning it is not definitively clear as to whether they are  
284 conditionally essential. For those genes that have been either characterised or for which a putative  
285 function prediction is possible, a variety of functions are evident. There is not a particular functional  
286 category that is richly represented within this group and thus there is not a specific process  
287 identifiable that is essential in only the Bvg+ phase.

288 In contrast, there are 79 genes that are classified as essential in only the Bvg- phase. 19 of these are  
289 classed as ambiguous in Bvg+ phase bacteria, but this still identifies a large number of genes as  
290 essential only under Bvg- phase conditions. Among these genes three broad functional categories  
291 stand out: 19 genes are predicted to encode cell wall biosynthesis functions, 12 genes are involved in  
292 translation/ribosome structure and 6 genes encode components of TCA cycle enzyme complexes.



293 Many genes are not essential but mutation of them carries a fitness cost that results in transposon  
294 insertions in that gene being recovered at relatively low frequency compared to genes that are  
295 completely dispensable for growth under the conditions tested. Statistical comparison of the  
296 number of sequence reads mapping to each gene between the Bvg phases enabled the identification  
297 of genes whose mutation induces significantly different fitness costs between Bvg phases. This  
298 analysis identified 50 genes for which insertion mutants were recovered at lower frequency for Bvg+  
299 phase and 51 genes for which mutants were recovered at a lower frequency for the Bvg- phase,  
300 Table S3. Of these genes only 8 (Bvg+) and 16 (Bvg-) were essential (conditionally or otherwise),  
301 demonstrating the power of this approach to extend the analysis of the fitness of mutants between  
302 Bvg phases. It is important to note that this does not mean that mutations in conditionally essential  
303 genes in the permissive phase does not cause a fitness defect. For some of these genes, their  
304 insertion indices in the permissive phase were low (but above the threshold for essentiality)  
305 suggesting that while viable mutants were recovered, their numbers were low compared to mutants  
306 for truly dispensable genes. However, when comparing the read counts for these genes between the  
307 the two phases, as in our analysis of fitness, the difference does not reach significance.

308 While these data contain a wealth of gene-specific information, overall, there was a strong signal for  
309 disruption of cell wall biosynthesis and energy metabolism producing greater fitness costs for *B.*  
310 *pertussis* in the Bvg- phase compared to the Bvg+ phase.

311 **Cell wall biosynthesis.** A number of cell wall biosynthesis genes were identified as essential for  
312 growth in the Bvg- phase but not the Bvg+ phase. Others were not essential in either phase but their  
313 mutation affected the fitness of Bvg- *B. pertussis* more than it did for Bvg+ phase *B. pertussis*. In  
314 particular, genes involved in assembly of new cell wall are represented in this gene set, for example  
315 *ampG*, the entire *mre/mrd* operon, D-alanyl-D-alanyl carboxypeptidase DacC, and a number of  
316 murein endopeptidases or murein transglycosylases (Table 2, Figure 2).

317 **Table 2.** Penicillin binding proteins and murein lytic transglycosylases of *B. pertussis* and their  
 318 essentiality/role in fitness.

Gene	Protein name	Function	Essential/Fitness affected?
BP0102	DacC	D-ala-D-ala carboxypeptidase	Essential Bvg- Fitness Bvg-
BP0377	MrdA (Pbp2)	Cell elongation transpeptidase	Essential Bvg- Fitness Bvg-
BP0905	MrcA (Pbp1A)	DD-transpeptidase	Non-essential
BP1051	DacB	D-ala-D-ala carboxypeptidase	Non-essential
BP1545	DacC2	D-ala-D-ala carboxypeptidase	Non-essential
BP2754	Pbp1C	DD-transpeptidase	Non-essential
BP3028	FtsI	Cell division transpeptidase	Essential Bvg+/-
BP3655	Pbp?	DD-transpeptidase	Non-essential
BP1061	MltE	Murein lytic transglycosylase	Fitness Bvg-
BP3214	MltD	Murein lytic transglycosylase	Fitness Bvg-
BP3268	MltA	Murein lytic transglycosylase	Fitness Bvg-

319

320 AmpG functions to recycle peptidoglycan fragments, released during cell growth, back into the cell  
321 (21). *B. pertussis* releases tracheal cytotoxin during culture. TCT is N-acetylglucosaminyl-1,6-  
322 anhydro-N-acetylmuramyl-(L)-alanyl-γ-(D)-glutamyl-mesodiaminopimelyl-(D)-alanine and is cytotoxic  
323 towards mammalian epithelial cells (22). Its release from *B. pertussis* is ascribed to *B. pertussis*  
324 AmpG being non-, or poorly, functional as expression of *E. coli ampG* in *B. pertussis* greatly reduced  
325 the amount of TCT in culture supernatants (23). TCT was released by both Bvg+ and Bvg- phase *B.*  
326 *pertussis* (24) but the relative amounts released by the two phases was not noted. Thus although *B.*  
327 *pertussis* AmpG may have low levels of activity compared to other bacteria, our data suggest that for  
328 growth of the Bvg- phase, recycling of peptidoglycan fragments is a factor.

329 The *mre* locus contains genes encoding proteins commonly annotated as 'rod shape determining  
330 proteins'. Briefly, these proteins are key components of the cell wall synthesis machinery. Together  
331 they coordinate and participate in the insertion of newly synthesised peptidoglycan fragments into  
332 the peptidoglycan sheath and this is vital for maintenance of cell shape, cell rigidity and cell growth  
333 (25, 26). Viable mutants of these genes have been constructed in other bacteria. Usually they have  
334 altered cell shape, and often they are able to grow only under specific conditions, such as in minimal  
335 media (25). It is striking that none of the genes in this locus are essential for growth in the Bvg+  
336 phase. Transposon insertions were recovered throughout each of these genes in Bvg+ phase  
337 conditions (Figure 2) strongly suggesting that inactivation of each gene gave rise to viable bacteria.

338 DacC is a penicillin-binding, D-alanyl-D-alanyl carboxypeptidase involved in transpeptidation  
339 reactions in peptidoglycan biosynthesis. Most gram negative bacteria encode several/multiple  
340 penicillin binding proteins (PBPs) suggesting redundancy among their functions or that different  
341 PBPs are utilised under different conditions. Analysis of the *B. pertussis* genome sequence identified  
342 eight genes encoding putative PBPs, Table 3. One of these was the cell division transpeptidase FtsI,  
343 and one was MrdA in the *mre/mrd* locus described above. Interestingly, of the other putative DD-  
344 transpeptidases, only that encoded by BP0102 was essential, and then only in the Bvg- phase.

345 Another DacC homologue, encoded by BP1545, was identified. It displays little homology to BP0102  
346 DacC at the amino acid level although it contains strong homology to the DacC conserved domain  
347 (COG1686) with a BlastP score of  $1.49e^{-72}$ . This suggests that BP0102 encoded DacC has a function  
348 that is distinct from those of the other putative DD transpeptidases, and that is essential for viability  
349 in the Bvg- phase.

350 BP1061 (MltE), BP3214 (MltD) and BP3268 (MltA) all encode putative murein lytic transglycosylases.  
351 These enzymes cleave the glycosidic bond between the MurNAc and GlcNAc residues of  
352 peptidoglycan during recycling of cell wall or biosynthesis of new cell wall regions; during cell growth  
353 and division. None were essential but all three were identified as affecting the fitness of Bvg- phase  
354 *B. pertussis* more than Bvg+ phase.

355 To validate findings made using TraDIS, and to further investigate differences in cell wall biosynthesis  
356 between Bvg phases, a defined mutant in *mreB* was constructed. During construction, the mutant  
357 strain was grown only in Bvg+ phase conditions and in doing so viable *mreB* mutants were obtained.  
358 These mutants produced normal looking colonies on charcoal agar after 72 hours incubation at 37°C,  
359 as does WT, confirming that *mreB* is not required for the normal growth rate of *B. pertussis* in the  
360 Bvg+ phase. The viability of the mutant under Bvg- phase conditions was tested. Serial dilutions of  
361 WT and *B. pertussis mreB* grown under Bvg+ phase conditions were plated onto Bvg+ and Bvg- phase  
362 condition charcoal agar plates and the number of viable colonies recovered were compared, Figure  
363 3. Under Bvg+ phase conditions there was no difference between the number of WT and mutant  
364 colonies recovered. However, under Bvg- phase conditions, no colonies of the *mreB* mutant were  
365 recovered, confirming the TraDIS-derived observation that *mreB* is conditionally essential for the  
366 growth of *B. pertussis* on charcoal agar, dependent on Bvg activity. Complementation of the  
367 mutation with a plasmid-borne copy of *mreB* restored growth under Bvg- phase conditions  
368 demonstrating that disruption of *mreB* produced this phenotype. It is possible that the lethality of  
369 the *mreB* mutation on Bvg minus condition plates was due to the high levels of  $MgSO_4$  used to

370 induce Bvg- phase conditions, possibly via effects on osmotic pressure. However, the same lethality  
371 was observed when using nicotinic acid to induce Bvg- phase conditions (data not shown), another  
372 known modulator of Bvg. In addition, attempts to construct the *mreB* mutation in a Bvg mutant  
373 background were unsuccessful, consistent with knock out of *mreB* being lethal to Bvg- phase  
374 bacteria.

375 Together these data strongly suggest that growth of Bvg- phase *B. pertussis* is more sensitive to  
376 disruption of new cell wall synthesis than is the Bvg+ phase. It was reasoned that the higher  
377 sensitivity of Bvg- *B. pertussis* to disruption of cell wall synthesis than Bvg+ *B. pertussis* would result  
378 in them being more sensitive to cell wall-disrupting antibiotics. The MIC for ampicillin was  
379 determined for *B. pertussis* in both phases and was significantly lower for cells growing in the Bvg-  
380 phase, i.e. these bacteria were significantly more susceptible to growth inhibition by ampicillin than  
381 Bvg+ phase bacteria (Figure 4). The sensitivity to gentamicin, an inhibitor of protein synthesis,  
382 although slightly greater for Bvg+ phase bacteria, was not significantly different to that of Bvg- phase  
383 bacteria suggesting that this effect was specific for the cell wall disrupting activity of ampicillin  
384 (Figure 4).

385 Mutation of these genes appears to cause different effects on the Bvg phases of *B. pertussis*. Bvg is a  
386 global regulator of gene expression in *B. pertussis* and thus these differences might be due to  
387 different levels of expression of these genes between the phases. However, these genes are not Bvg-  
388 regulated as determined by RNAseq-mediated analysis of gene expression of Bvg+ and Bvg- phase *B.*  
389 *pertussis* (our unpublished observations and (10)) and each of the genes was expressed in both Bvg  
390 phases. This suggests that the increased sensitivity of the Bvg- phase to mutation of these genes was  
391 not due to lack of expression in the Bvg+ phase, but due to different sensitivity of the phases to  
392 disruption of the activities in which the proteins encoded by these genes are involved.

393 **TCA Cycle/Energy Metabolism.** Succinate dehydrogenase (SDH) participates in both the TCA and  
394 electron transport chain. *B. pertussis* cannot metabolise sugars and preferentially uses glutamate as

395 a carbon source. Thus many biosyntheses in *B. pertussis* stem from metabolism of glutamate via  
396 alpha-ketoglutarate and the TCA cycle. Genes encoding all four subunits of SDH were identified as  
397 being essential for Bvg- *B. pertussis*, as perhaps would be expected for such a key metabolic enzyme  
398 complex. Surprisingly, SDH was not essential for growth of Bvg+ *B. pertussis* although the insertion  
399 indices for *sdhA-D* in Bvg+ *B. pertussis* were low. It is not clear how glutamate might be metabolised  
400 in the absence of SDH, as metabolism of the  $\alpha$ -ketoglutarate through which glutamate derived  
401 carbon enters the TCA requires SDH. In addition, SDH generates FADH<sub>2</sub> that transfers electrons to  
402 the electron transport chain for generation of the proton motive force and ATP production.

403 A difference between phases in the effect of mutation of the cytochrome bc complex (complex III)  
404 was observed. The insertion indices for *petABC* were significantly lower for Bvg- *B. pertussis* (0.078,  
405 0.053, 0.064) compared to Bvg+ phase (0.183, 0.165, 0.162),  $p=3.88E-09$ ,  $3.43E-14$ ,  $1.36E-06$   
406 respectively (Figure 5). The cytochrome bc complex is a key link between electron transport and  
407 proton translocation for utilising metabolically derived energy to generate the proton motive force.  
408 Most bacteria contain alternative pathways for quinol oxidation that bypass the cytochrome bc  
409 complex but many of these do not have the same energy transduction, and thus the same proton  
410 translocation power, as the cytochrome bc complex. The relative lack of sensitivity of Bvg+ *B.*  
411 *pertussis* to mutation of *petABC* suggests that growth in the Bvg+ phase is less reliant on this  
412 electron transport complex than is growth in the Bvg- phase. These findings suggest that energy  
413 metabolism in *B. pertussis* differs between the Bvg phases.

414 There were an equal number of genes whose insertion index was significantly lower under Bvg+  
415 phase conditions compared to Bvg- conditions. Interestingly, a different set of cell wall  
416 biogenesis/cell division genes than those with lower insertion indices under Bvg- conditions were in  
417 this group, again suggesting that there are differences in these processes between Bvg phases. Of  
418 note, also within this group were a number of genes involved in synthesis and export of the  
419 *Bordetella* extracellular polysaccharide (EPS), often referred to as the *Bordetella* capsule. Although

420 relatively poorly characterized, the genes in this locus are maximally expressed under Bvg<sup>-</sup> phase  
421 conditions, although basal levels of transcription are observed under Bvg<sup>+</sup> phase conditions (9).  
422 Recently, upregulation of the capsule locus was observed in *B. pertussis* during infection of mice,  
423 conditions under which *B. pertussis* had been assumed to be in the Bvg<sup>+</sup> phase (27). The same study  
424 identified an interaction between components of the EPS export apparatus and BvgS, and mutation  
425 of EPS export genes caused wide ranging effects on the expression of a number of Bvg-regulated  
426 genes. While interesting, it is unclear why mutation of the capsule locus causes fitness costs in the  
427 Bvg<sup>+</sup> phase *in vitro*, conditions under which the locus is expressed at only low levels, whereas there  
428 is no apparent defect under Bvg<sup>-</sup> phase conditions in which there is high level expression of the  
429 locus.

## 430 **Discussion.**

431 Previously, the genes essential for growth of *B. pertussis* *in vitro*, on Bordet-Gengou agar and those  
432 conditionally essential for survival of *B. pertussis* *in vivo* in a mouse model of infection were defined  
433 (12). Here, we focused on the effect of the global regulator of *B. pertussis* virulence, the Bvg two  
434 component system, and identified genes that were conditionally essential for *in vitro* growth of *B.*  
435 *pertussis*, dependent on the activity of Bvg. Bvg is hypothesized to sense stimuli that signal to  
436 *Bordetella* that they have entered a host, as switching from ambient temperature to 37°C switches  
437 them from the Bvg<sup>-</sup> to the Bvg<sup>+</sup> phase, that is characterised by high level expression of many of the  
438 proteins (e.g. adhesins, toxins) involved in infection. In this way, Bvg has been regarded as a switch  
439 that regulates the transition between virulent and avirulent states. Several studies have identified  
440 that Bvg regulates the expression of a large number of genes (for example, 550 genes (10)) including  
441 those maximally expressed in either the Bvg<sup>+</sup> or Bvg<sup>-</sup> phase. A spectrum of states exists between  
442 these two extremes, with some genes maximally expressed in intermediate states. The role of Bvg in  
443 regulation of virulence/infection in the *Bordetellae* has been well studied. Much less is known  
444 regarding Bvg-regulation of basic physiology. Bvg-regulation of growth rate has been observed. For

445 example, during laboratory culture, compared to Bvg+ phase *B. pertussis*, Bvg- phase *B. pertussis* exit  
446 lag phase earlier, grow at a faster rate during exponential growth and reach higher biomass final  
447 yields (11), but the genetic basis for this is unknown.

448

449 We identified 396 genes that were unconditionally essential for growth of strain BP536, compared to  
450 609 genes defined as essential for strain UT-25 ((12)). As expected, many of the genes identified  
451 here were also essential for UT-25, TableS1. Thirty seven genes identified as essential for BP536  
452 were not essential for UT-25 whereas 134 genes essential for UT-25 were dispensable for growth of  
453 BP536. The two studies used different laboratory agars for growth (charcoal vs Bordet Gengou) but  
454 both are similar, utilising peptone as the major carbon source for growth, and a source of starch (not  
455 metabolised by *B. pertussis*) to complex small, hydrophobic inhibitory molecules. Here, Tn5 was used  
456 whereas Gonyar et al used a Mariner-based transposon. However, in both cases high-density  
457 transposon libraries were recovered, suggesting saturation of the genome. Thus, these differences in  
458 gene essentiality may represent differences between the two strains in their requirements for  
459 growth. Of particular note, two loci were identified as essential in UT-25 but not BP536, comprising  
460 20 of the 23 genes between BP0911-BP0934 and 17 of the 19 genes between BP1158-BP1176. The  
461 functions of genes in these regions are unknown, but many appear to encode transport and  
462 metabolic functions. Fascinatingly, the genes BP0910-0934 represent one of the known regions of  
463 difference between strains in that this region is absent from many *B. pertussis* strains (28, 29). Thus,  
464 it is surprising that it is essential for strain UT-25, when this region is clearly dispensable to many  
465 other strains. No loci of contiguous genes were identified as essential in BP536 but not UT-25, and it  
466 is possible that differences in the sensitivity of approaches used to identify essential genes  
467 contributes to some of these differences.

468 Here, we identified some surprising differences in the effect of mutations in genes involved in  
469 certain core processes, dependent on the activity of Bvg. Seventy nine genes were identified as



470 being essential only in the Bvg<sup>-</sup> phase, not the Bvg<sup>+</sup> phase. Analysis of the predicted functions of  
471 these genes identified cell wall biogenesis, cofactor biosynthesis and translation as being processes  
472 for which more genes were essential in the Bvg<sup>-</sup> phase compared to Bvg<sup>+</sup>.

473 Identification of genes whose mutation is not lethal, but appears to affect the fitness of the  
474 bacterium, expands this observation. Of the 51 genes whose mutation significantly affect the fitness  
475 of Bvg<sup>-</sup> phase bacteria but not Bvg<sup>+</sup> phase, 35 encode functions involved in these same three  
476 processes, compared to only 14 genes for the Bvg<sup>+</sup> phase. Among these genes are those considered  
477 central to cell wall biogenesis, for example the *mreBCDmrdAB* operon that encodes proteins which  
478 coordinate the insertion of newly synthesized peptidoglycan fragments into the existing cell wall. In  
479 some other bacteria, strains carrying knock out mutations of these genes are viable, but only under  
480 certain conditions such as nutrient limitation. For example, *E. coli mreB* mutants are spheroid rather  
481 than the WT rod shape (25). Interestingly, for *B. subtilis mre* mutants, the growth and shape defects  
482 usually associated with mutations in these genes were completely suppressed by supplementing the  
483 growth media with magnesium, presumably through stabilization of the cell wall (30). In *B. pertussis*,  
484 mutations in this locus appear remarkably well tolerated under Bvg<sup>+</sup> phase conditions, as reasonable  
485 numbers of insertions in these genes were counted in the mutant library, and insertions were  
486 recovered throughout each gene of the locus. The mutants were recovered from plates at a single  
487 time point, 72 hours after plating, which would be considered normal growth for *B. pertussis* and  
488 suggests that these mutants grew at normal rates. We constructed a defined knock out mutation of  
489 *mreB* in *B. pertussis*. Full characterisation of the phenotypes of this mutant, and of other *mre/mrd*  
490 mutants, is on-going but it confirmed that *mreB* was not required for the wild type growth rate of  
491 Bvg<sup>+</sup> phase *B. pertussis* whereas this mutant was non-viable under Bvg<sup>-</sup> phase conditions despite the  
492 Bvg<sup>-</sup> phase being induced by the addition of Mg<sup>2+</sup> to the medium.

493 A number of studies have identified the essentiality of genes as being conditional through changing  
494 growth conditions, often using different media compositions or growth in the presence of different

495 stressors. Here, is demonstrated the conditional essentiality of genes involved in core processes  
496 dependent on the activity of a regulator of gene expression. It is recognised that Bvg state was  
497 manipulated by the addition of MgSO<sub>4</sub> to the agar but this was the only difference in the growth  
498 conditions between the Bvg phases.

499 It might be expected that Bvg would thus influence the essentiality of genes, as strong down-  
500 regulation of genes (switching off expression) in one Bvg phase would probably render them non-  
501 essential in that phase. Thus, Bvg-dependent essentiality might be expected to mirror Bvg-regulation  
502 patterns. However, surprisingly, this was not the case. None of the conditionally essential genes, or  
503 those for which mutation was identified as affecting the fitness of one Bvg phase, were identified as  
504 Bvg-regulated (>2 fold change in expression between phases) in an RNAseq-mediated study of gene  
505 expression in Bvg+ and Bvg- phase growth (our own unpublished observations and (10)). Thus,  
506 conditional essentiality is not due to the differential expression of these genes between Bvg phases  
507 but implies fundamental differences in the contribution of specific genes to *B. pertussis* physiology,  
508 dependent on Bvg.

509 Here, we identify that the process of cell wall biosynthesis can proceed differently between Bvg  
510 phases, as Bvg+ phase growth can tolerate the loss of a range of key cell wall biosynthesis genes. We  
511 have identified a number of specific genes involved in this difference, providing targets for further  
512 study of the phenomenon. Our findings suggest that Bvg not only regulates virulence in *B. pertussis*,  
513 but that it coordinates much wider aspects of *B. pertussis* physiology and understanding the  
514 coordination between growth physiology and virulence will be important to fully dissecting *B.*  
515 *pertussis*-host interactions. In addition, *B. pertussis* is able to maintain normal growth rates in the  
516 absence of seemingly key cell wall biosynthesis genes, and is able to do this without mutation, or  
517 acquisition of novel genes. We suggest that understanding mechanisms by which bacteria may  
518 withstand disruption of cell wall biosynthesis, a key target of important antibiotics, is important to

519 understanding mechanisms of antibiotic resistance and for developing effective new antimicrobial  
520 drugs.

521 **Author Statements.**

522 TB: investigation, formal analysis, writing – original draft; IM: investigation, formal analysis; JDK:  
523 investigation, formal analysis; GCL: formal analysis, data curation, software; MM: formal analysis,  
524 data curation, software; JP: resources, supervision, writing-review and editing; AP:  
525 conceptualisation, formal analysis, funding acquisition , project administration, resources,  
526 supervision, writing-review and editing.

527

528 **Conflicts of Interest.**

529 There are no conflicts of interest for any of the authors.

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534

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- 609

610 Figure Captions.

611 Figure 1

612 Gamma fitted plots of the frequency distribution of the insertion indices for Bvg+ and Bvg- phase  
613 transposon mutant libraries. The bimodal distribution highlights essential genes (left peaks) and non-  
614 essential genes (right peaks). The cut off values of insertion indices for designating genes as essential  
615 or whether their essentiality is ambiguous are shown.

616 Figure 2

617 Plots of insertion sites and frequencies in the genes discussed in the text, for Bvg+ and Bvg- phase  
618 conditions. The height of the lines indicates the frequency of insertions at that site. Green and red  
619 distinguish the orientation of the transposon insertion into the chromosome. Plots were generated  
620 from mapped read files and visualized in Artemis as described (3).

621 Figure 3.

622 *B. pertussis mreB* is unable to grow under Bvg- phase conditions. Strains were grown under on plates  
623 Bvg+ phase conditions. Bacteria were resuspended in PBS, serially diluted and dilutions plated onto  
624 agar and grown under either Bvg+ and Bvg- phase conditions for 72 hours until colonies were clearly  
625 visible. All strains grew very similarly under Bvg+ and Bvg- phase conditions except for the *B.*  
626 *pertussis mreB* mutant (BpmreB) that did not grow under Bvg- phase conditions. BpmreBpBBR: the  
627 mreB mutant carrying the shuttle vector pBBR1MCS. BpmreBpBBRmreB: the *mreB* mutation  
628 complemented by a copy of *mreB* carried on the shuttle vector pBBR1MCS.

629 Figure 4

630 The MIC for ampicillin and gentamicin of *B. pertussis* grown under Bvg+ and Bvg- phase conditions.  
631 The graphs show the average and standard error of triplicate samples. Significance was calculated  
632 using a T-test. \* p<0.05.

633 Figure 5

634 Plots of insertion sites in the *petABC* locus for Bvg+ and Bvg- phase conditions. The height of the  
635 lines indicates the frequency of insertions at that site. Green and red distinguish the orientation of  
636 the transposon insertion into the chromosome. Plots were generated from mapped read files and  
637 visualized in Artemis as described (3).

638 S1 Table. *B. pertussis* essential genes, unconditional and Bvg-dependent conditionally essential  
639 genes.

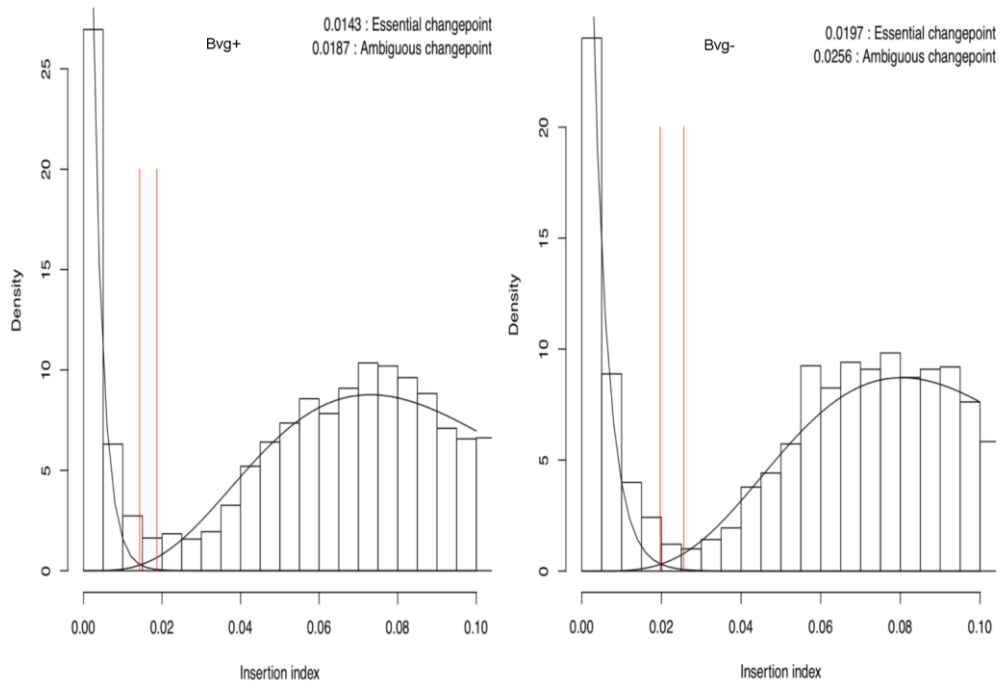
640 S2 Table. Ambiguously essential genes, Bvg+ and Bvg- phases.

641 S3 Table. Fitness-affected genes, Bvg+ and Bvg- phases.

642

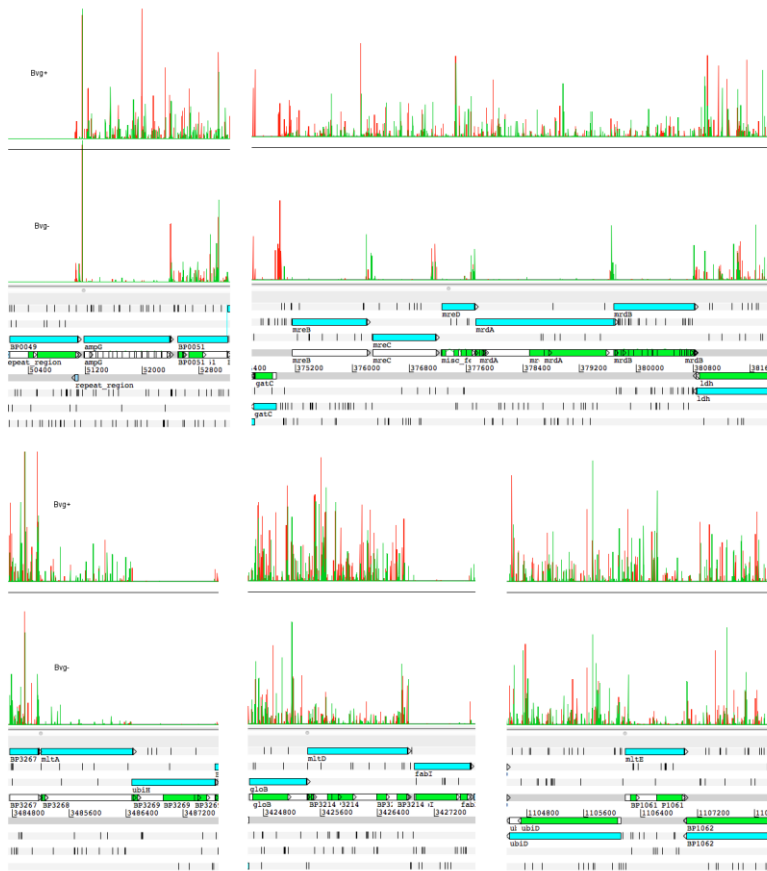


643 Figure 1.



644

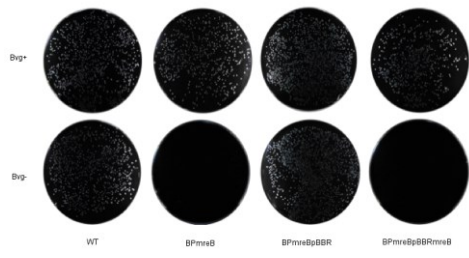
645 Figure 2.



646

647

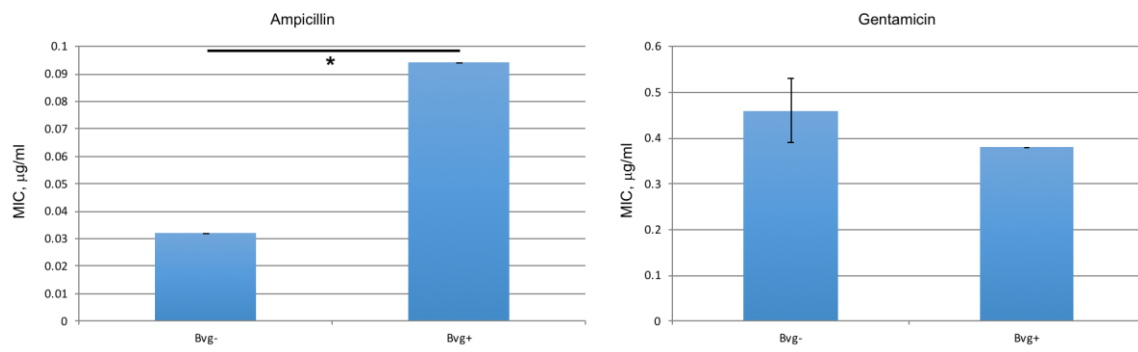
648 Figure 3.



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651 Figure 4.



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